

FORMAL METHODS IN CELL BIOLOGY

Jasmin Fisher

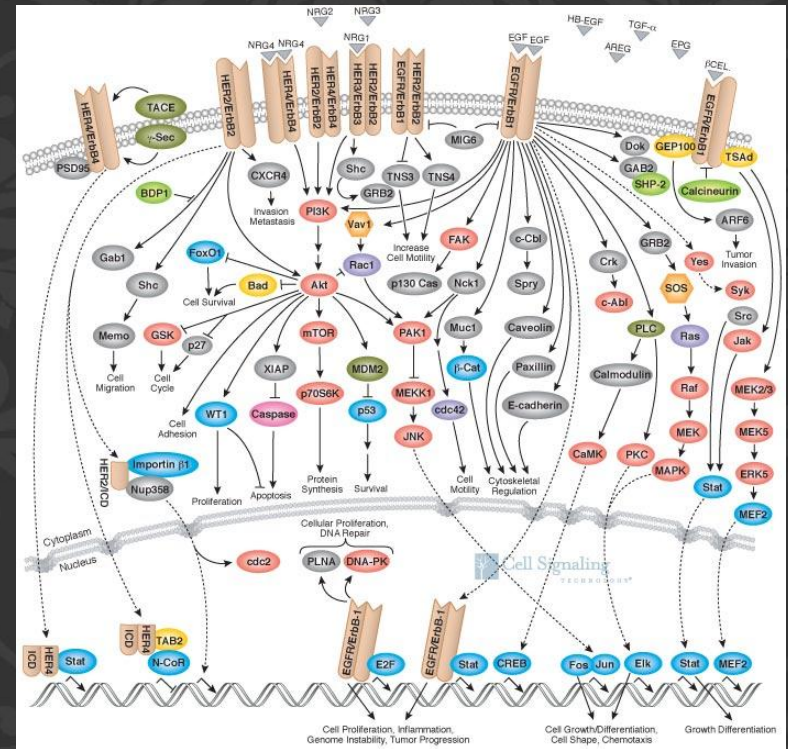
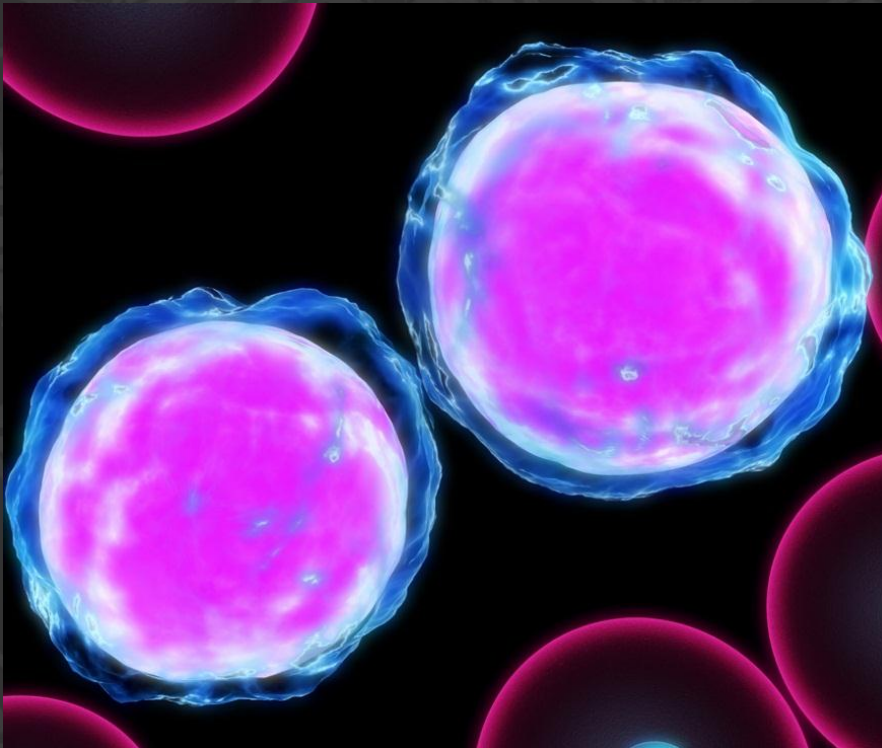
Microsoft Research Cambridge

Tutorial, FMCAD 2012

Cambridge, UK

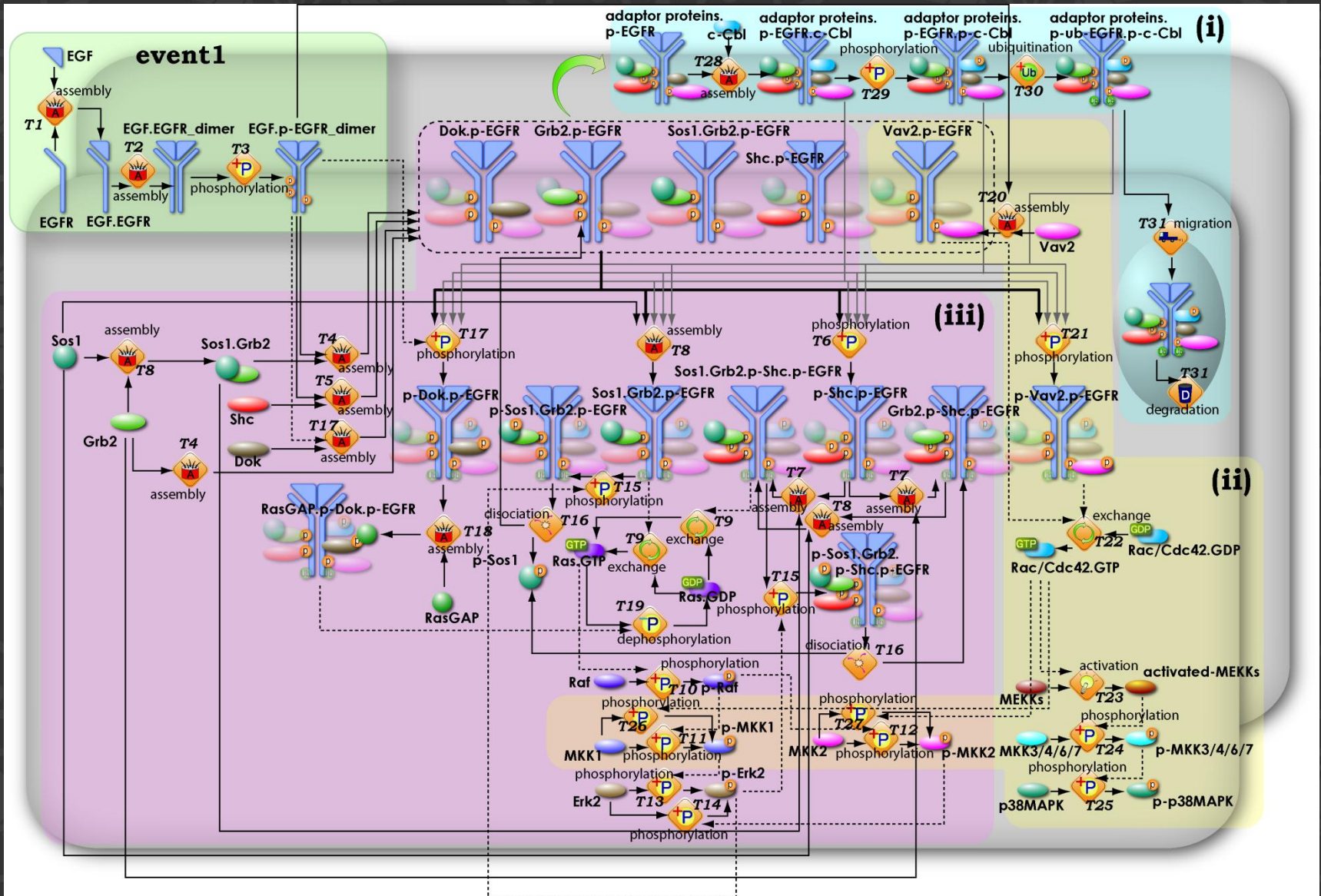
Executable Biology

Tackling the problem of complexity



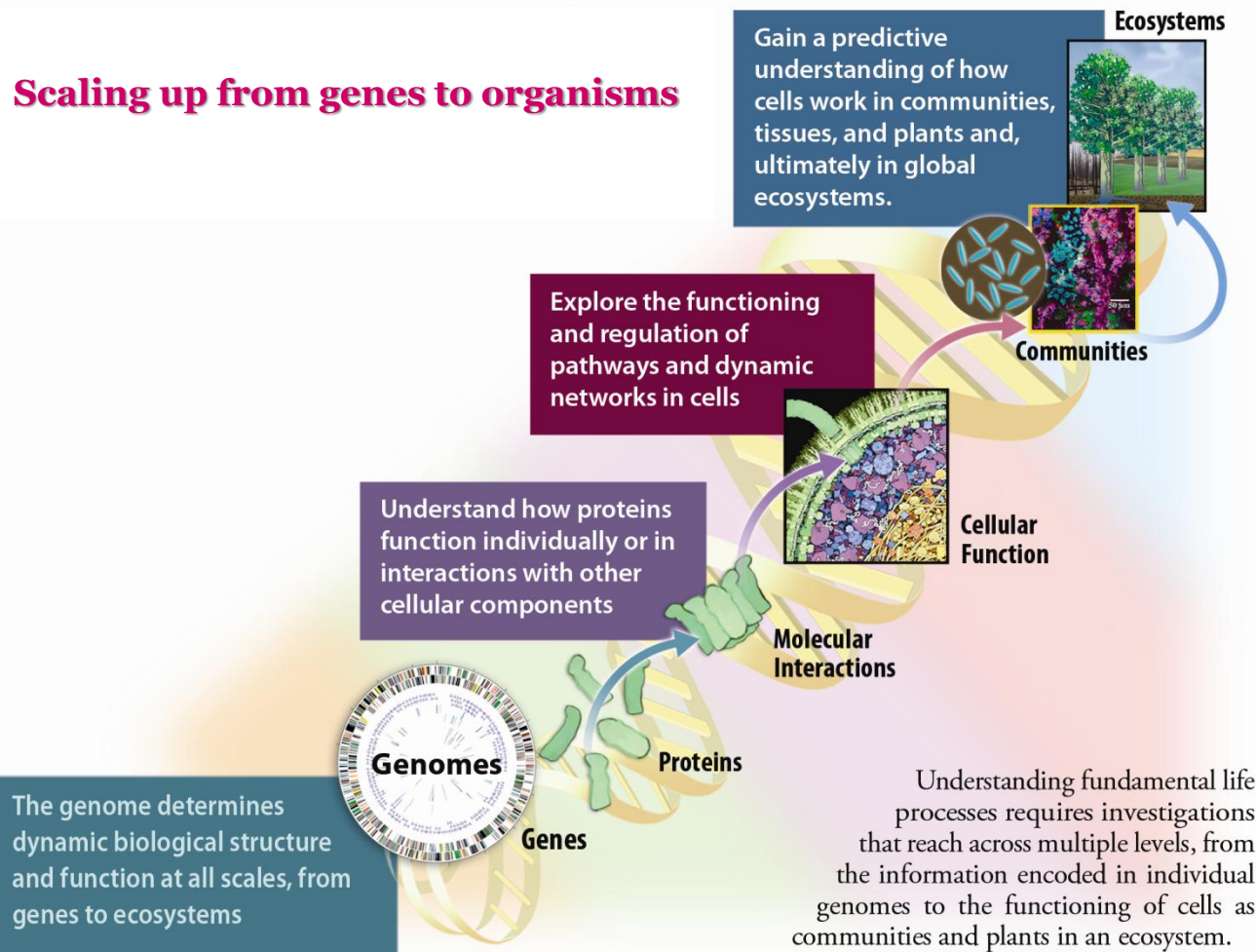
Biocomplexity

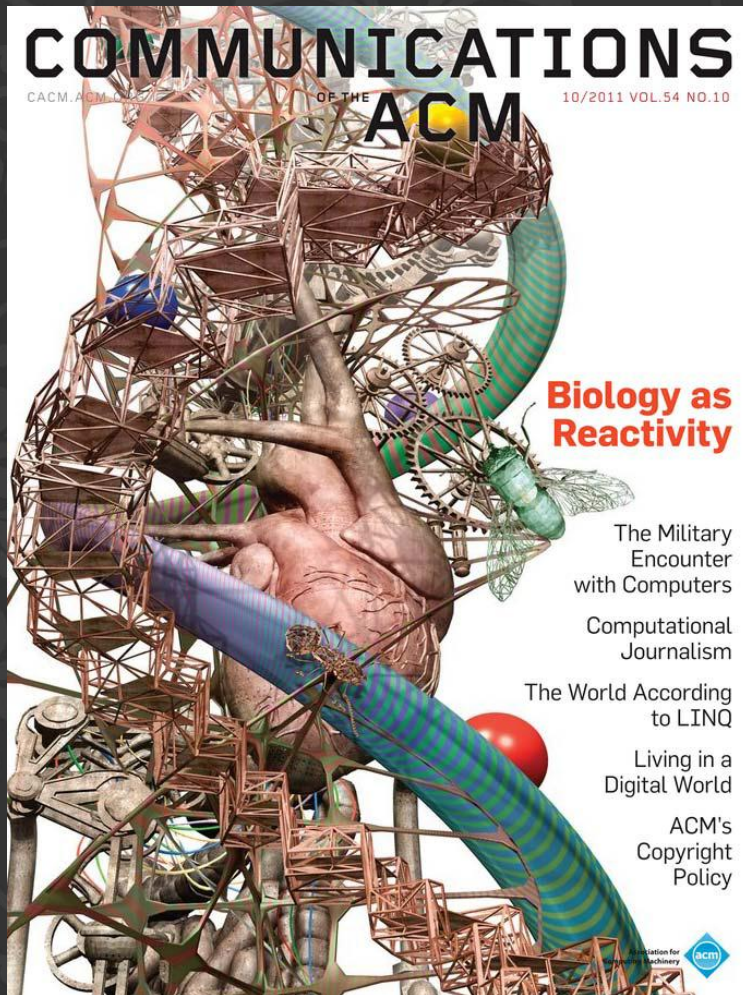
e.g. EGFR signalling pathway



Scaling up...

Scaling up from genes to organisms





Fisher, Harel & Henzinger CACM (2011)

Systems Biology

- System level understanding of living systems.
- Establish the methodologies and techniques to understand biological complexity.

From data collection to data analysis...

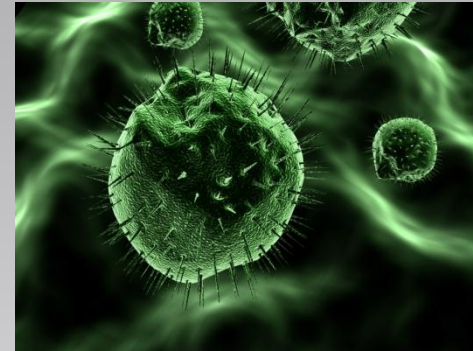


- Data mining
- Classical bioinformatics (omics...)
- Synthetic Biology
- **Executable Biology**

Engineering



Reverse - Engineering

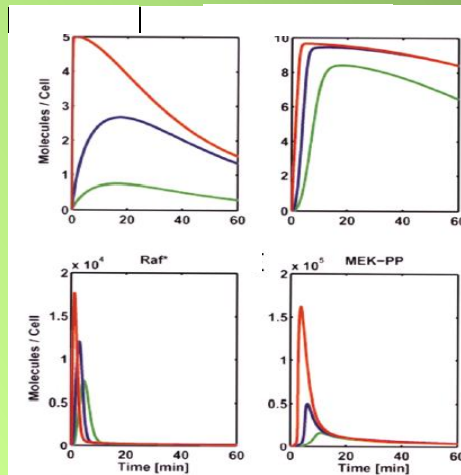


Use of formal methods to model biological systems

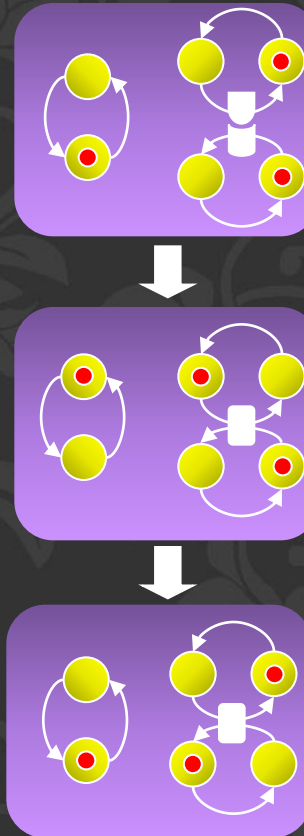
A tale of two cultures...

A Mathematical models

$$\begin{aligned}\frac{d[c_1]}{dt} &= 3.84 \times 10^{-3} \times c_3 - 3.00 \times 10^7 \times c_1 \times c_2 \\ \frac{d[c_2]}{dt} &= 3.84 \times 10^{-3} \times c_3 + 5.00 \times 10^{-4} \times c_8 + \\ &\quad - 3.00 \times 10^7 \times c_3 - 5.00 \times 10^{-3} \times c_8 \\ \frac{d[c_3]}{dt} &= 3.00 \times 10^7 \times c_1 \times c_2 - 3.84 \times 10^{-3} \times c_3 \\ \frac{d[c_8]}{dt} &= 1.73 \times 10^{-7} \times c_5 \times c_6 - 0.2 \times c_8 \\ \frac{d[c_5]}{dt} &= 1.46 \times 10^{-2} \times c_3 - 1.66 \times 10^{-6} \times c_5\end{aligned}$$



B Computational models



Mathematical vs. Computational Models

Mathematical

Denotational semantics

Equations describing the relations between quantities and change over time.

Equations do not prescribe an algorithm for solving them.

Computational

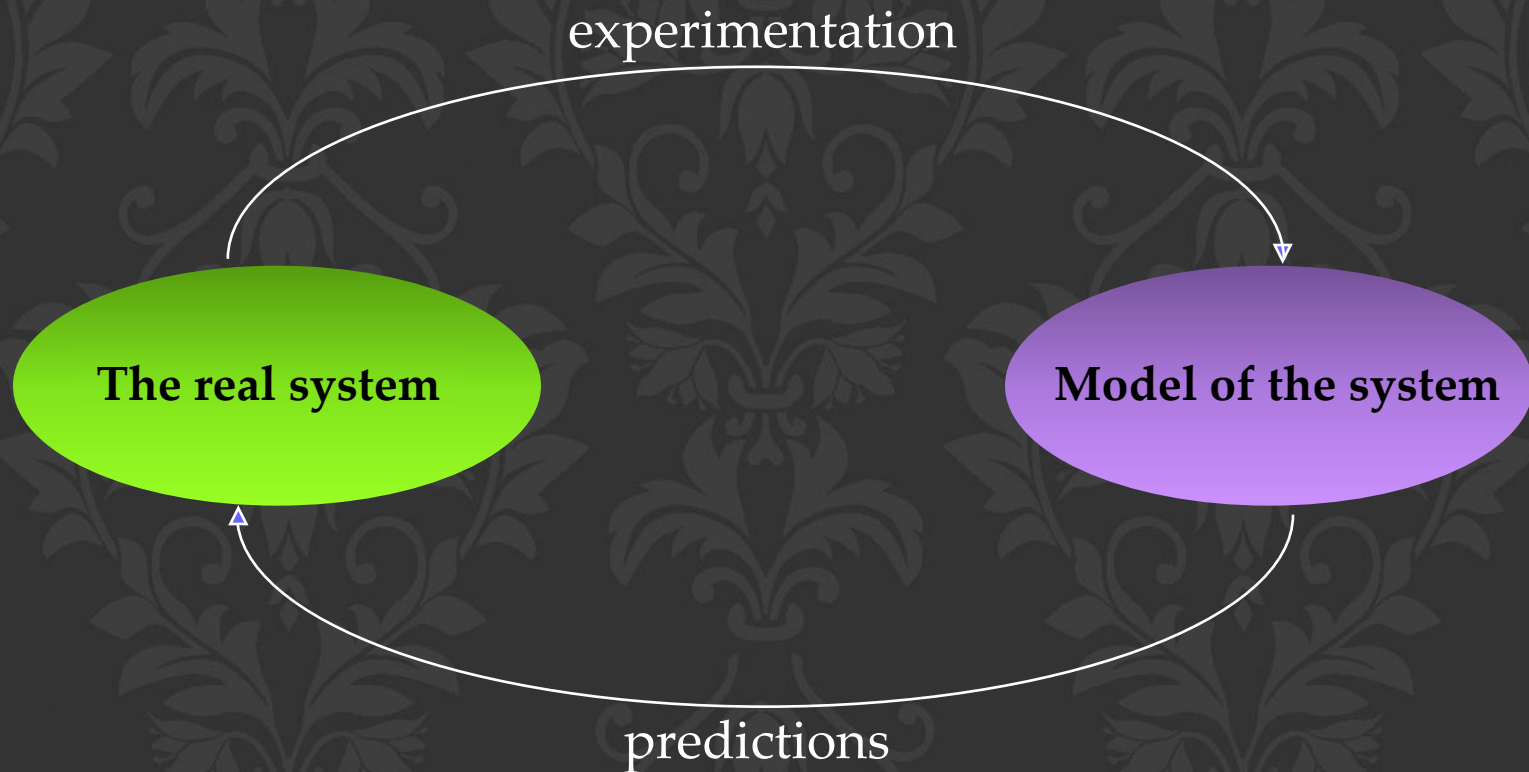
Operational semantics

Sequence of steps to be taken by abstract machine.

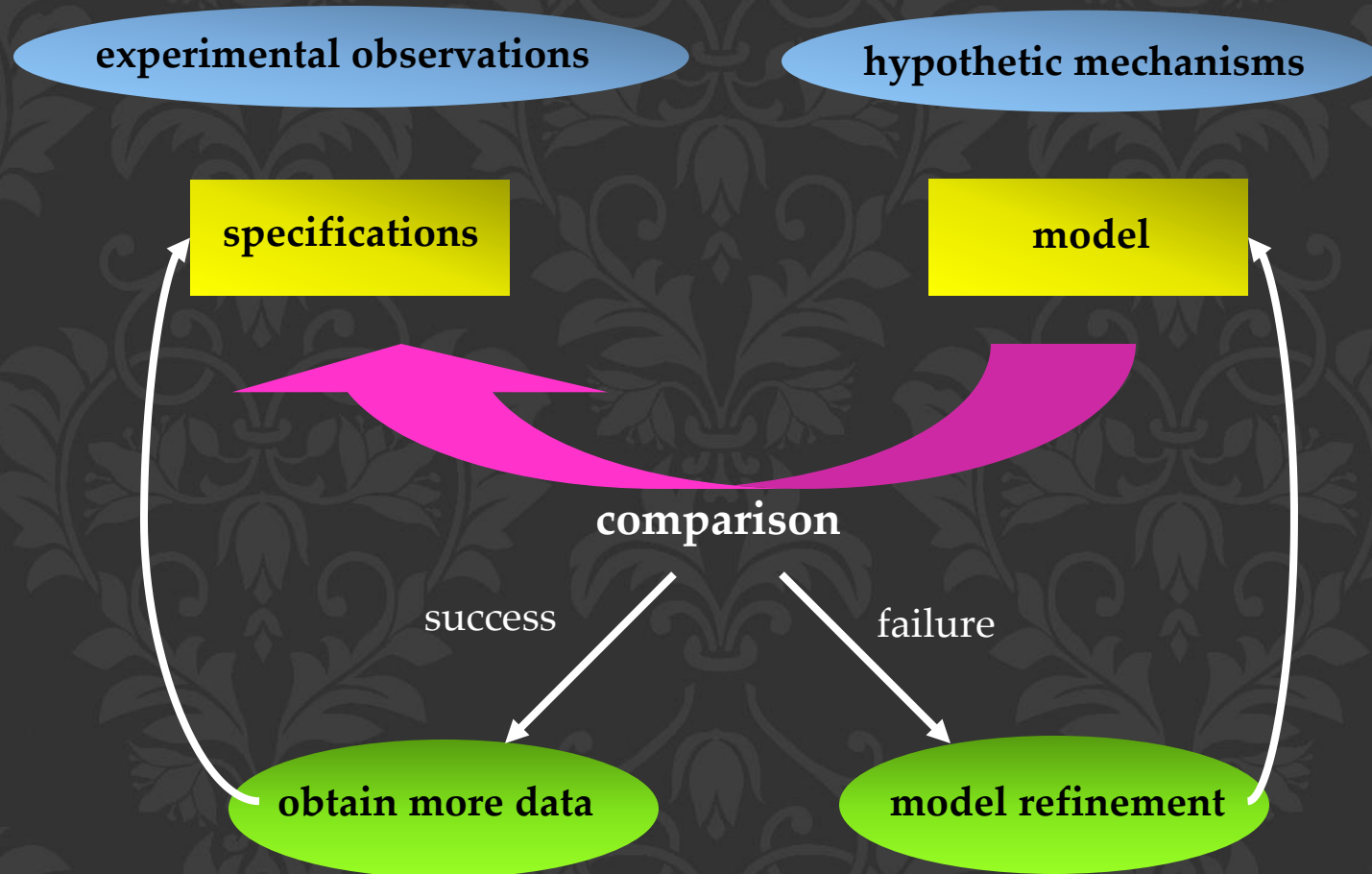
Provide a recipe for execution on a computer.

Fisher J. & Henzinger T.A. *Nature Biotechnology* (2007) 25(11):1239-1249

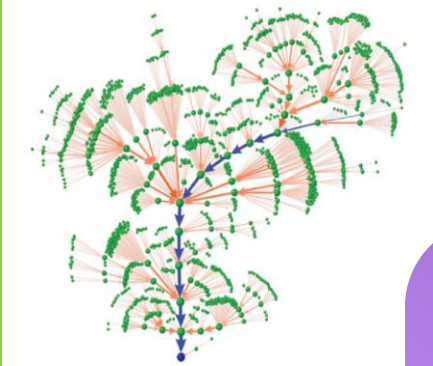
Scientific Method



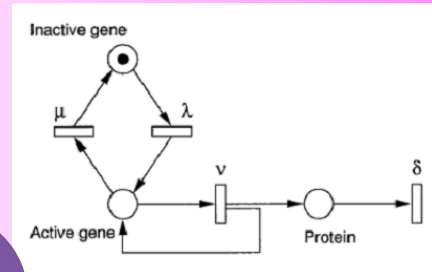
Bio-CEGAR



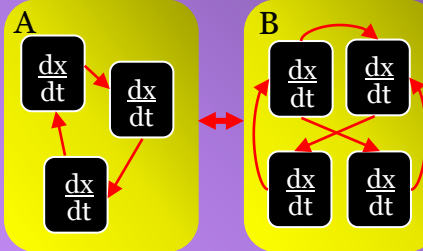
Models for Executable Biology



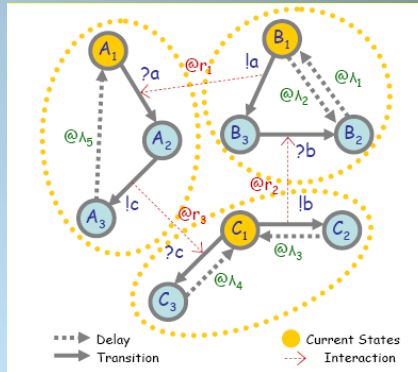
Qualitative networks



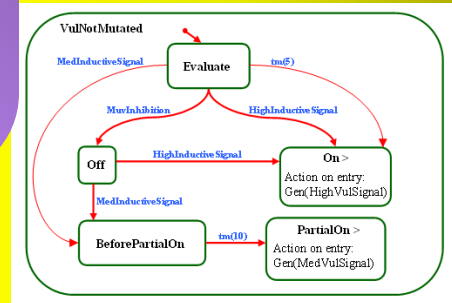
Petri nets



Hybrid models

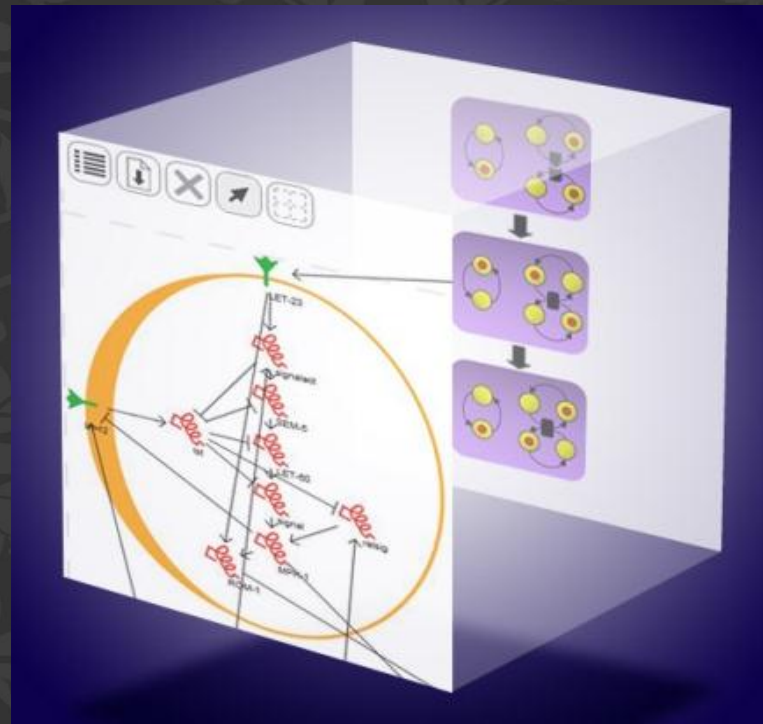


Pi calculus



State-machines

Modelling biological regulatory networks using Qualitative Networks



Qualitative Networks

- Extension of Boolean networks
- Larger range of possible values
- More flexible target functions
- Modelling of hierarchy
- Increased sophistication in analysis
- **Bio Model Analyzer**

Schaub *et al.*, *BMC Systems Biology* (2007)

Schaub *et al.*, *RECOMB Systems Biology LNBI* (2008)

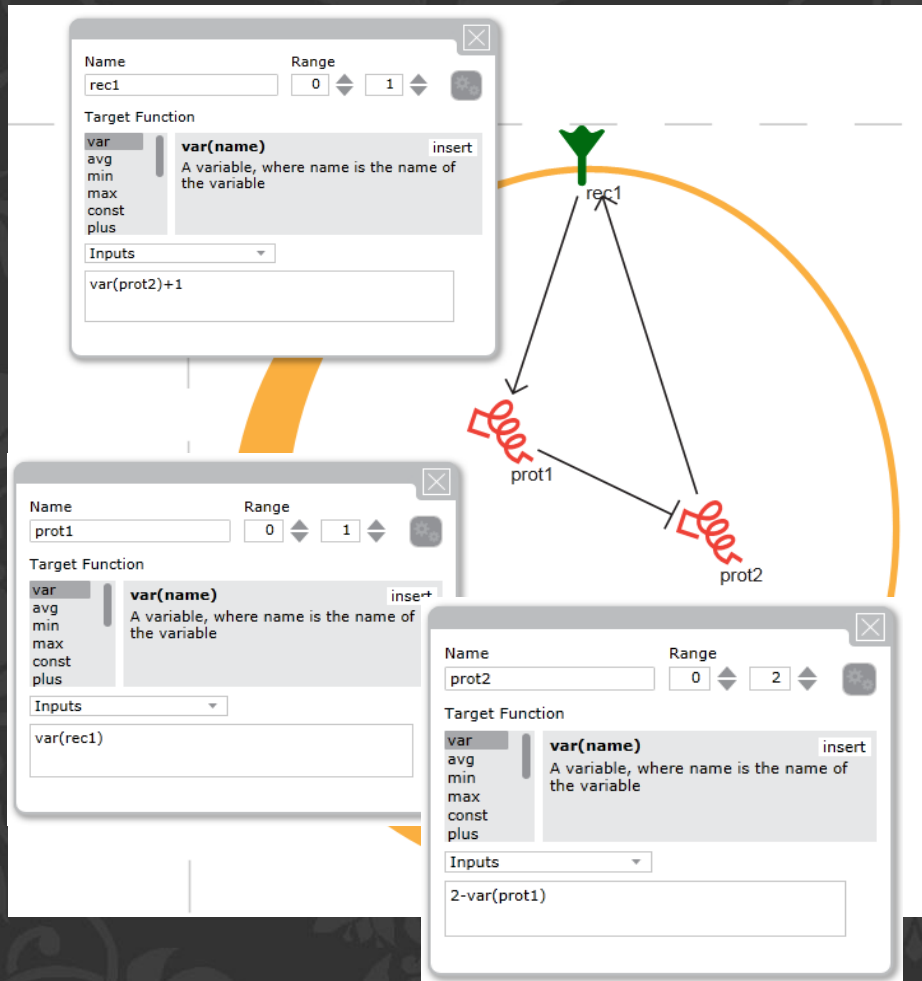
Cook *et al.*, *VMCAI* (2011)

Benque *et al.*, *CAV* (2012)

...more formally:

A *qualitative network* (QN) is $Q = (V, T, N)$, where $V = (v_1, v_2, \dots, v_n)$ is a set of variables ranging over $\{0, 1, \dots, N\}$ and $T = (T_1, \dots, T_n)$ are their respective target functions.

A state of the system is an assignment $s: V \rightarrow \{0, 1, \dots, N\}$. Let Σ denote the set of all possible states.



...more formally:

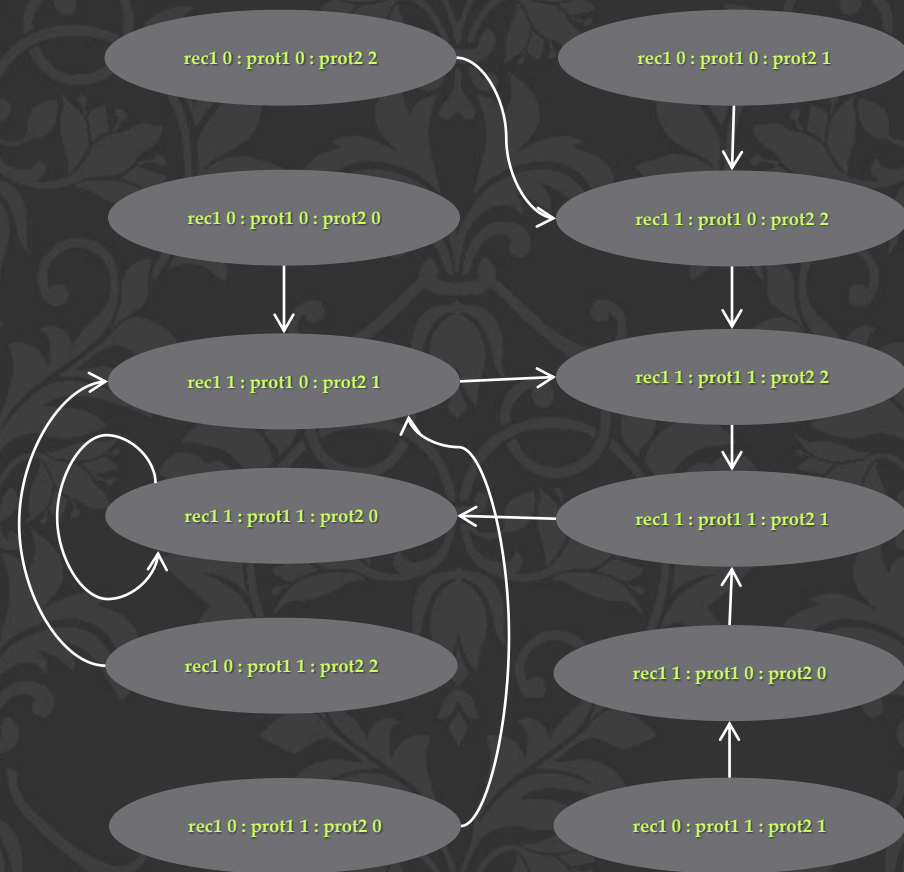
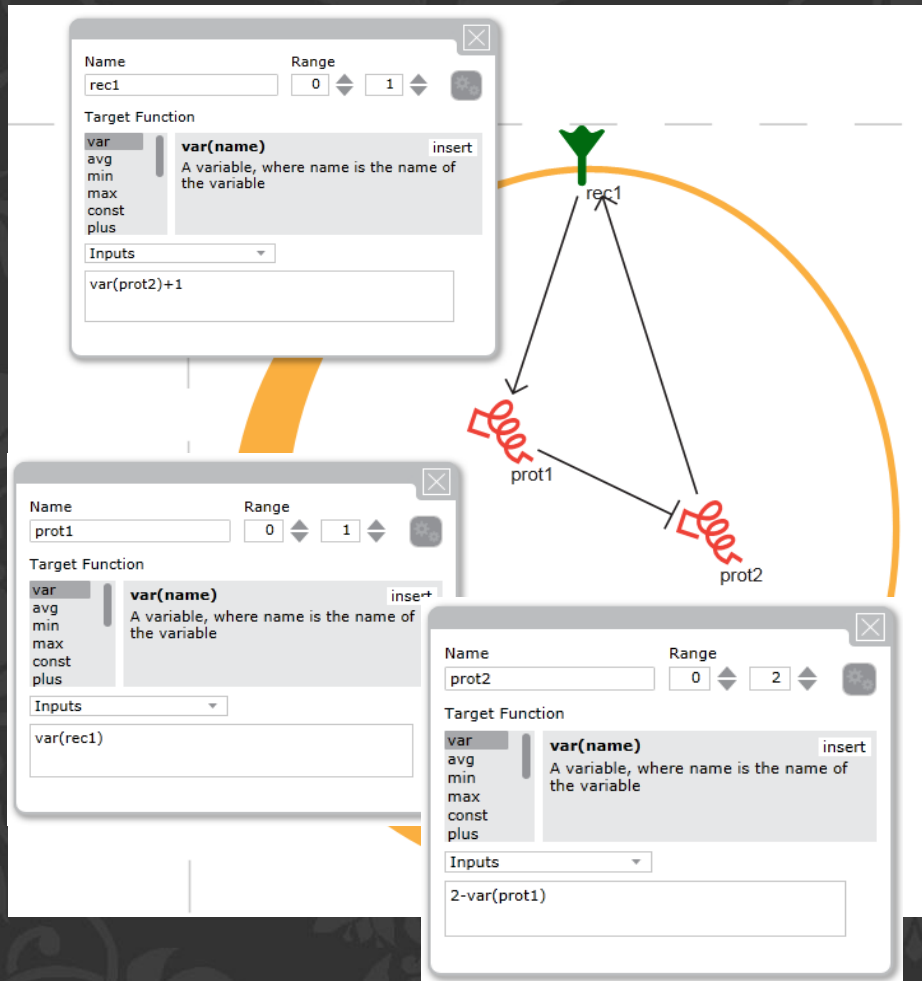
A **qualitative network** (QN) is $Q = (V, T, N)$, where $V = (v_1, v_2, \dots, v_n)$ is a set of variables ranging over $\{0, 1, \dots, N\}$ and $T = (T_1, \dots, T_n)$ are their respective target functions.

A state of the system is an assignment $s: V \rightarrow \{0, 1, \dots, N\}$. Let Σ denote the set of all possible states.

A **target function** $T_i \in T$ is $T_i: \Sigma \rightarrow \{0, 1, \dots, N\}$. Intuitively, in a given state s , variable v_i “would like” to get the value $T_i(s)$. However, values of variables change by at most 1. The *successor* of state s is s' , where for

every $v_i \in V$ we have:
$$s'_i(v_i) = \begin{cases} s(v_i) + 1 & \text{If } s(v_i) < T_i(s) \text{ and } s(v_i) < N \\ s(v_i) - 1 & \text{If } (v_i) > T_i(s) \text{ and } s(v_i) > 0 \\ s(v_i) & \text{Otherwise} \end{cases}$$

Scalable analysis is possible through combination of static analysis and verification techniques.



Executable modelling of cancer signalling in mammalian skin



Freddy Radtke
EPFL



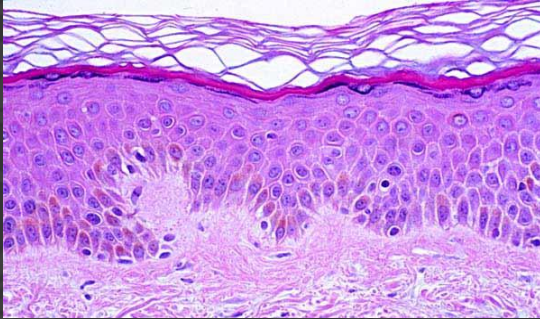
Marc Schaub
Stanford

Mammalian skin (epidermis)

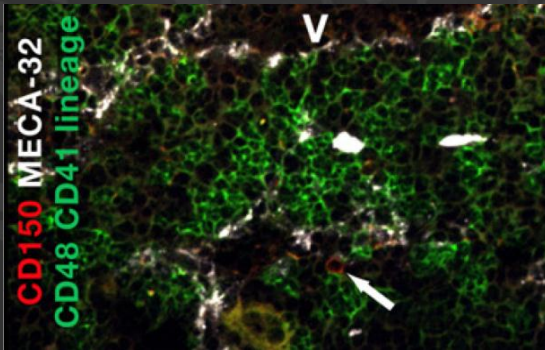


instability

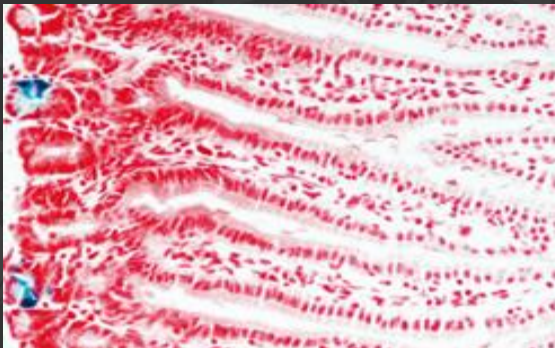
Tissue homeostasis is *critical*



Cells lost = new cells made

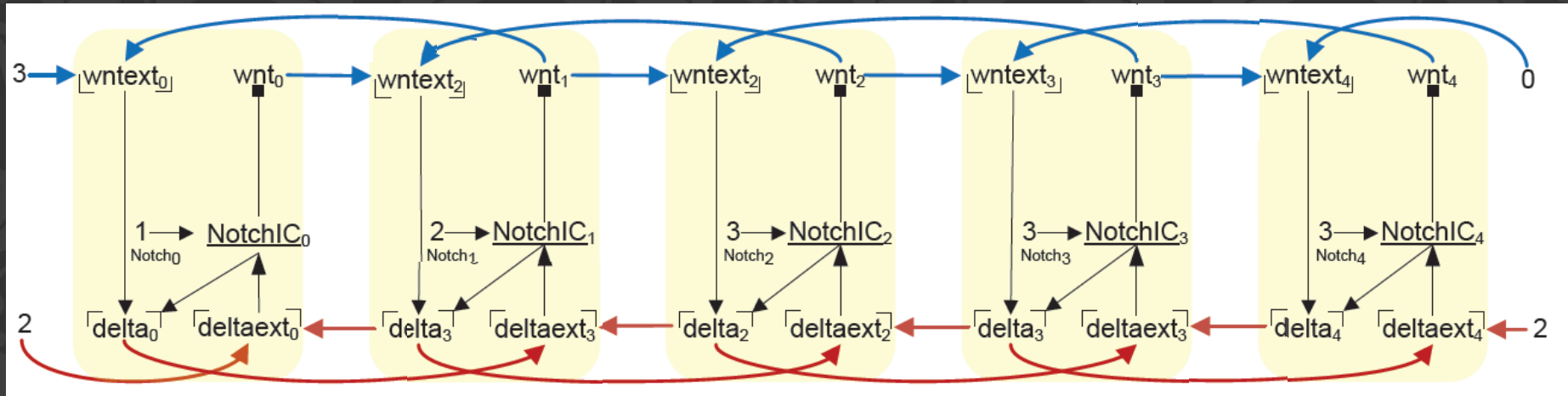


Cells lost > new cells made **FAILURE**



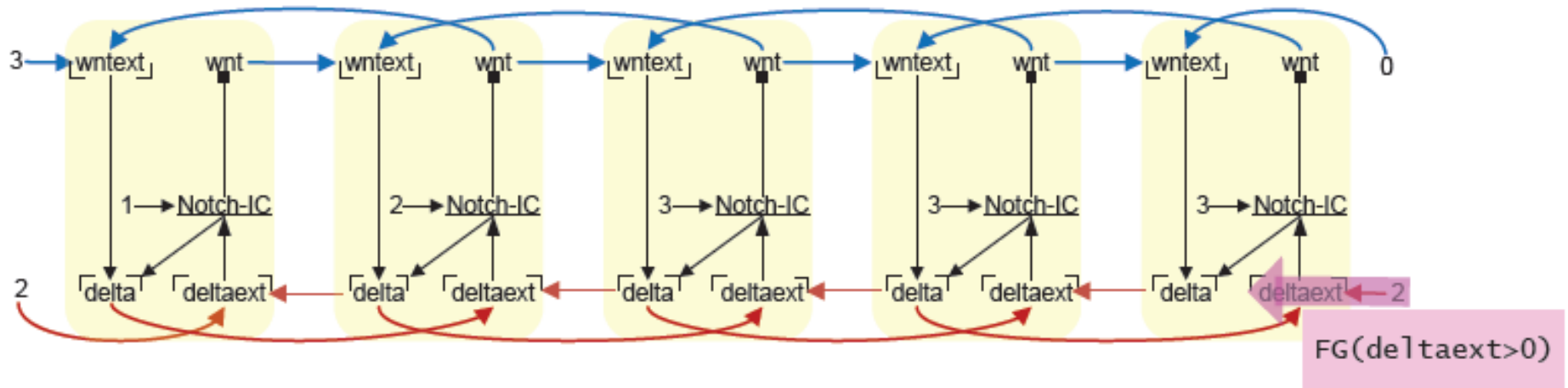
New cells made > cells lost **CANCER**

Mammalian skin model

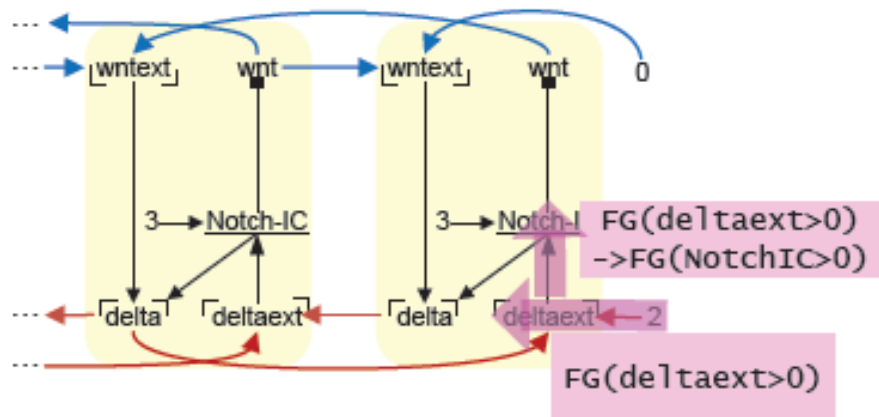


Does there exist a unique fix point that is eventually reached?

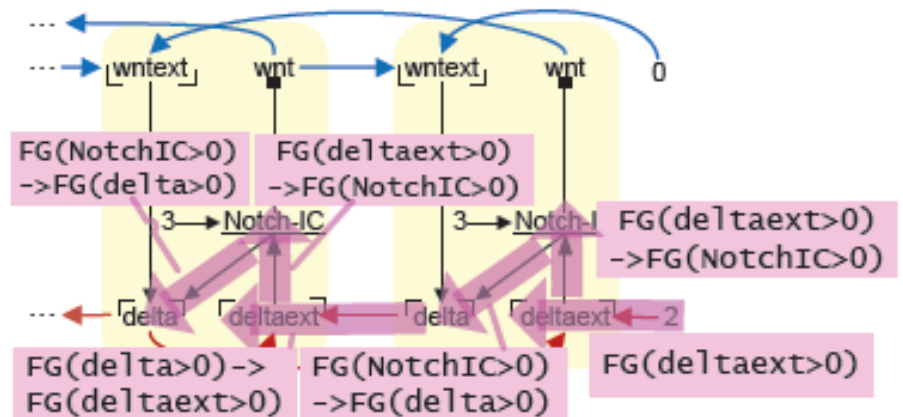
Geographic Proof



(a)

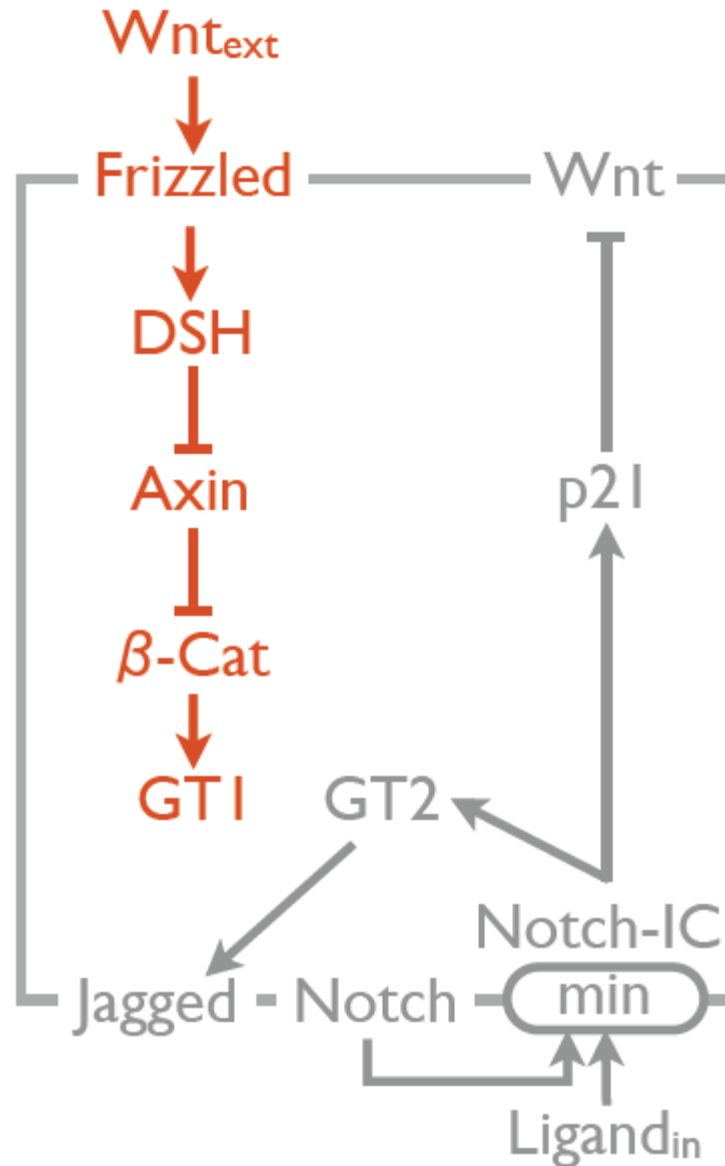


(b)

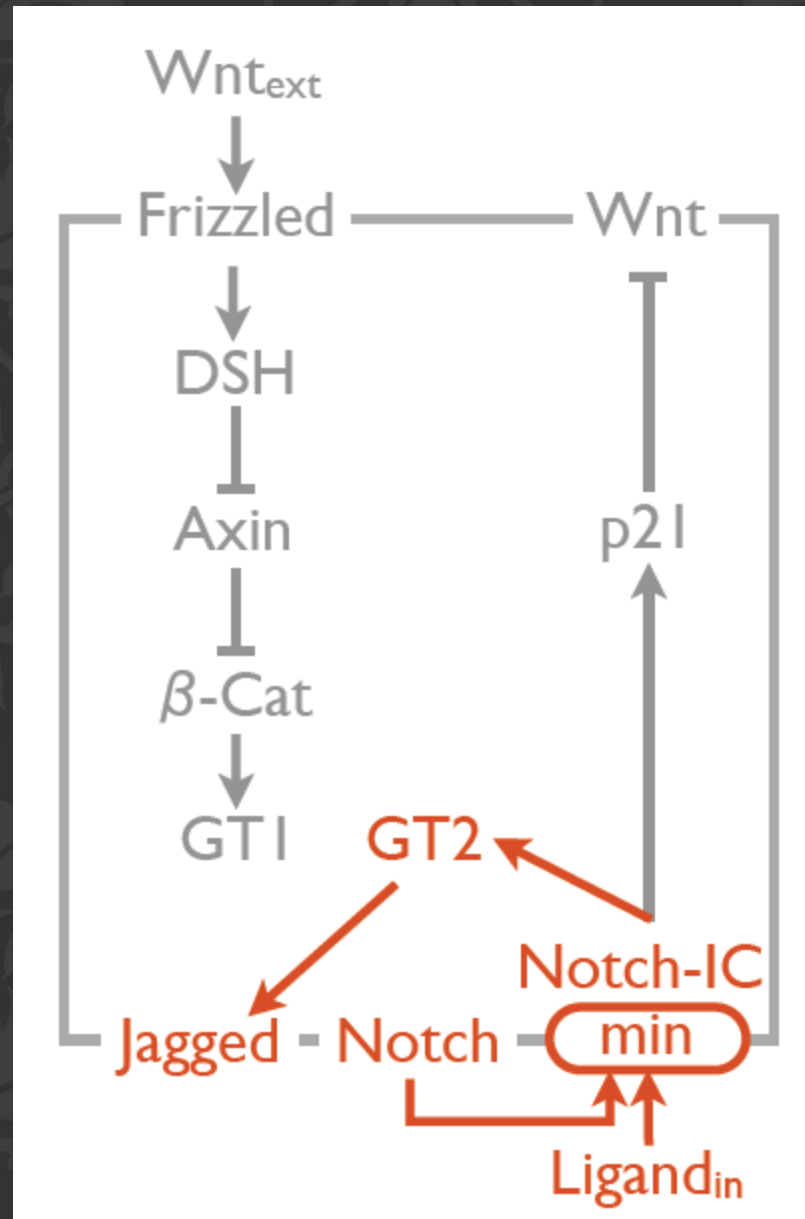


(c)

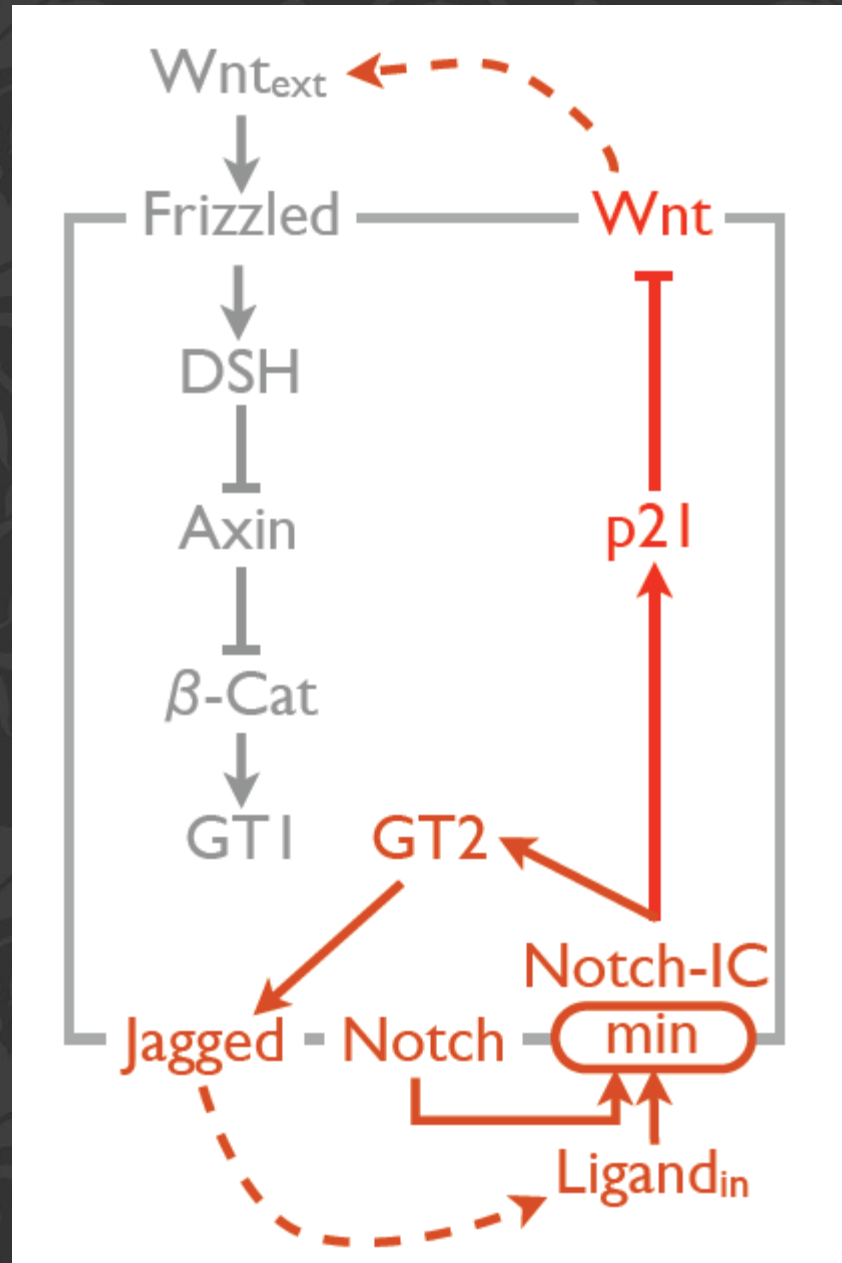
Wnt pathway



Notch pathway



Crosstalk between Notch & Wnt



Requirements

Target genes of Wnt signaling (GT1) maintain the cell in a proliferating state.

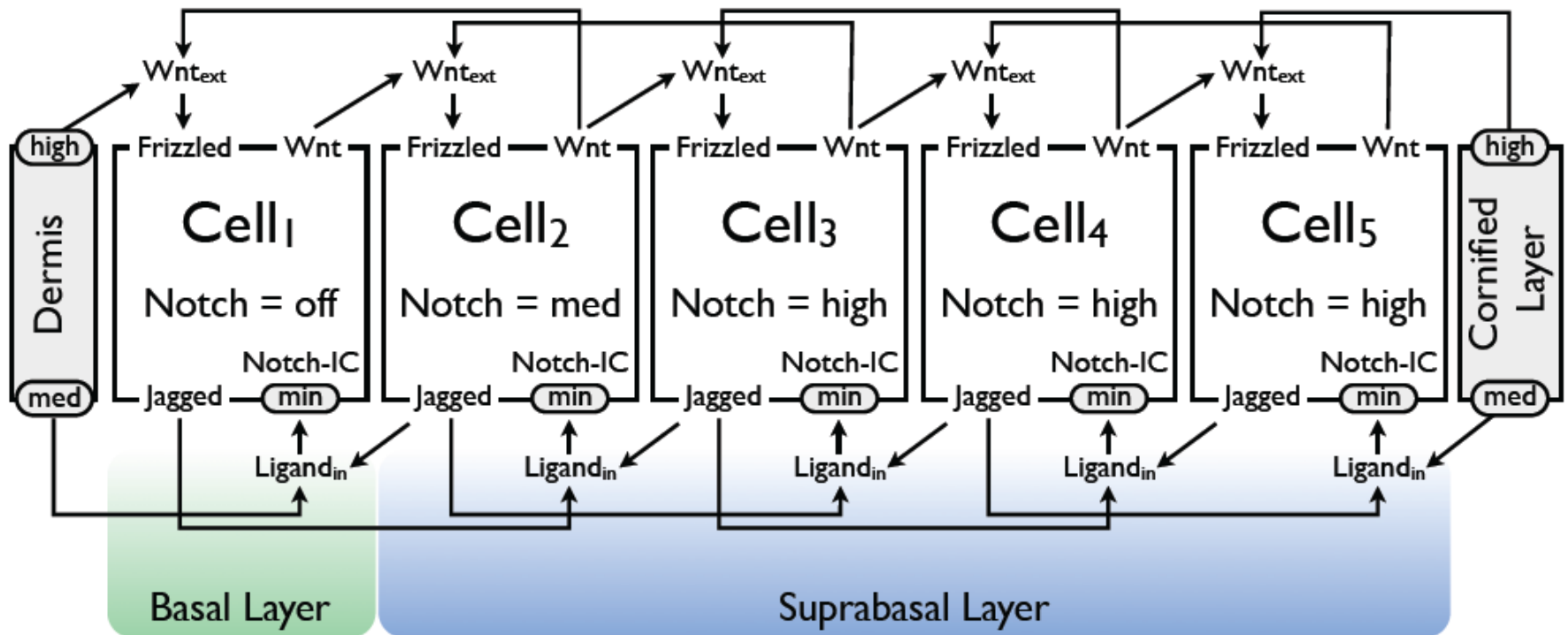
Target genes of Notch signaling (GT2) initiate terminal differentiation.

High-level requirements:

$GT1 > GT2 \rightarrow$ proliferation

$GT1 < GT2 \rightarrow$ differentiation

Model & Requirements



Requirements

GT1 > GT2
proliferating

GT1 = GT2

GT1 < GT2
differentiated

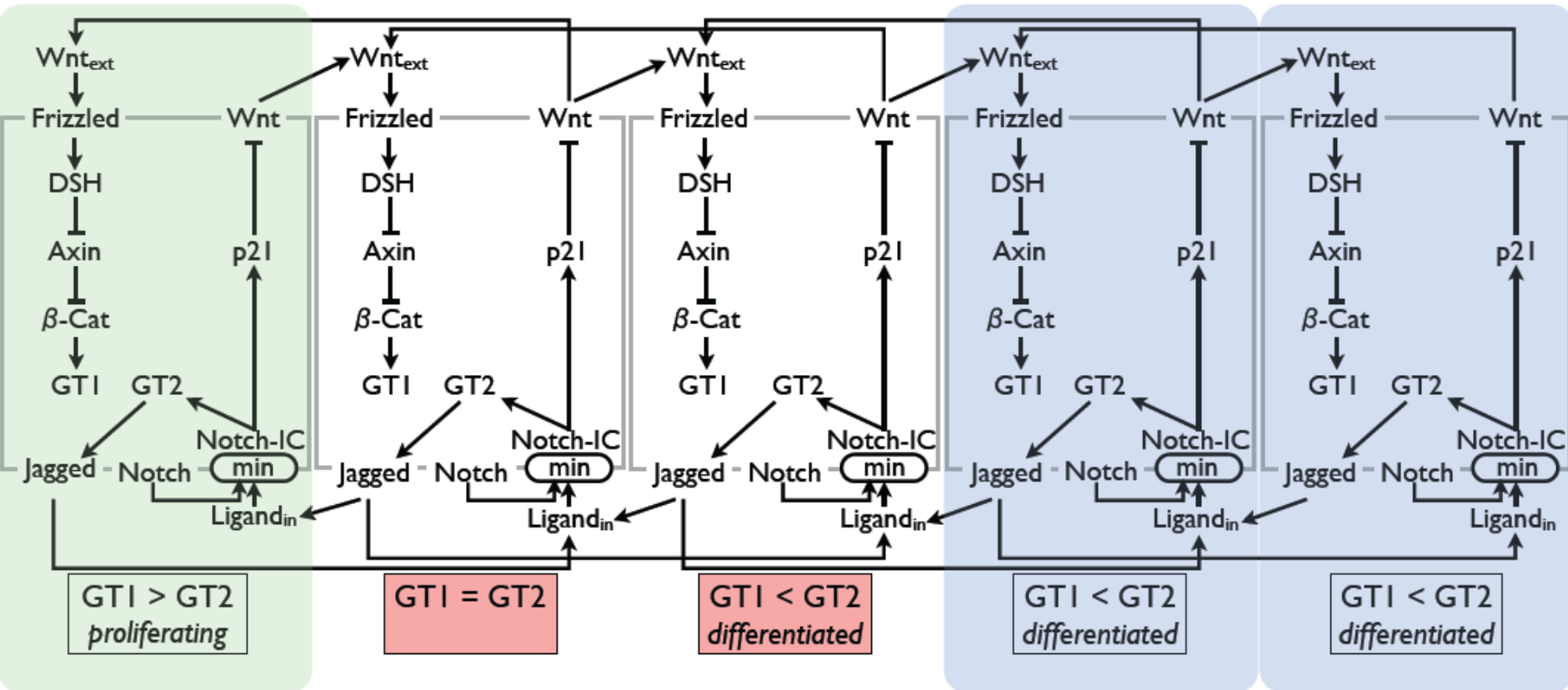
GT1 < GT2
differentiated

GT1 < GT2
differentiated

Gaining new biological insights...



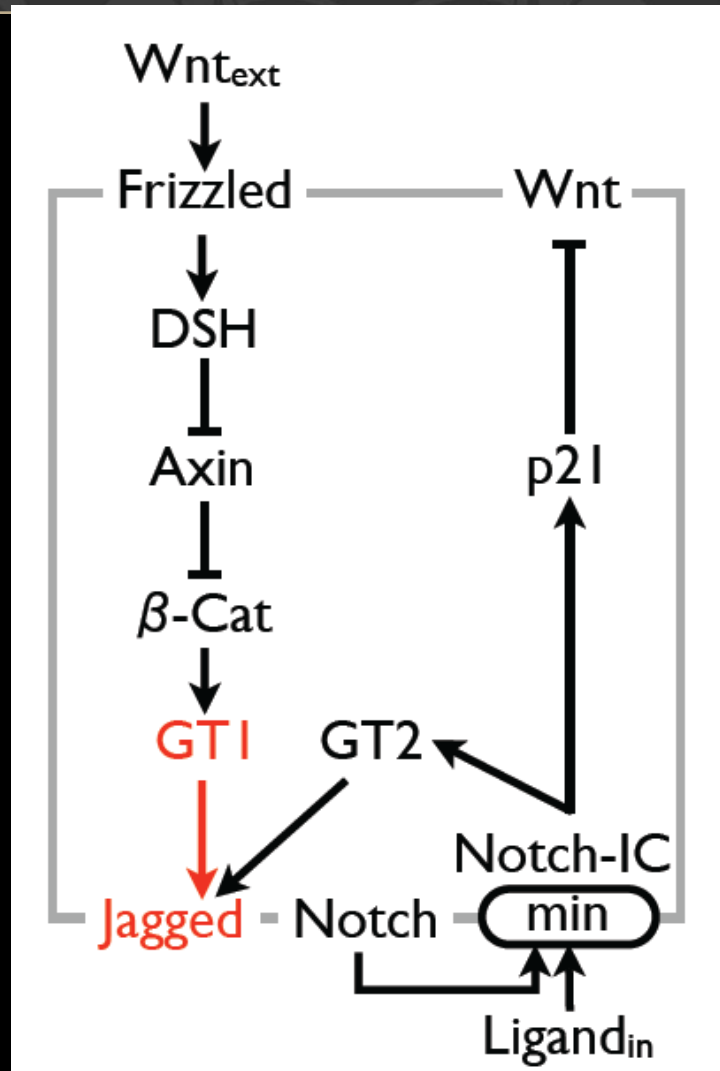
Analysis



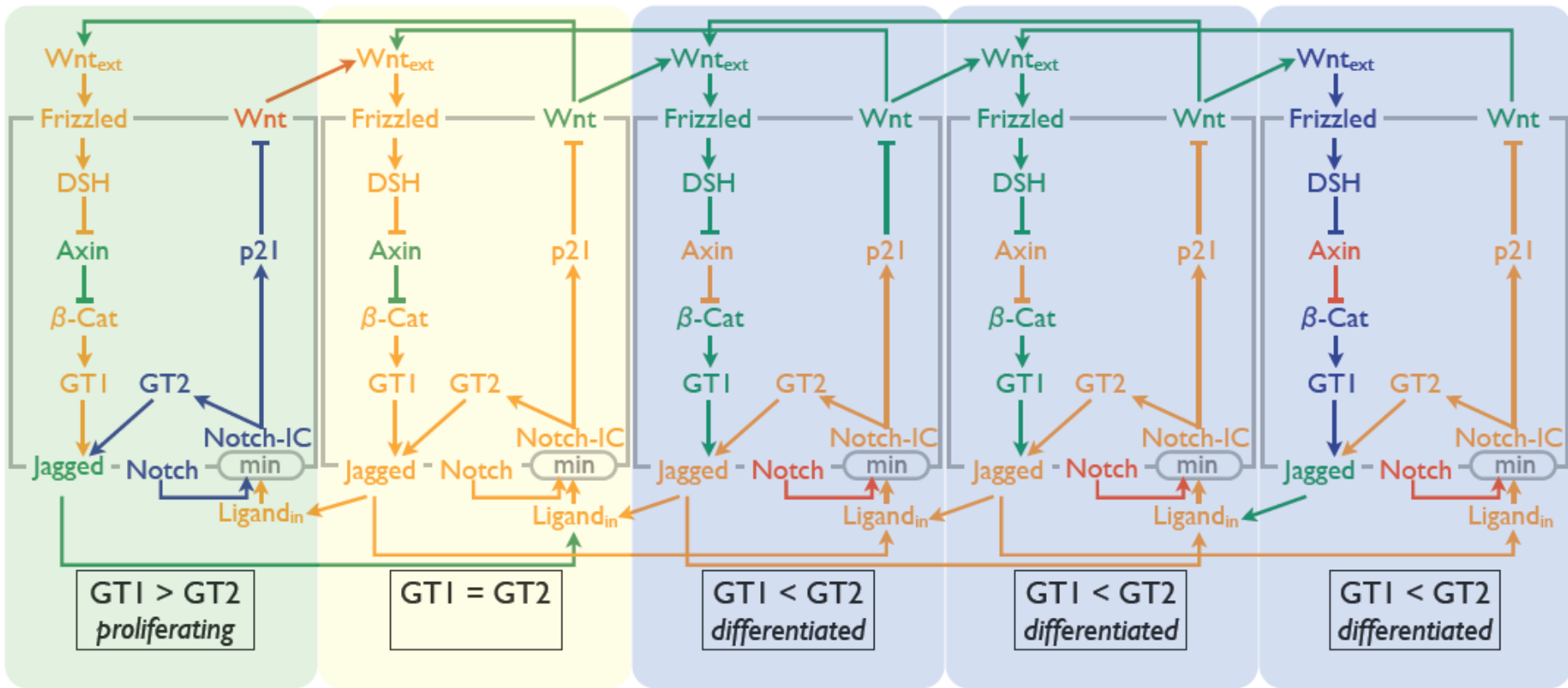
- 6561 infinitely visited states.
- Requirements on cells 1,4,5 are satisfied.
- Requirements on cells 2,3 are **not satisfied**.

New Hypothesis:

Jagged is a downstream target of Wnt signaling



Analysis

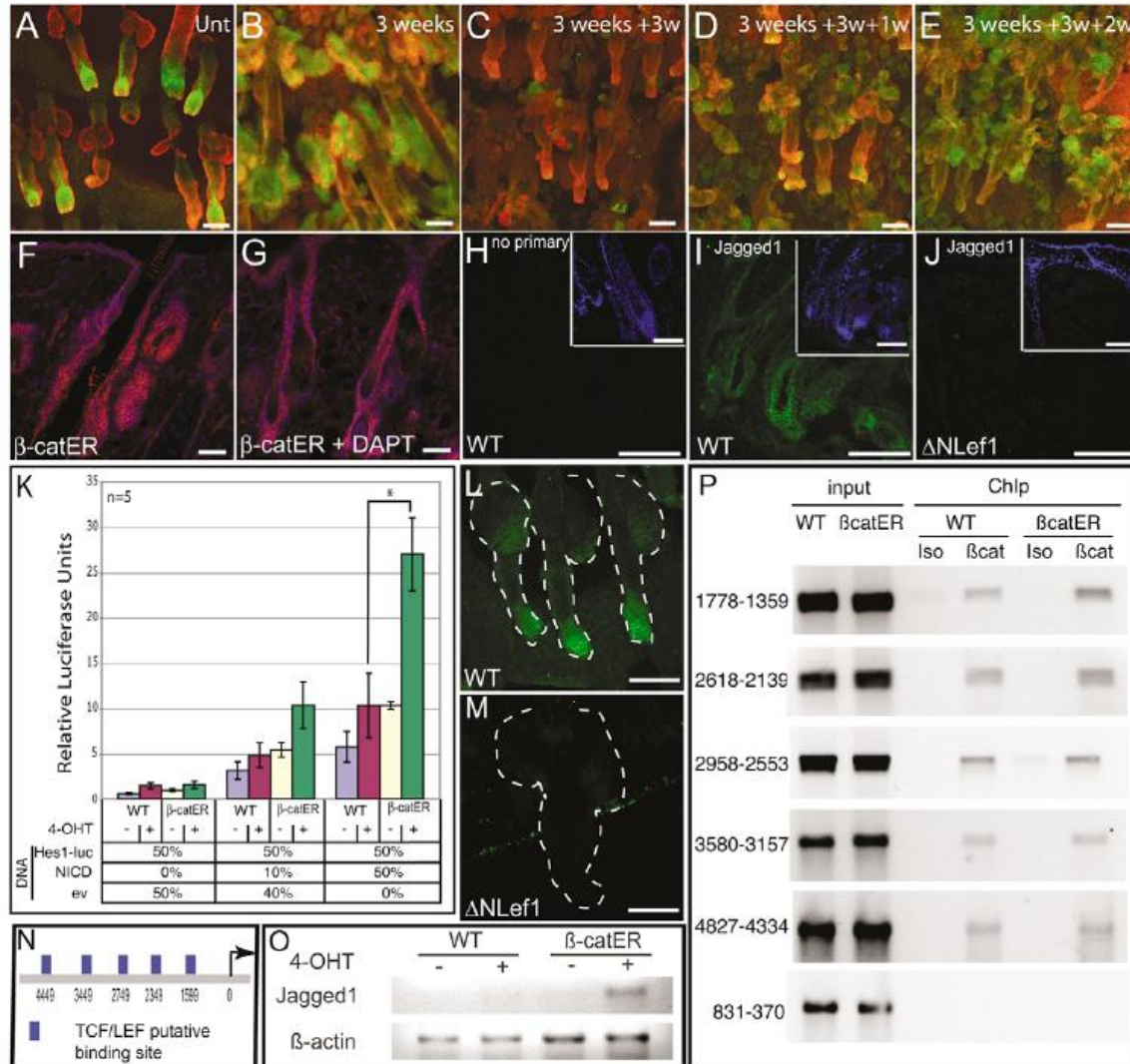


Level of a component: *off*, *low*, *medium*, *high*

- One infinitely visited state.
- Satisfies **all** requirements.

Experimental validation

Jag1 is a direct target gene of β -catenin



Executable modelling of blood cell development from pluripotent embryonic stem cells

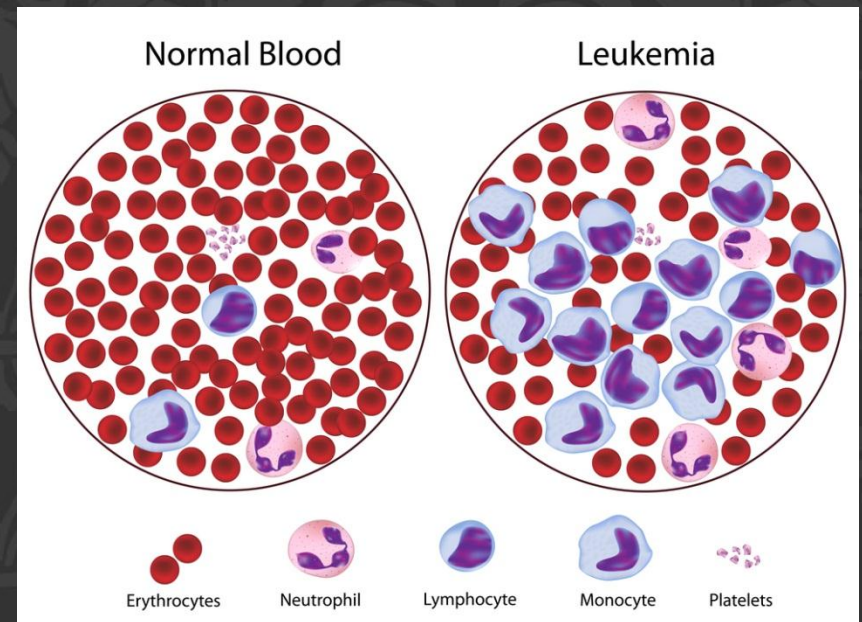
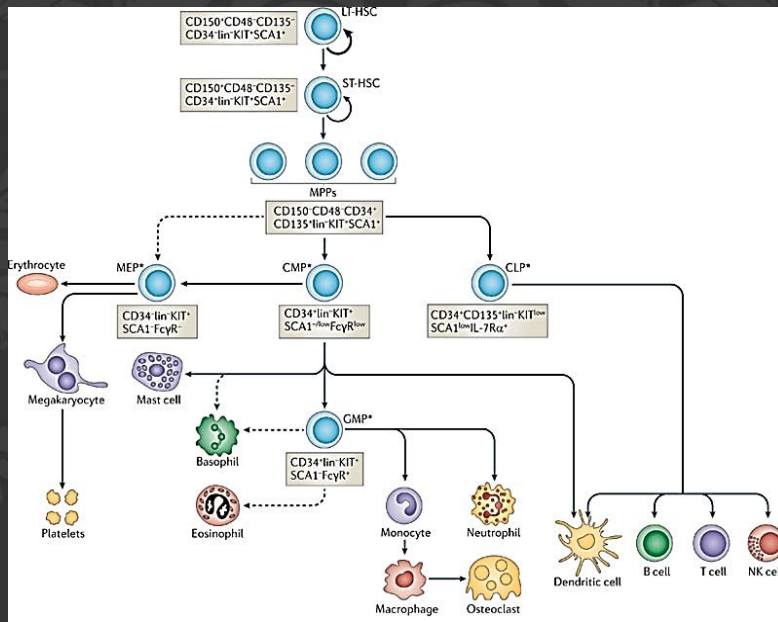


Bertie Gottgens
Cambridge Institute
for Medical Research

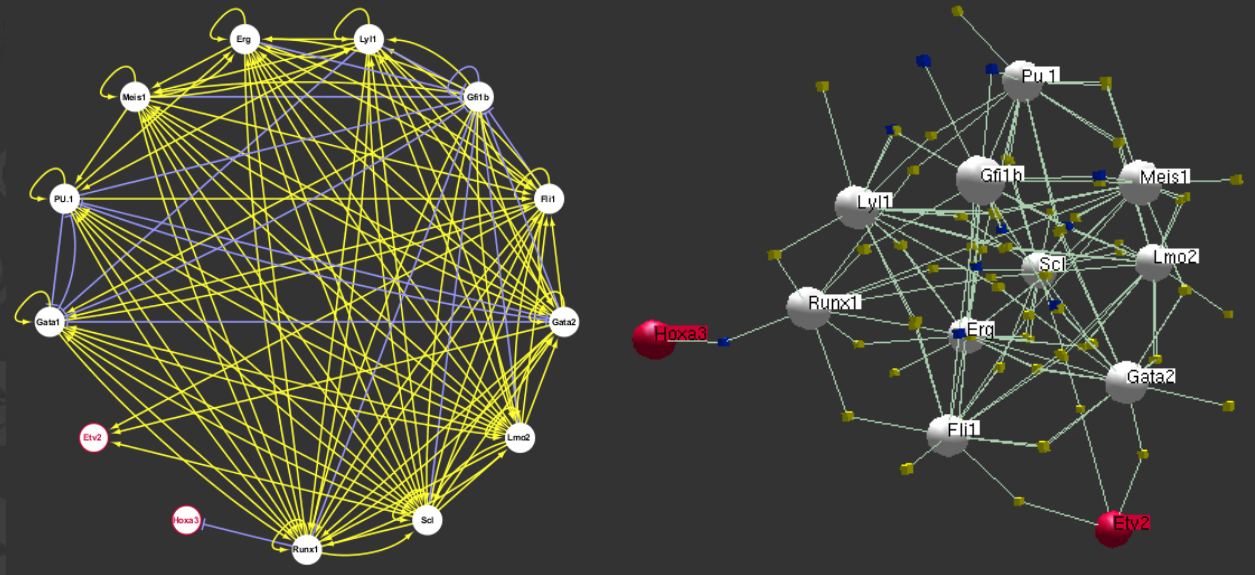


Lucinda Moore
University of
Cambridge

How does normal blood stem cell development turn into leukaemia?

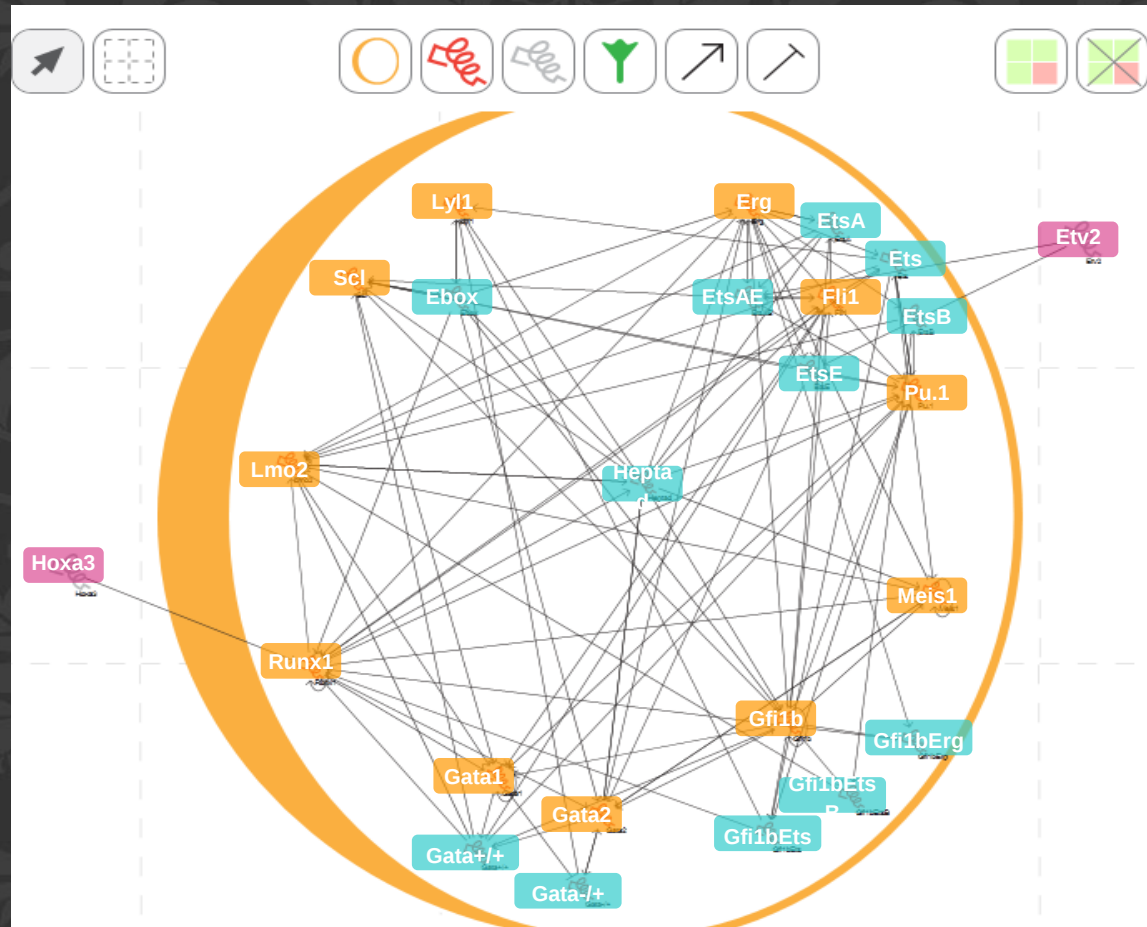


The grand challenge of analysing highly-connected gene regulatory networks

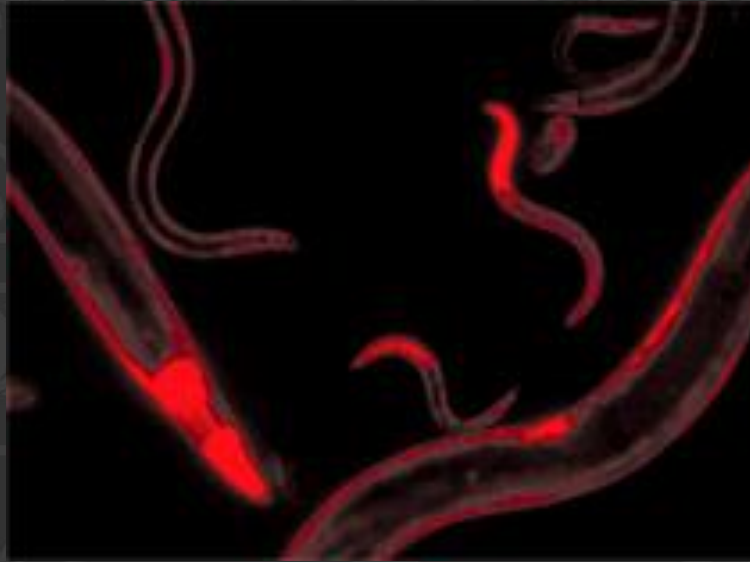


Genes regulating haematopoiesis

Mechanistic insights into leukaemia development



Modelling Cancer Signalling using **State-machines**



PNAS (2005) **102** (6): 1951-1956

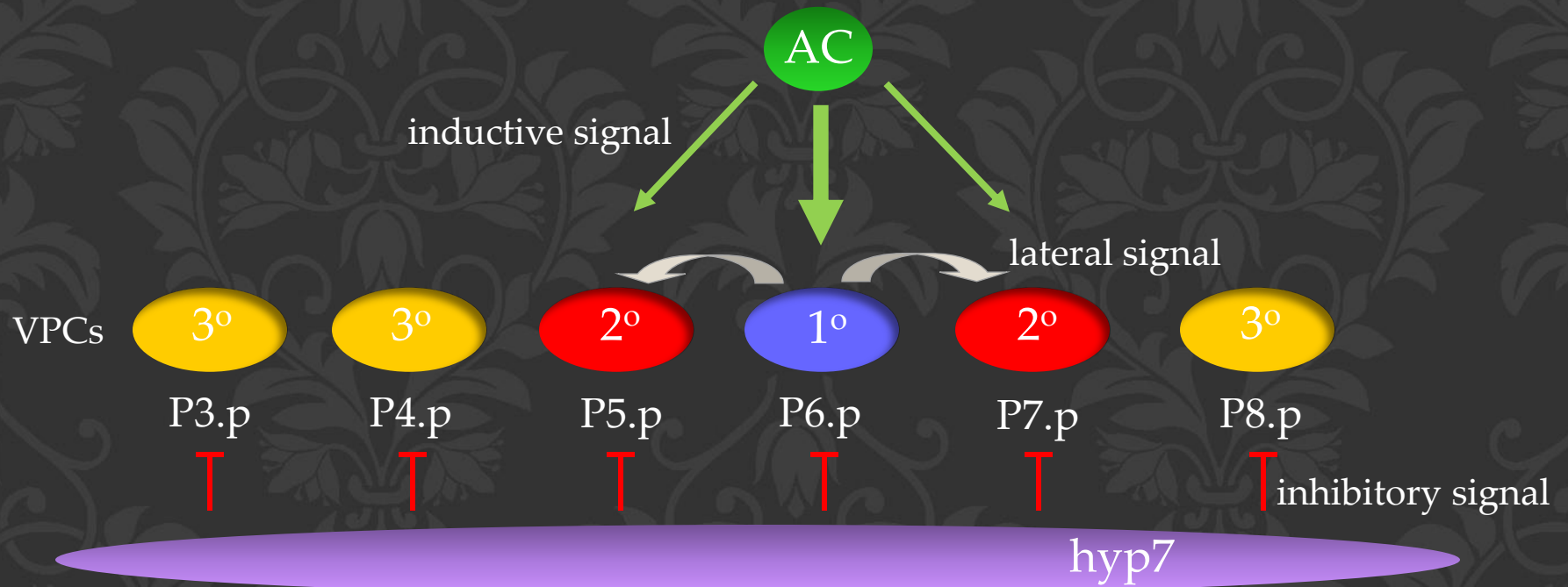
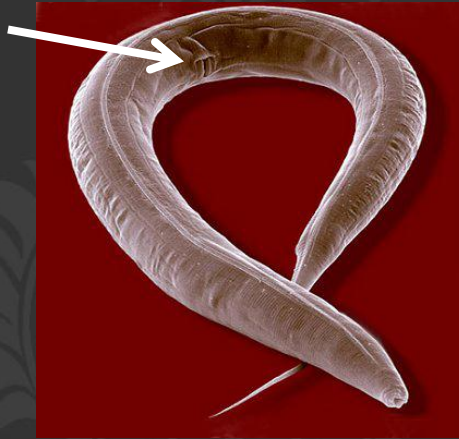
PLoS Comp. Biol. (2007) **3**(5):e92

Mol Sys Biol (2012) **8**:618

Caenorhabditis elegans (C. elegans)

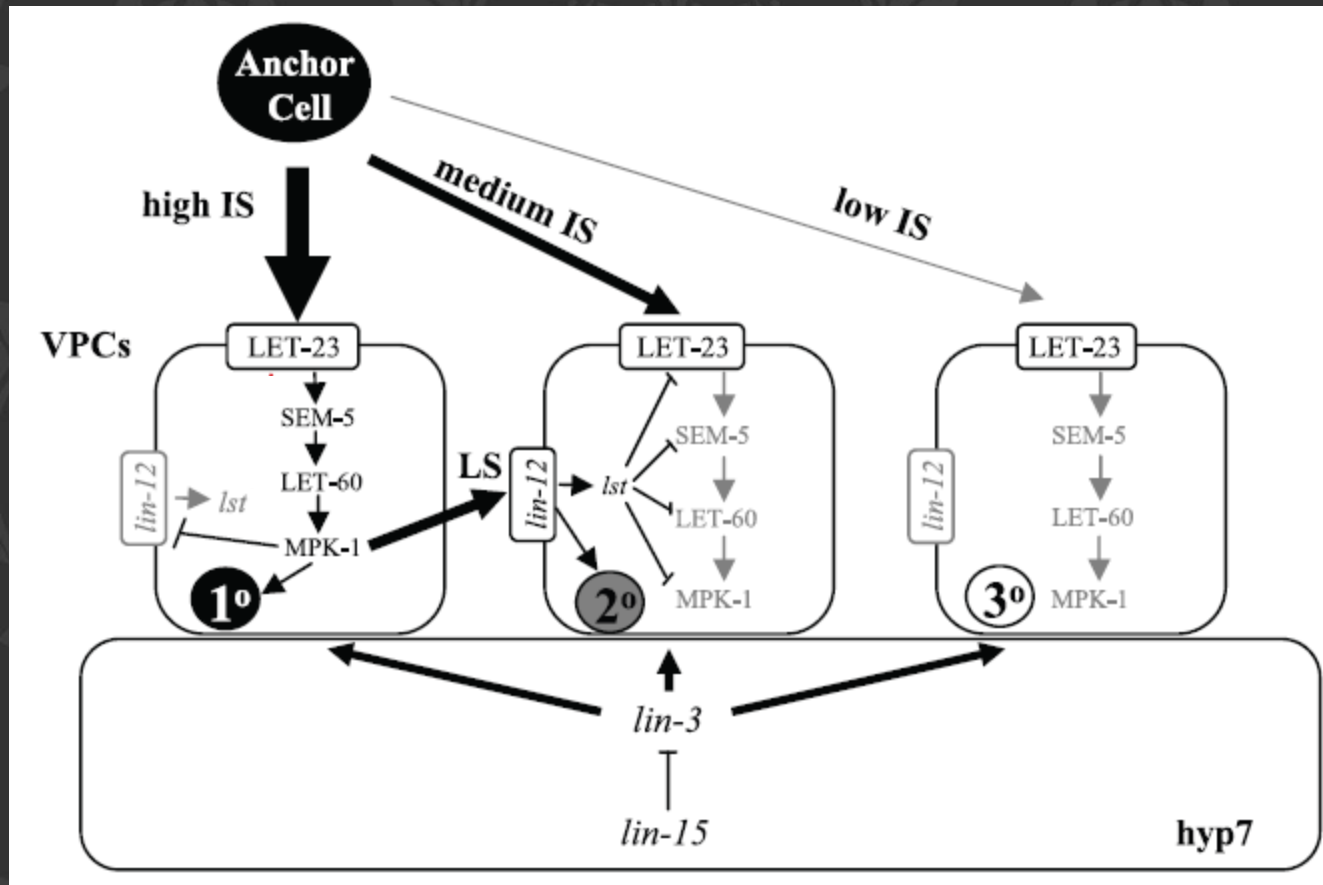
- 1mm long worm
- ~ 1000 cells
- transparent
- easy to handle in the lab
- model for human biology
- very well studied



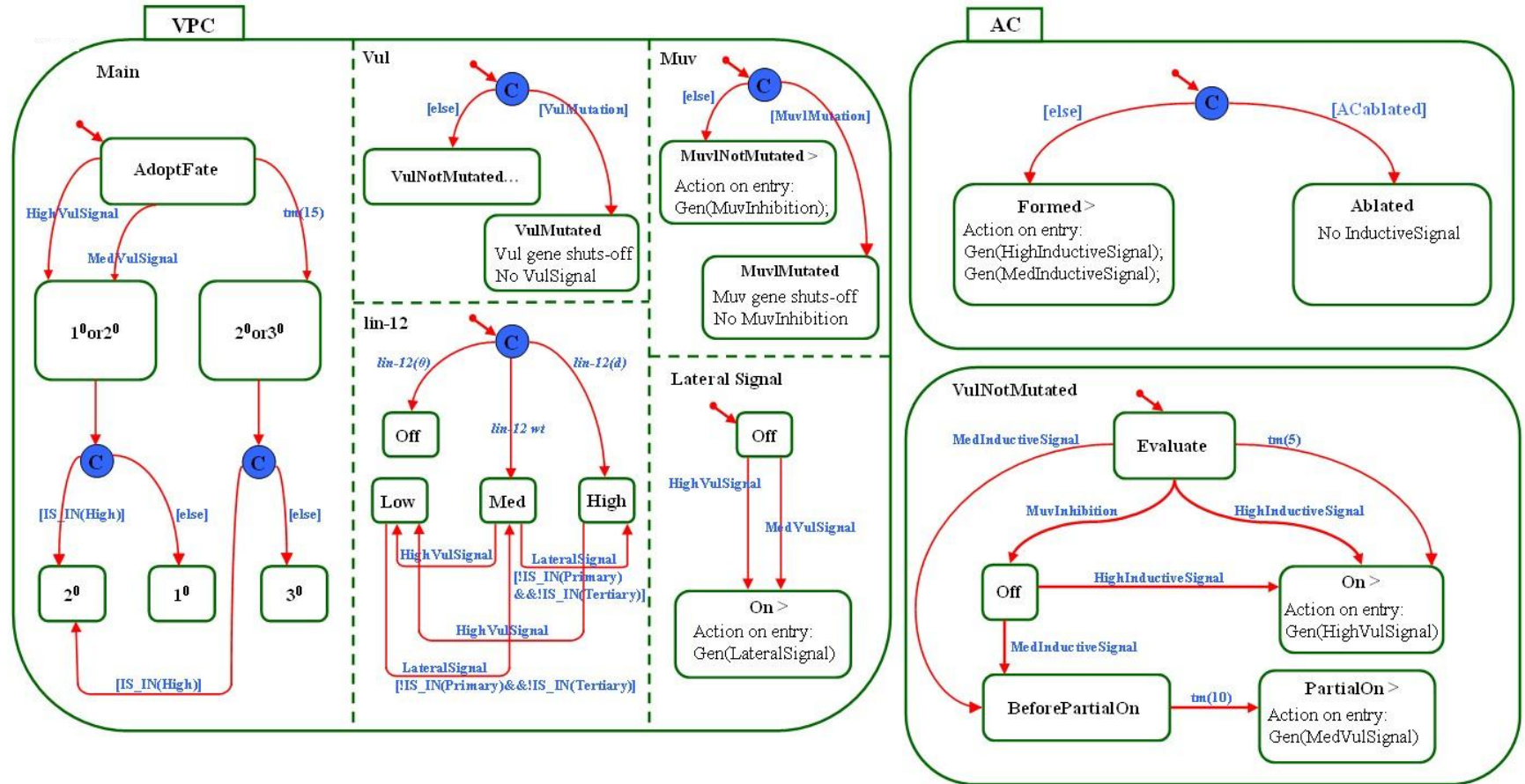


Conceptual understanding

Sternberg & Horwitz, '89; Berset et al., '01; Shaye & Greenwald, '02; Yoo et al., '04, Cui et al., '06

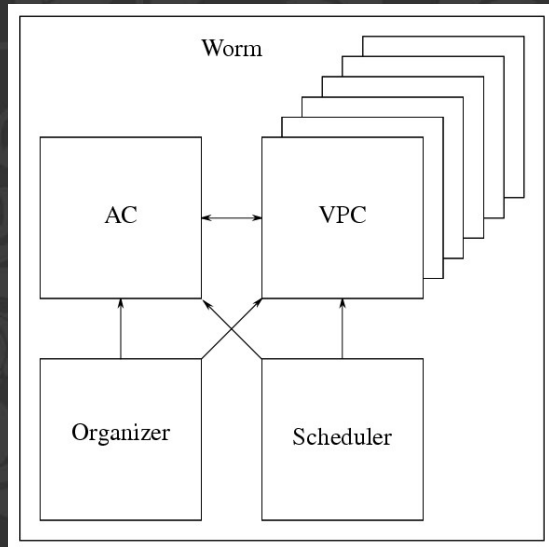


Statecharts VPC Model

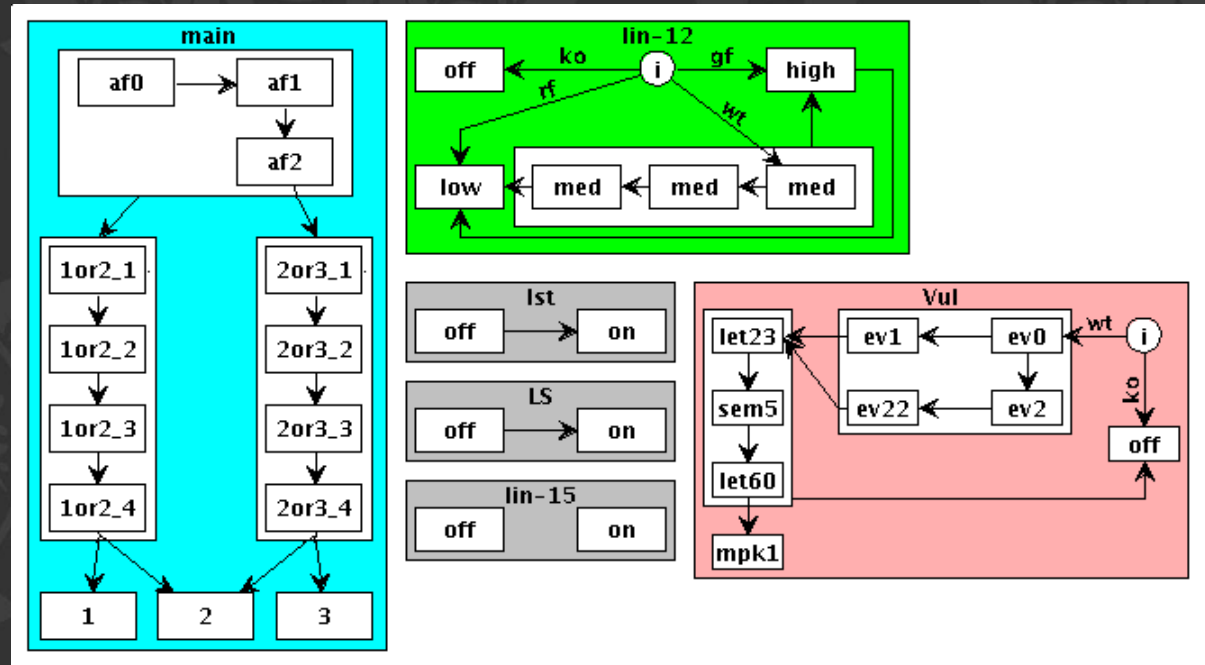


Fisher et al., *PNAS* (2005) 102 (6): 1951-1956

Interacting state-machine model of the worm vulva



Modules comprising the Worm Vulva model



Fisher *et al.*, *PLoS Comp. Biol.* (2007) 3(5):e92

Experimental observations

Table 4. Summary of VPC Fates

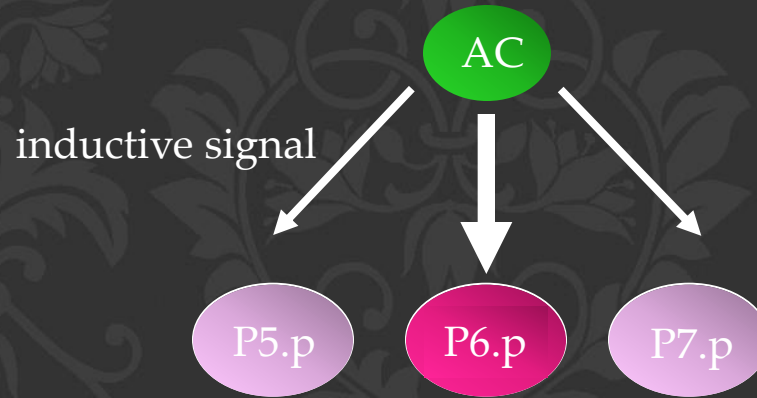
		<i>lin-2(d)</i>						Wild Type						<i>lin-12(0)</i>					
		P3.p	P4.p	P5.p	P6.p	P7.p	P8.p	P3.p	P4.p	P5.p	P6.p	P7.p	P8.p	P3.p	P4.p	P5.p	P6.p	P7.p	P8.p
Muv	ac ⁺	2	1/2	2	ac 1	2	1/2	1/2	1/2	2	ac 1	2	1/2	1	1	1	acac 1	1	1
	ac ⁻	1/2	1/2	1/2	—	1/2	1/2	1/2	1/2	1/2	—	1/2	1/2	1	1	1	—	?	1
+	ac ⁺	2	2	2	ac 1	2	2	3	3	2	ac 1	2	3	3	3	1	acac 1	1	3
	ac ⁻	2	2	2	—	2	2	3	3	3	—	3	3	3	3	3	—	3	3
Vul	ac ⁺	2	2	2	ac 2	2	2	3	3	3	ac 3	3	3	3	3	3	acac 3	3	3
	ac ⁻	2	2	2	—	2	2	3	3	3	—	3	3	3	3	3	—	3	3

In the presence of AC when *Muv* & *lin-12* are mutated, P7.p becomes secondary

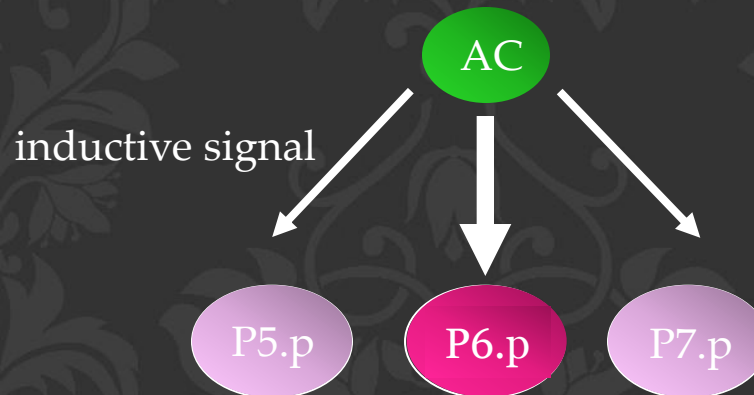
new biological insights...



Known: The inductive signal is spatially graded



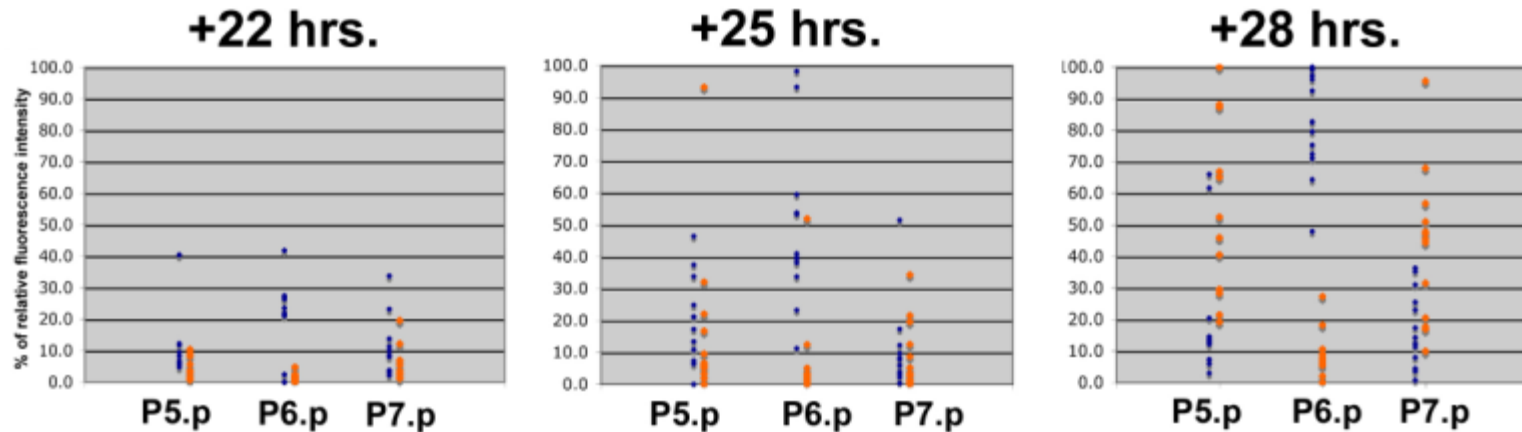
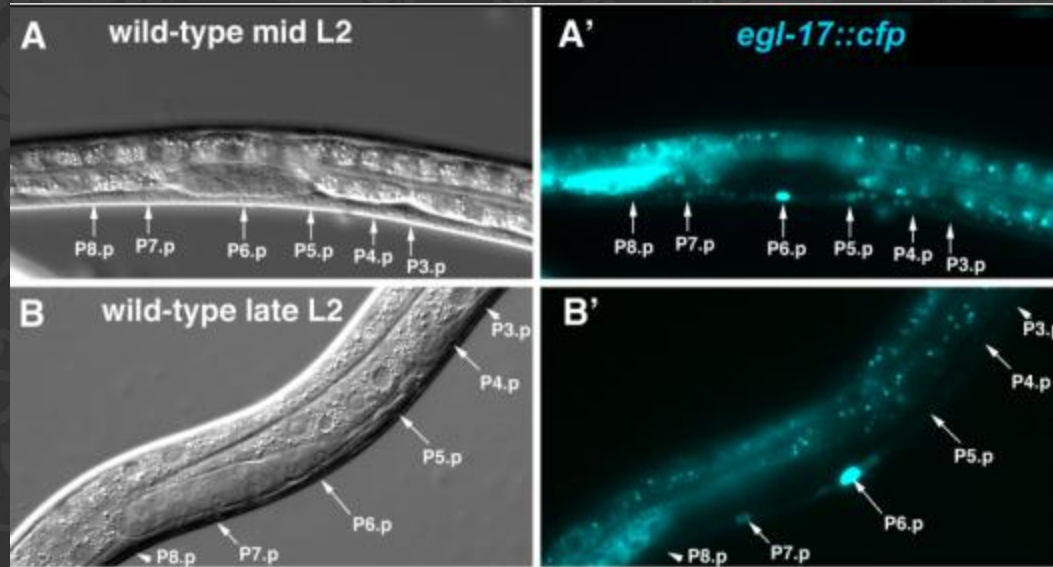
New insight: A temporally-graded response to the inductive signal



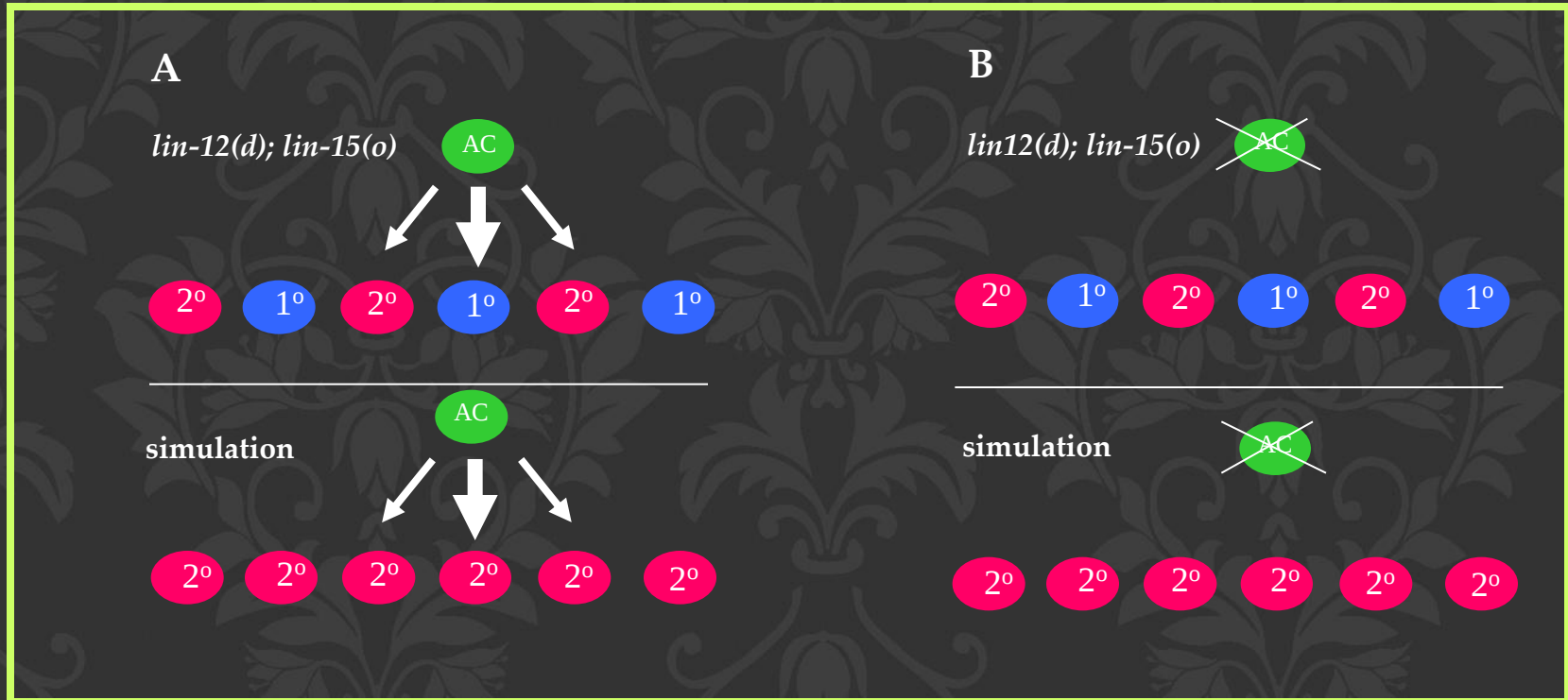
Experimental validation of the model's predictions

Temporal gradient activation of inductive signaling in wild-type animals

Together with
A. Hajnal

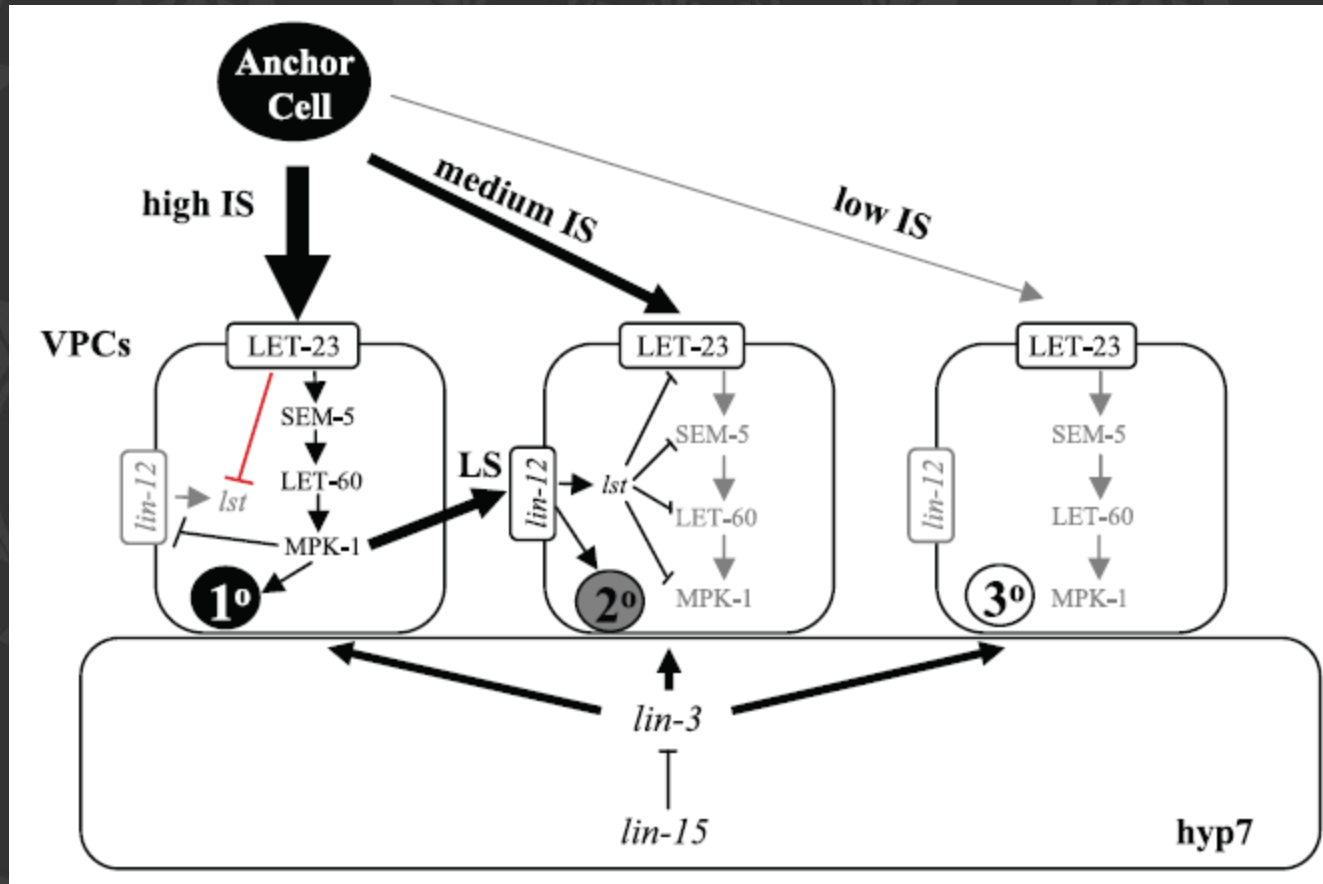


Analyzing the behavior of *lin-12(d);lin-15(o)* mutants



Predicting a new negative feedback loop: EGFR signaling negatively regulates *lst* gene activity

new
insight!



Fisher *et al.*, *PLoS Comp. Biol.* (2007) 3(5):e92

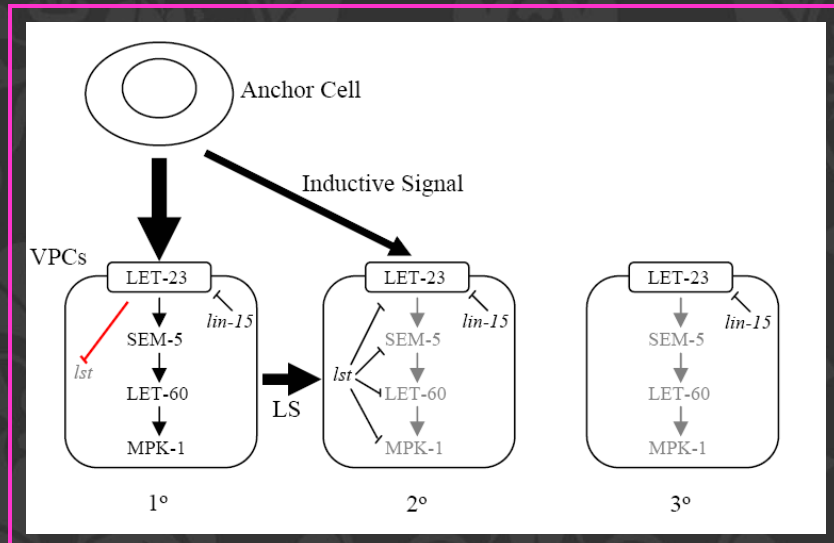
model-checking checks **ALL** possible runs of the model

observations

Table 4. Summary of VPC Fates

		<i>lin-2(d)</i>					
		P3.p	P4.p	P5.p	P6.p	P7.p	P8.p
Muv	ac ⁺	2	1/2	2	ac 1	2	1/2
	ac ⁻	1/2	1/2	1/2	—	1/2	1/2
+	ac ⁺	2	2	2	ac 1	2	2
	ac ⁻	2	2	2	—	2	2
Vul	ac ⁺	2	2	2	ac 2	2	2
	ac ⁻	2	2	2	—	2	2

model



MC

does the model satisfies the data?

yes

continue

no

get a counter example

refinement

model checking enables to **query** the system

e.g., do all possible executions reach a stable state,
independent of the order of reactions between the
VPCs ?

Summary of VPC fate patterns according to the computational model

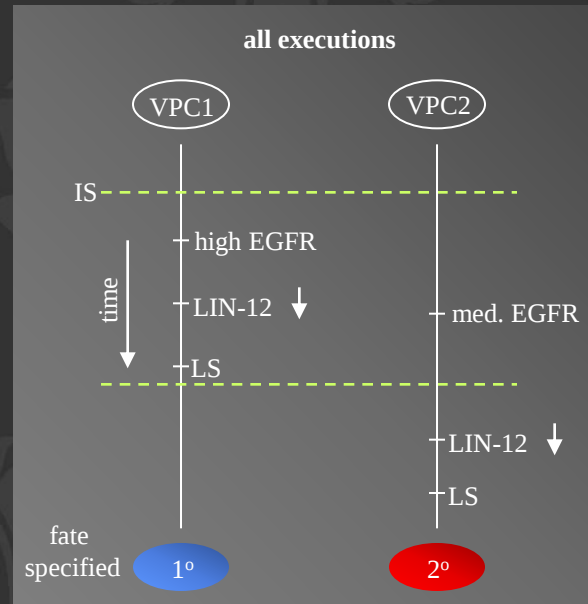
	AC	lin12	lin15	Vul	1st	Predicate name	Fate pattern						Reference & Remarks
							P3.p	P4.p	P5.p	P6.p	P7.p	P8.p	
1	formed	wt	wt	wt	wt	wild_type	3	3	2	1	2	3	Soulston & Horvitz '77
2	formed	wt	wt	wt	ko	1stko	3	3	1	1	1	3	Berset et al. '05, Yoo et al. '04 (by marker expression)
3	formed	wt	wt	ko	wt	Vulko	3	3	3	3	3	3	Sternberg & Horvitz '89
4	formed	wt	wt	ko	ko	Vulkolstko	3	3	3	3	3	3	n.d. (only partial Vul(rf) mutants tested)
5	formed	wt	ko	wt	wt	lin15ko	1/2	1/2	2	1	2	1/2	Sternberg & Horvitz '89
6	formed	wt	ko	wt	ko	lin15kolstko	1	1	1	1	1	1	Berset et al. '01(marker expression)
7	formed	wt	ko	ko	wt	lin15koVulko	3	3	3	3	3	3	Ferguson & Horvitz '87, Sternberg & Horvitz '89, Cui et al '06
8	formed	wt	ko	ko	ko	lin15koVulkolstko	3	3	3	3	3	3	n.d.
9	formed	ko	wt	wt	wt	lin12ko	3	3	1	1	1	3	Sternberg & Horvitz '89, Greenwald et al. '83
10	formed	ko	wt	wt	ko	lin12kolstko	3	3	1	1	1	3	Berset & Hajnal, unpublished results
11	formed	ko	wt	ko	wt	Vulkolin12ko	3	3	3	3	3	3	Sternberg & Horvitz '89 (rf Vul mutants)
12	formed	ko	wt	ko	ko	lin12koVulkolstko	3	3	3	3	3	3	n.d.
13	formed	ko	ko	wt	wt	lin15kolin12ko	1	1	1	1	1	1	Sternberg & Horvitz '89
14	formed	ko	ko	wt	ko	lin12kolin15kolstko	1	1	1	1	1	1	n.d.
15	formed	ko	ko	ko	wt	lin12kolin15koVulko	3	3	3	3	3	3	n.d.
16	formed	ko	ko	ko	ko	lin12kolin15koVulkolstko	3	3	3	3	3	3	n.d.
17	formed	gf	wt	wt	wt	lin12d	2	2	2	1	2	2	Sternberg & Horvitz '89 [(lin-12(gf)/lin-12(lf)]
18	formed	gf	wt	wt	ko	lin12dlstko	2	2	1	1	1	2	n.d.
19	formed	gf	wt	ko	wt	Vulkolin12d	2	2	2	2	2	2	Sternberg & Horvitz '89 [(lin-12(gf)/lin-12(lf) with vul(rf)]
20	formed	gf	wt	ko	ko	lin12dVulkolstko	2	2	2	2	2	2	n.d.
21	formed	gf	ko	wt	wt	lin15kolin12d	1/2	1/2	2	1	2	1/2	Sternberg & Horvitz '89 [(lin-12(gf)/lin-12(lf)]
22	formed	gf	ko	wt	ko	lin12dlin15kolstko	1	1	1	1	1	1	n.d.
23	formed	gf	ko	ko	wt	lin12dlin15koVulko	2	2	2	2	2	2	n.d.
24	formed	gf	ko	ko	ko	lin12dlin15koVulkolstko	2	2	2	2	2	2	n.d.
25	ablated	wt	wt	wt	wt	ac-	3	3	3	3	3	3	Kimble '81
26	ablated	wt	wt	wt	ko	ac-_1stko	3	3	3	3	3	3	Berset et al. '05
27	ablated	wt	wt	ko	wt	ac-_Vulko	3	3	3	3	3	3	n.d.
28	ablated	wt	wt	ko	ko	ac-_Vulkolstko	3	3	3	3	3	3	n.d.
29	ablated	wt	ko	wt	wt	ac-_lin15ko	1/2	1/2	1/2	1/2	1/2	1/2	Sternberg & Horvitz '89
30	ablated	wt	ko	wt	ko	ac-_lin15kolstko	1	1	1	1	1	1	n.d.
31	ablated	wt	ko	wt	ko	ac-_lin15koVulko	3	3	3	3	3	3	n.d.
32	ablated	wt	ko	ko	ko	ac-_lin15koVulkolstko	3	3	3	3	3	3	n.d.
33	ablated	ko	wt	wt	wt	ac-_lin12ko	3	3	3	3	3	3	Sternberg & Horvitz '89
34	ablated	ko	wt	wt	ko	ac-_lin12kolstko	3	3	3	3	3	3	n.d.
35	ablated	ko	wt	ko	wt	ac-_Vulkolin12ko	3	3	3	3	3	3	n.d.
36	ablated	ko	wt	ko	ko	ac-_lin12koVulkolstko	3	3	3	3	3	3	n.d.
37	ablated	ko	ko	wt	wt	ac-_lin15kolin12ko	1	1	1	1	1	1	Sternberg & Horvitz '89 (one animal)
38	ablated	ko	ko	wt	ko	ac-_lin12kolin15kolstko	1	1	1	1	1	1	n.d.
39	ablated	ko	ko	ko	wt	ac-_lin12kolin15koVulko	3	3	3	3	3	3	n.d.
40	ablated	ko	ko	ko	ko	ac-_lin12kolin15koVulkolstko	3	3	3	3	3	3	n.d.
41	ablated	gf	wt	wt	wt	ac-_lin12d	2	2	2	2	2	2	Sternberg & Horvitz '89
42	ablated	gf	wt	wt	ko	ac-_lin12dlstko	2	2	2	2	2	2	Berset et al. '05 (for lip-1)
43	ablated	gf	wt	ko	wt	ac-_lin12dVulko	2	2	2	2	2	2	Sternberg & Horvitz '89 (rf mutants), Han et al. '90
44	ablated	gf	wt	ko	ko	ac-_lin12dVulkolstko	2	2	2	2	2	2	n.d. (let-60dneg)
45	ablated	gf	ko	wt	wt	ac-_lin15kolin12d	1/2	1/2	1/2	1/2	1/2	1/2	Sternberg & Horvitz '89
46	ablated	gf	ko	wt	ko	ac-_lin12dlin15kolstko	1	1	1	1	1	1	n.d.
47	ablated	gf	ko	ko	wt	ac-_lin12dlin15koVulko	2	2	2	2	2	2	n.d.
48	ablated	gf	ko	ko	ko	ac-_lin12dlin15koVulkolstko	2	2	2	2	2	2	n.d.

another query:

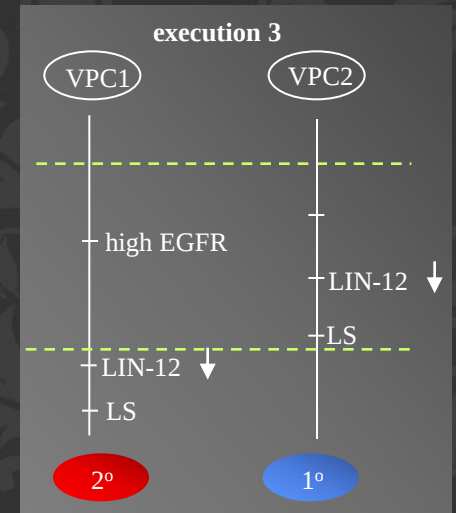
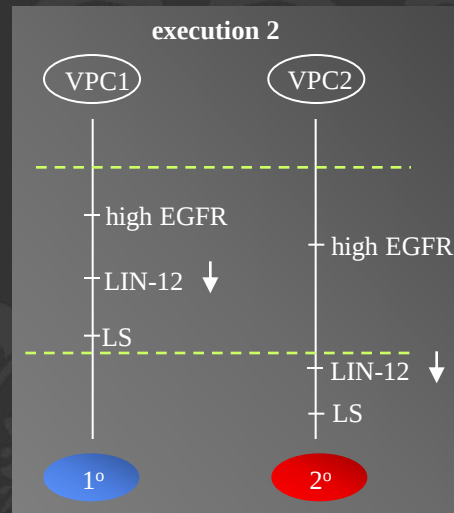
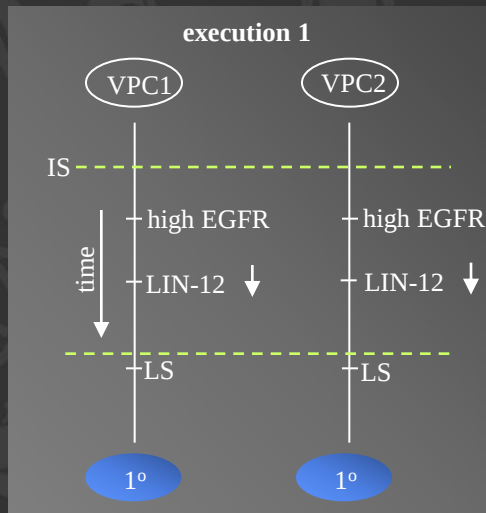
Is it possible to get an unstable fate pattern without allowing variations in the timing of the lateral signal?

Sequence of events leading to stable and unstable fate patterns

A Stable pattern

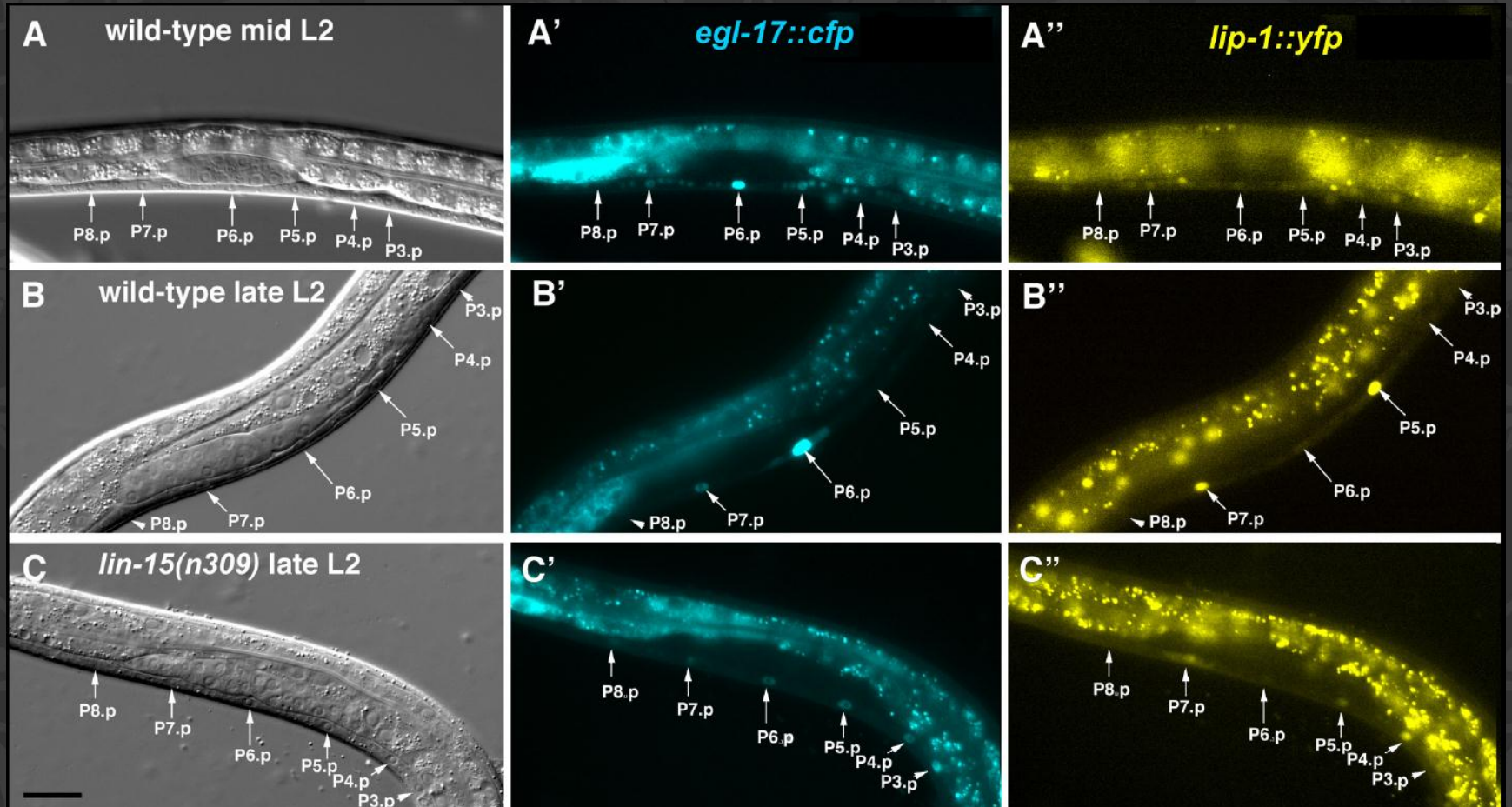


B Unstable pattern



Experimental validation of the model's predictions

Loss of sequential induction in *lin-15* mutants

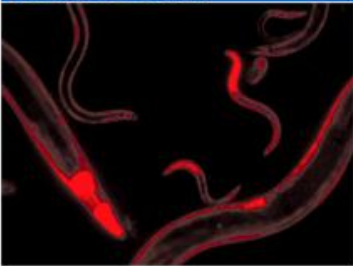


Concurrency reveals... a molecular synchronization mechanism

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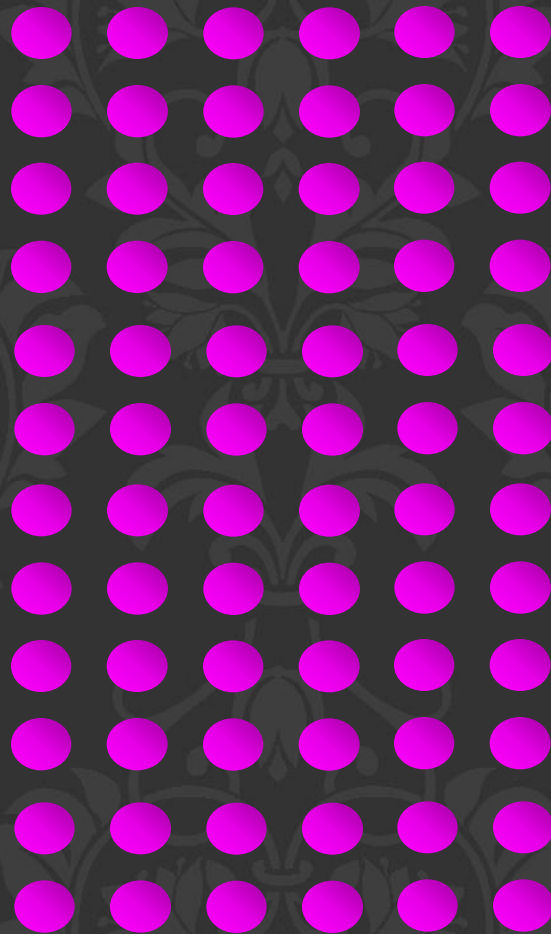
Welcome to *Molecular Systems Biology*

Featured article

Cell-cycle regulation of NOTCH signaling during *C. elegans* vulval development

Through an iterative process of computational modeling, prediction, and experimentation, a molecular synchronization mechanism is revealed by which the cell-cycle regulates Notch signaling to allow the formation of a stable cell fate pattern.

synchronous



synchronous

e.g., *lin-15(o)*

1°

1°

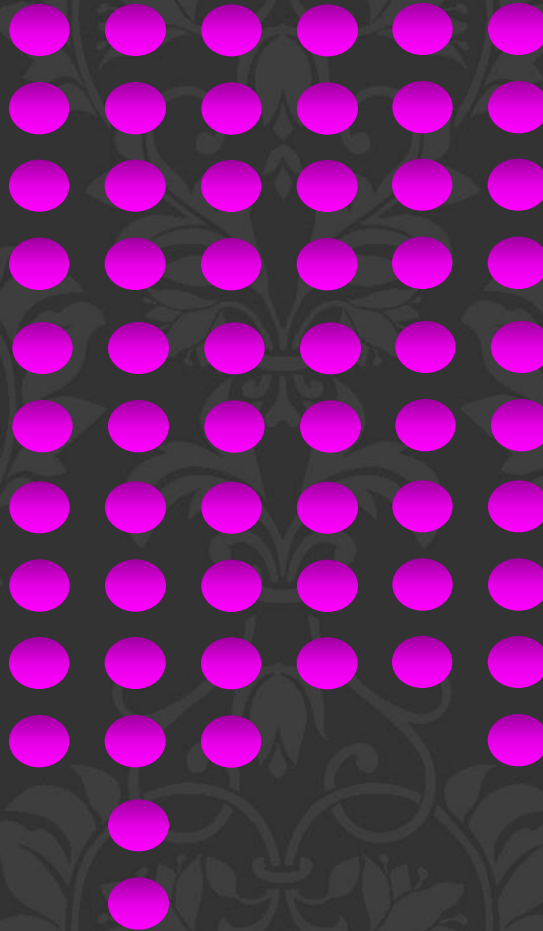
1°

1°

1°

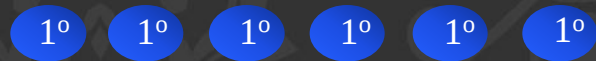
1°

asynchronous



synchronous

e.g., *lin-15(o)*



asynchronous

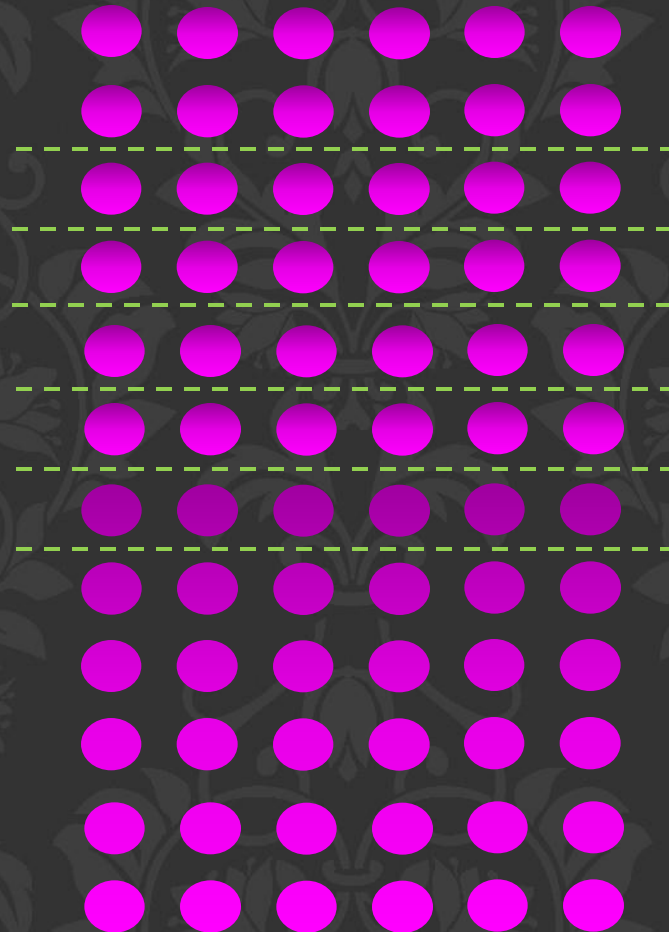
e.g., wt



**A notion of concurrency tailored for cell-cell interactions
'bounded-asynchrony'**

Fisher *et al.*, *Formal Methods in Systems Biology*; LNBI Vol. 5054, pp. 17-32, (2008)

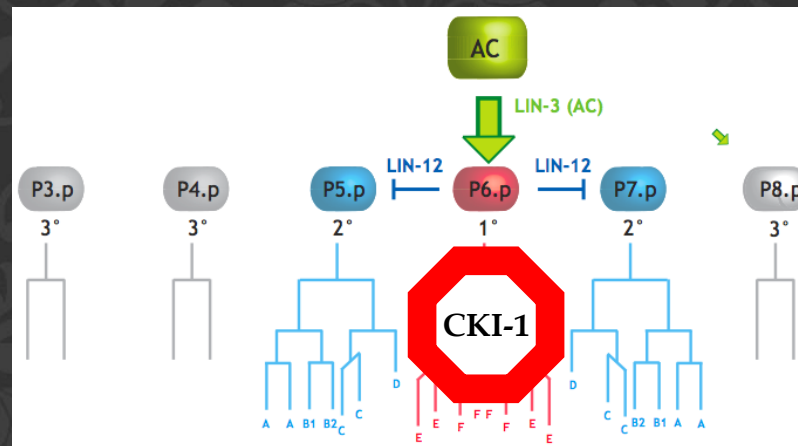
bounded asynchrony



What is the biological counterpart of the scheduler?

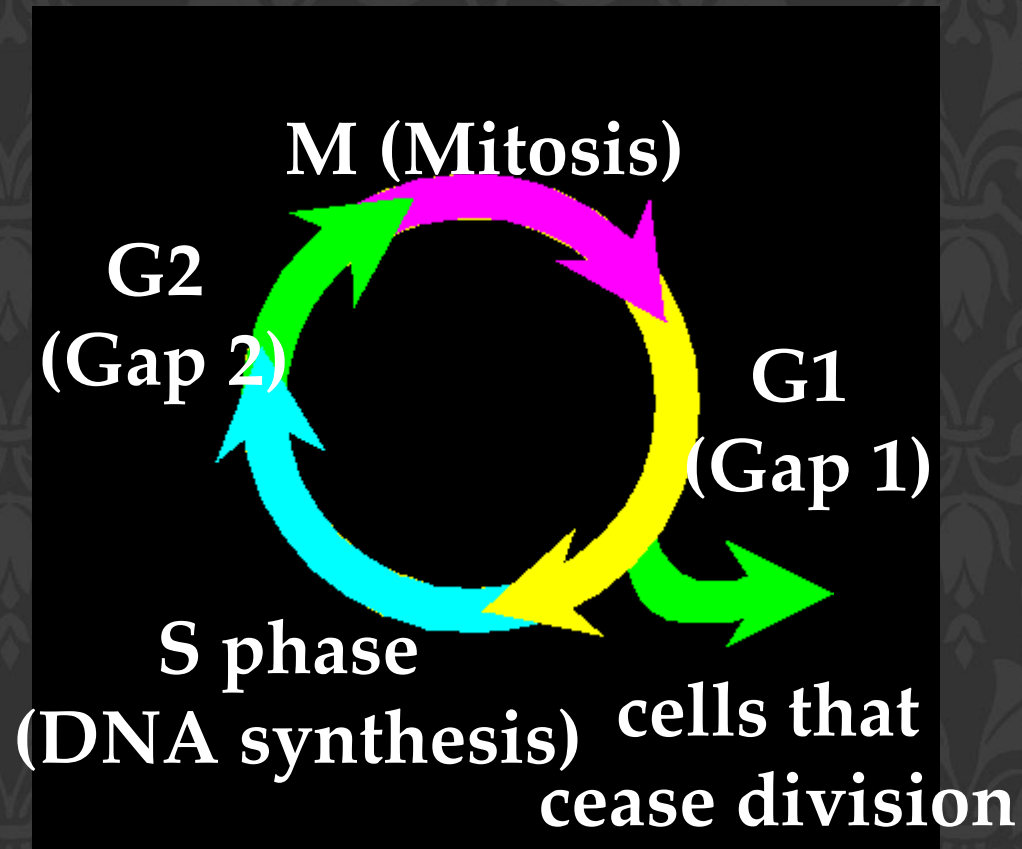
Model prediction: An impaired “scheduler” leads to affected cell fate patterning

Hypothesis: The cell cycle corresponds to the scheduler

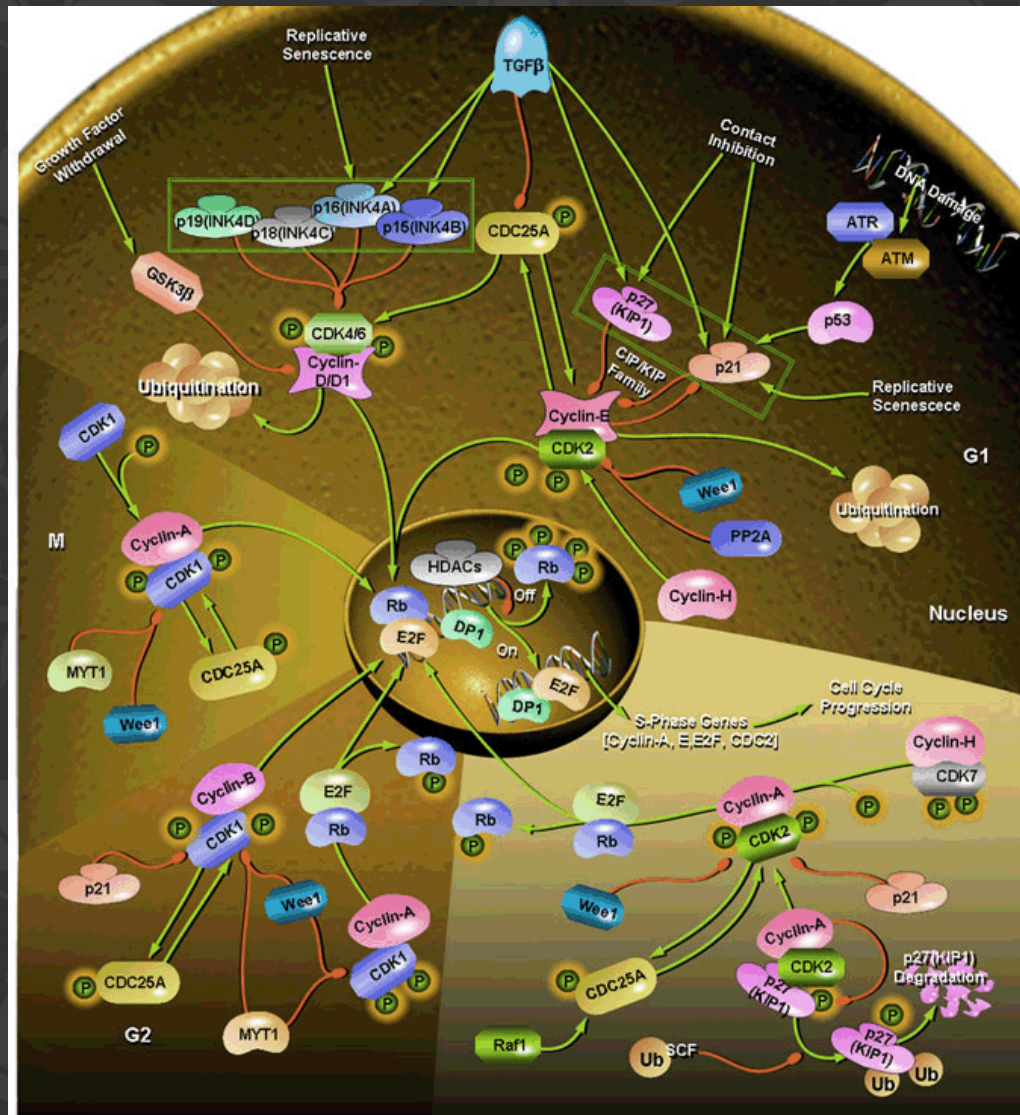


⇒ What happens to expression pattern of different cell fate markers in P6.p?

Cell-cycle



Cell-cycle regulation



Cell-cycle Regulation of Cancer Signalling



Alex Hajnal



Stefanie Nusser



Ivo Rimann



Magdalene Adamczyk



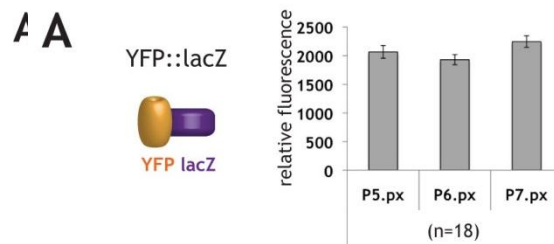
Antje Beyer
University of
Cambridge

Zurich University

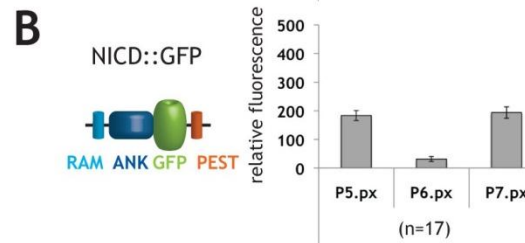
Nusser-Stein *et al.* *Molecular Systems Biology* 8:618, 2012

1400

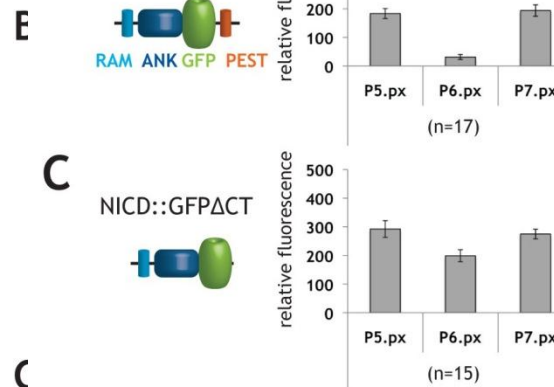
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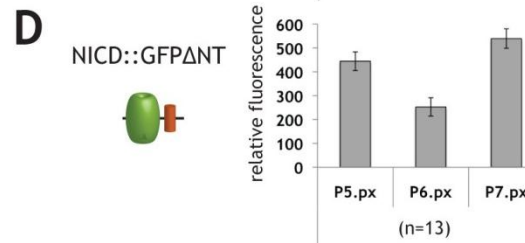
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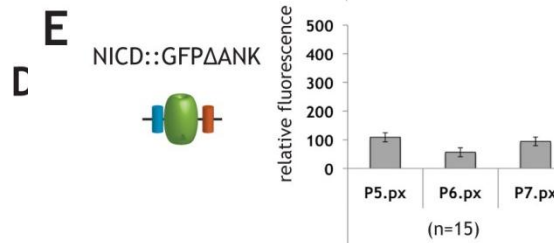
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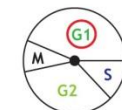
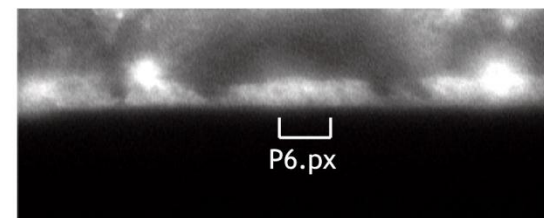
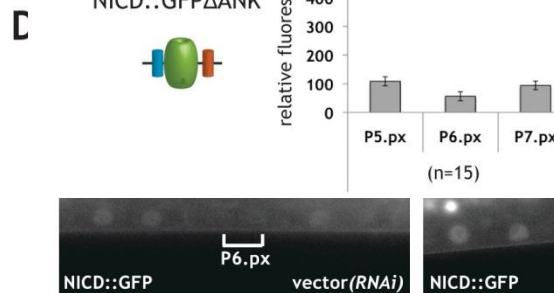
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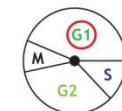
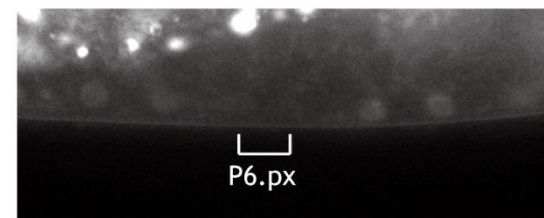
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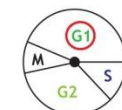
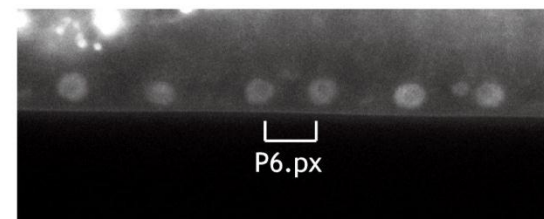
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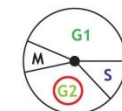
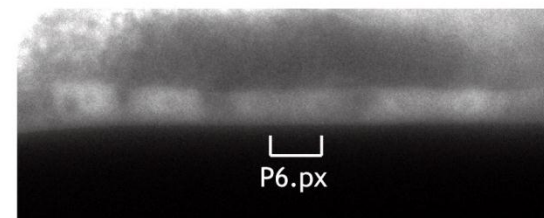
■ wild-type (n=24)
■ *cyd-1(q626)* (n=31)



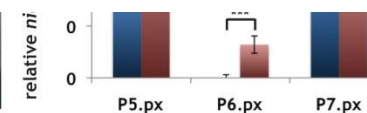
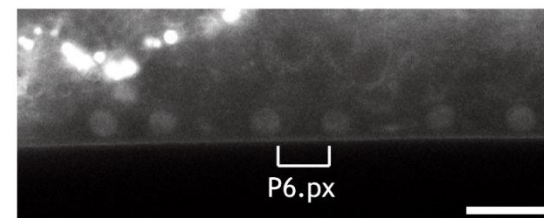
■ wild-type (n=14)
■ *cyd-1(q626)* (n=23)



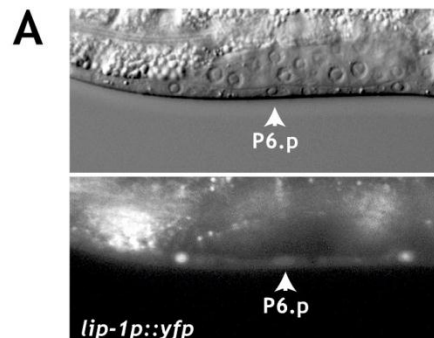
■ *cye-1(ku256)/+* (n=23)
■ *cye-1(ku256)* (n=23)



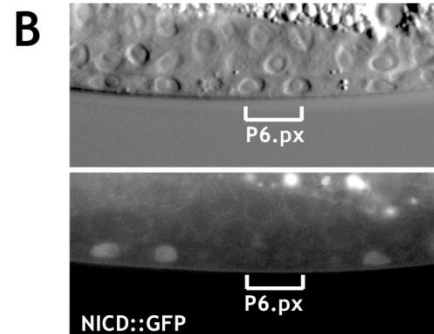
■ vector(RNAi) (n=33)
■ *cyb-3(RNAi)* (n=36)



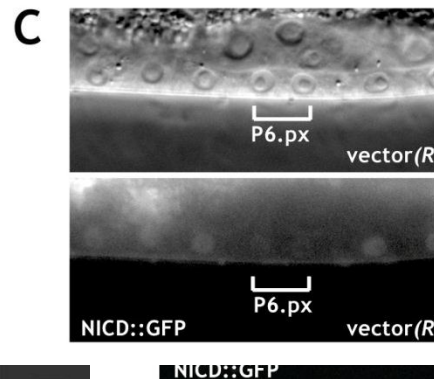
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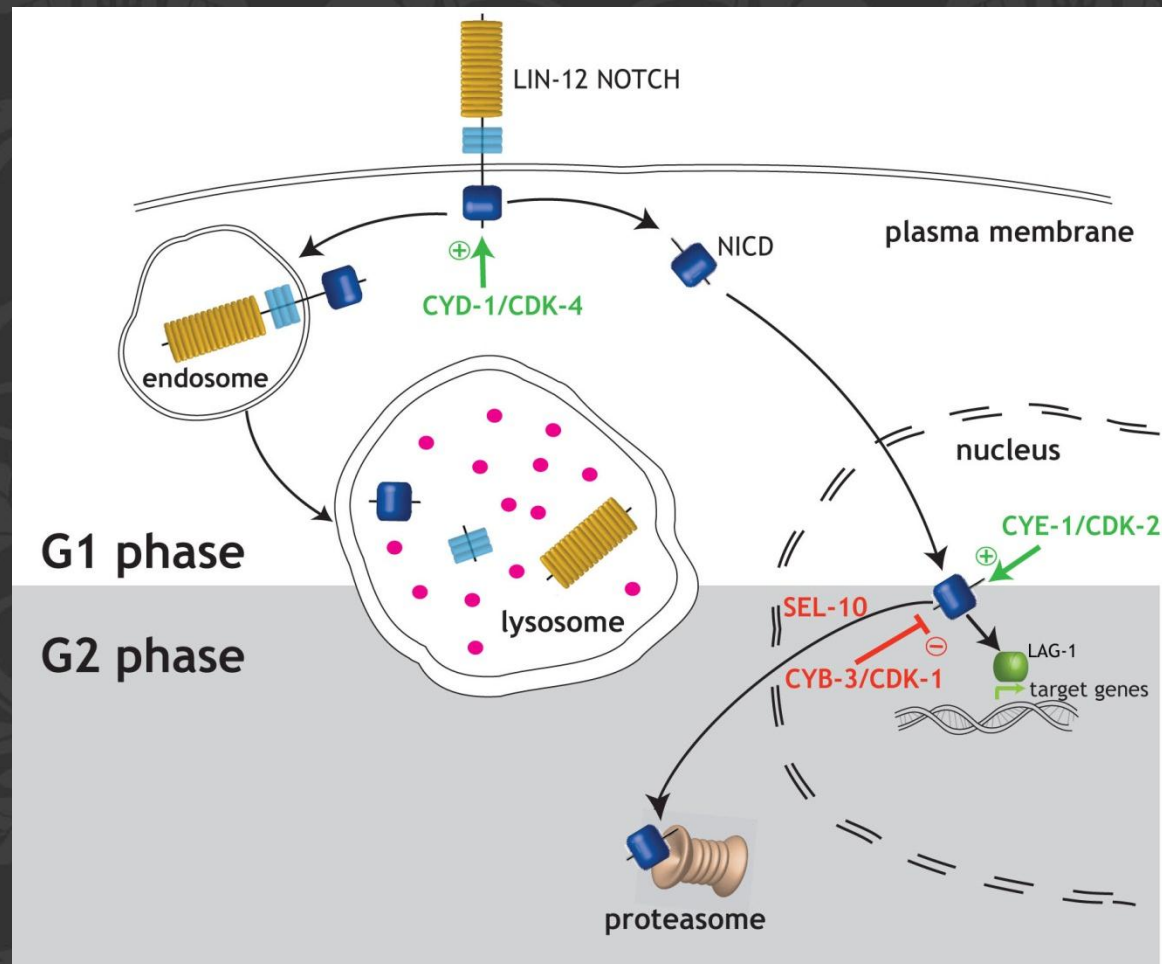
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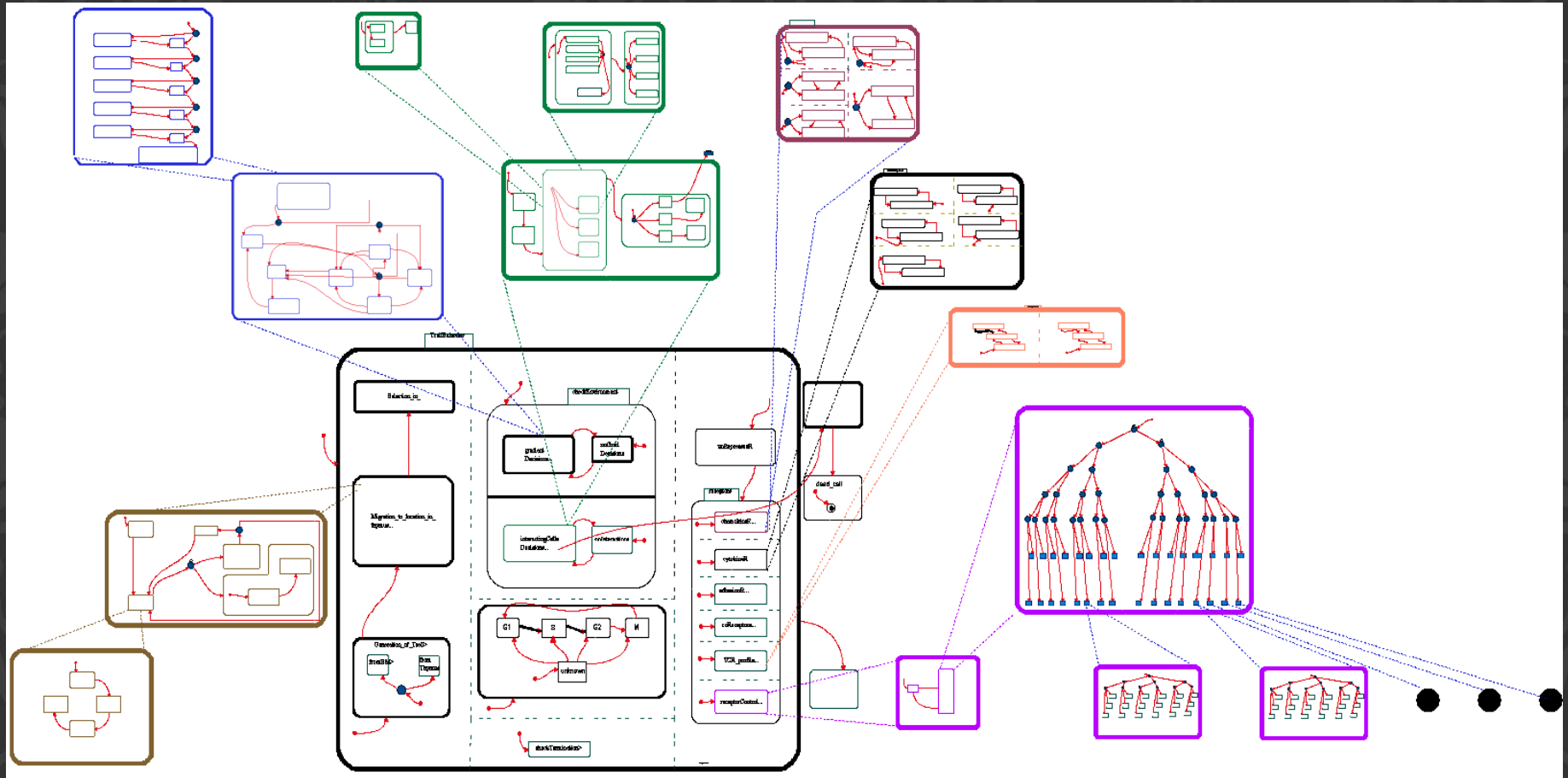
C



Model for Notch Regulation by the Cell Cycle Machinery

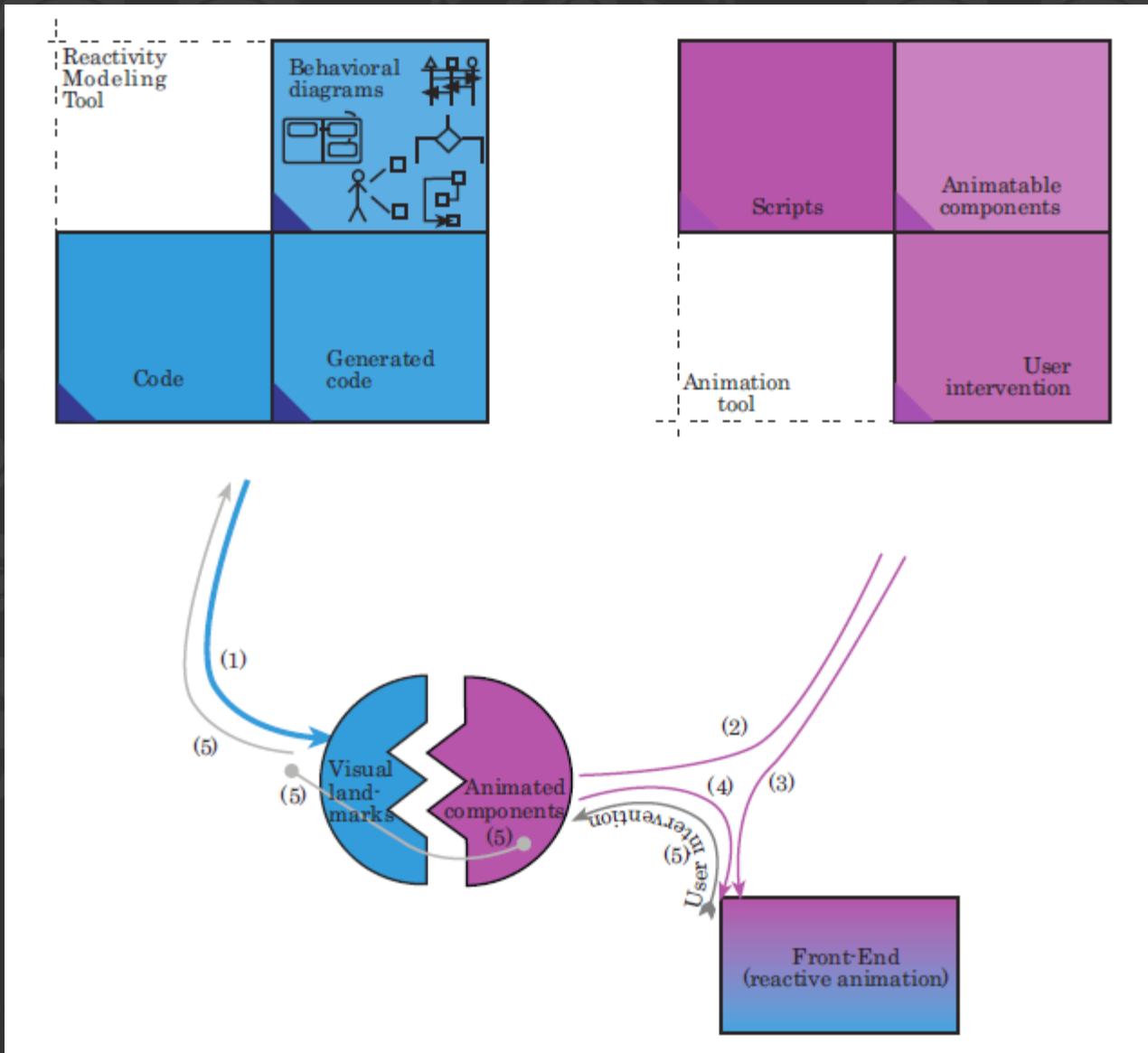


Statechart of a T cell in the thymus

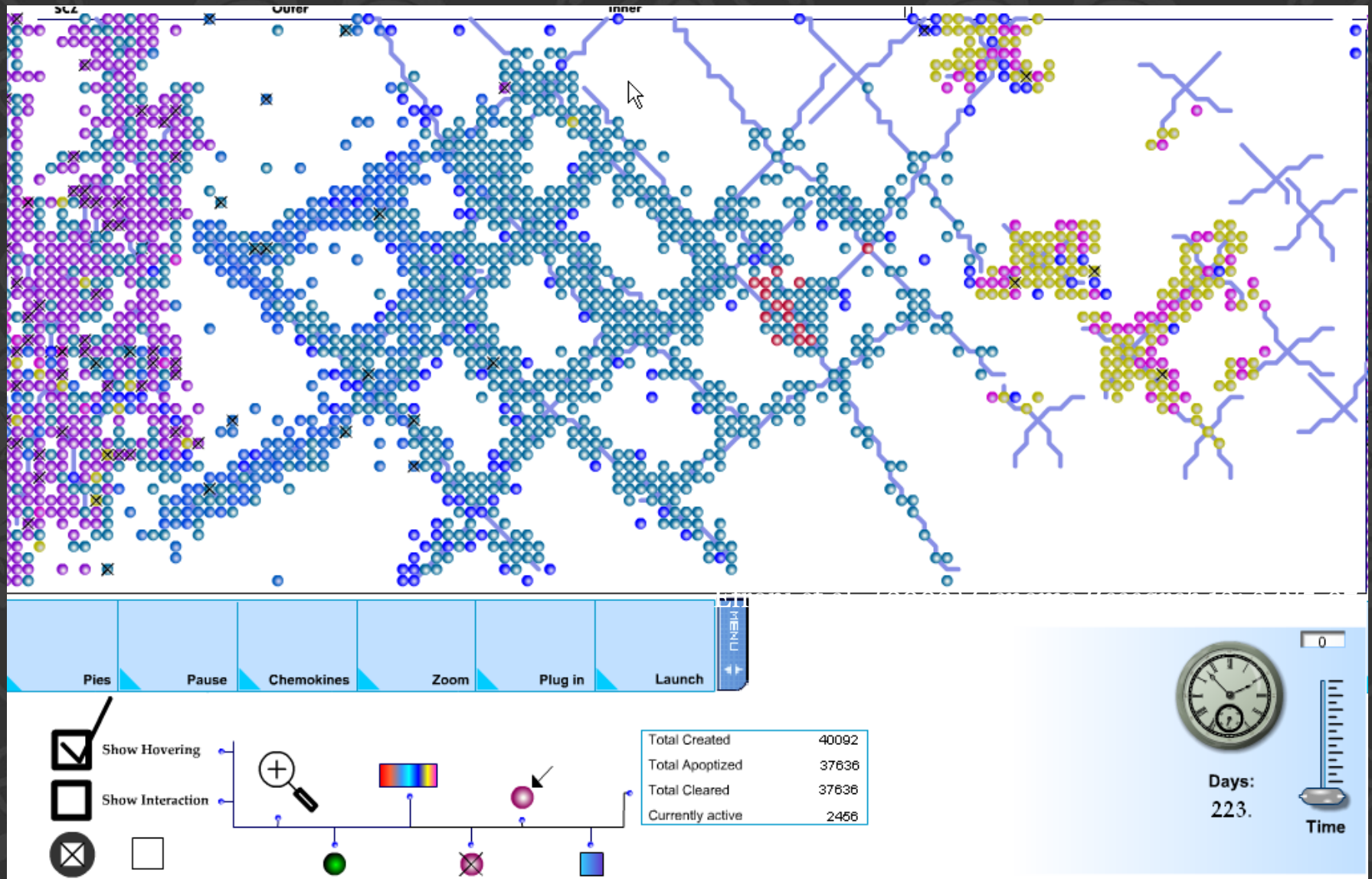


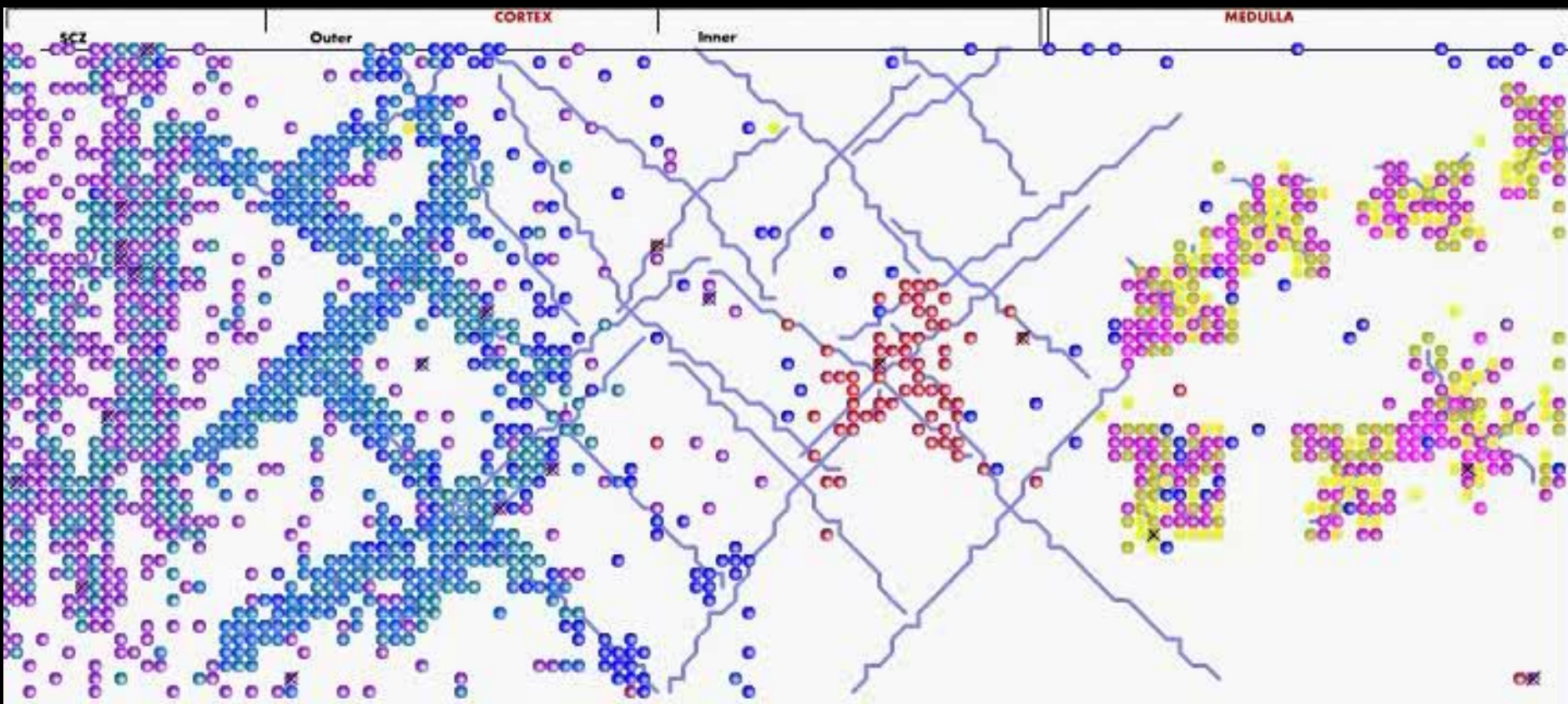
Efroni et al., (2003) *Genome Research* 13: 2485-97

Reactive Animation (Harel, Efroni & Cohen '03)

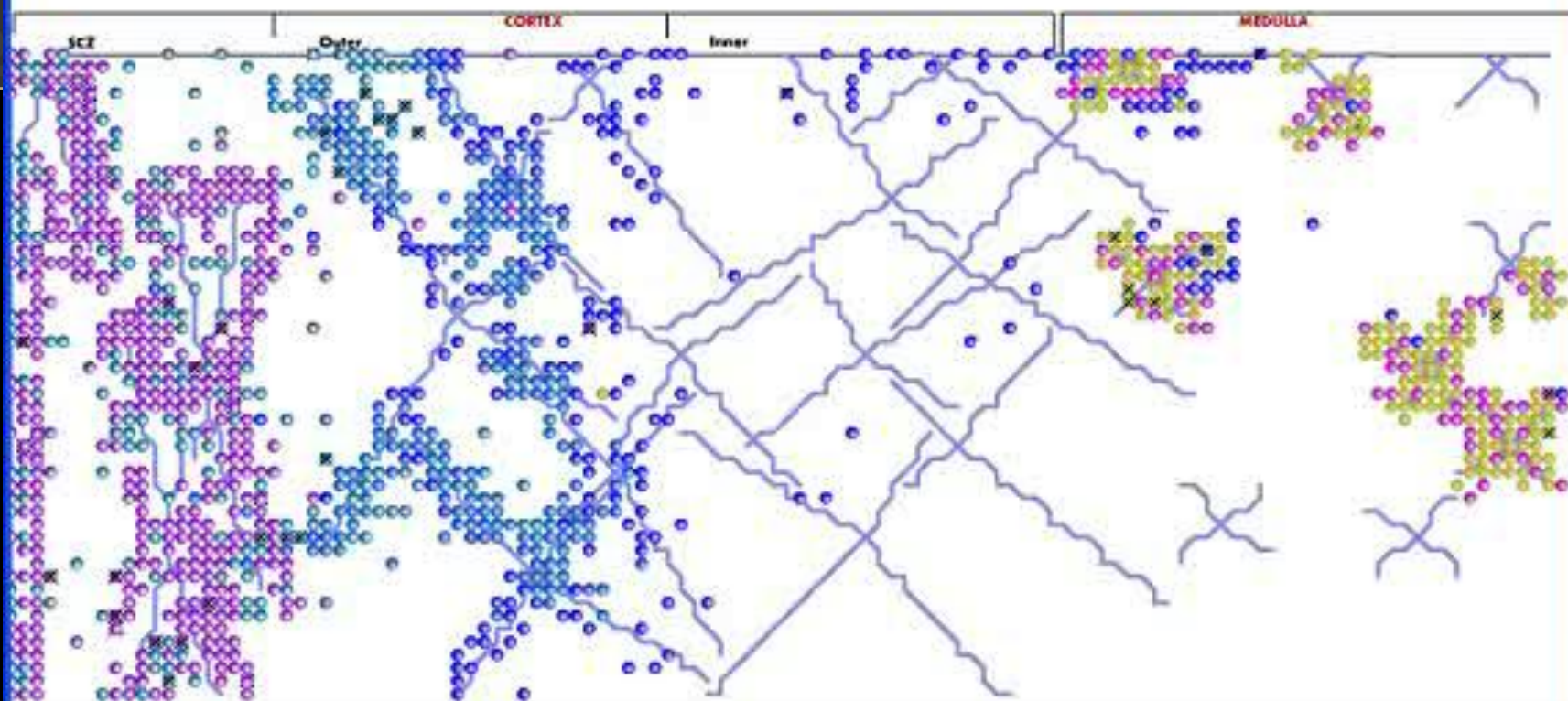


Reactive Animation (Harel, Efroni)





Efroni et al., (2003) *Genome Research* 13: 2485-97



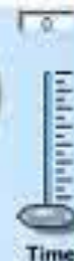
Show Hivering
Show Interaction



Total Created:	4713
Total Apoptosed:	7098
Total Cleared:	7098
Currently active:	3070

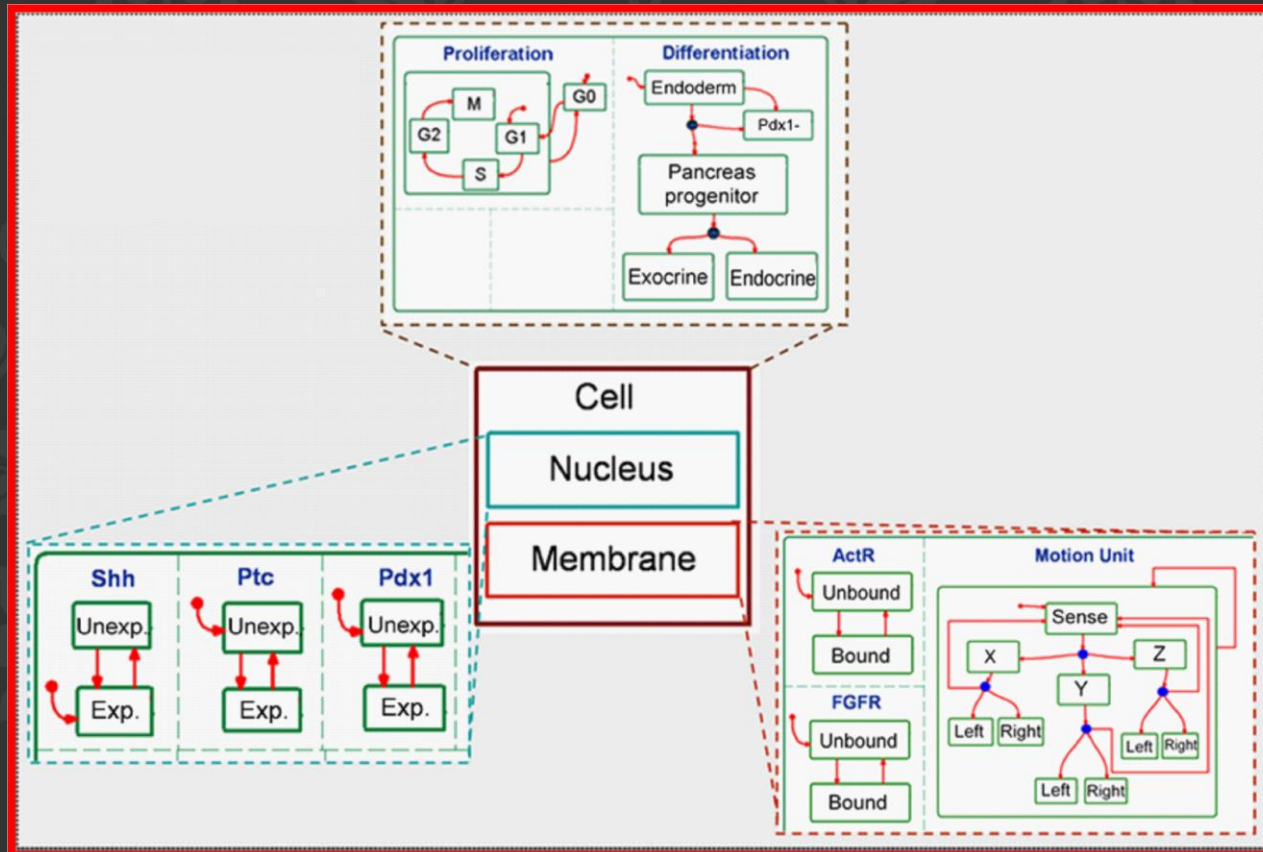


Days:
52.2



Time

Statechart of pancreas (Harel, Setty)



Harel et al., *Electronic Notes in Theoretical Computer Science* 194 (2008)

Setty et al., *PNAS* 105:51 (2008), 20374-20379

Animation Clock: 0 0:31

Number of Cells: 756

Current Day: 8.5



Endoderm: 756
Pancreatic: 0
Early Endocrine: 0
Dead: 0
Endocrine: 0
Exocrine: 0



Delta



[0]

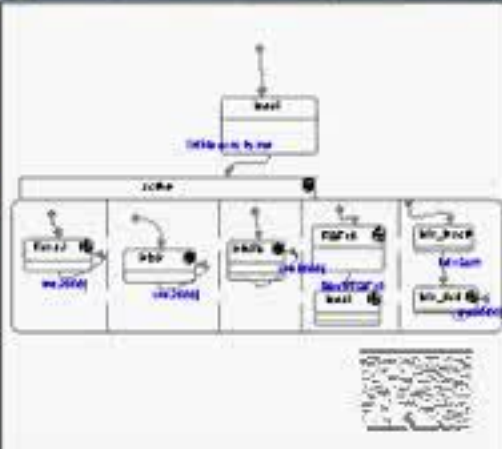
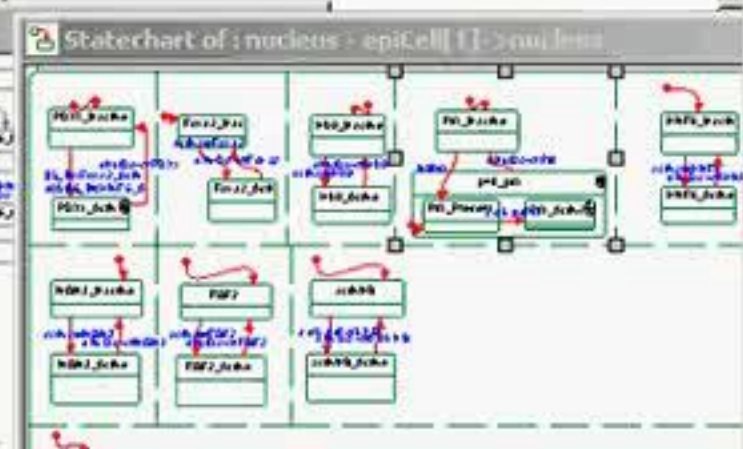
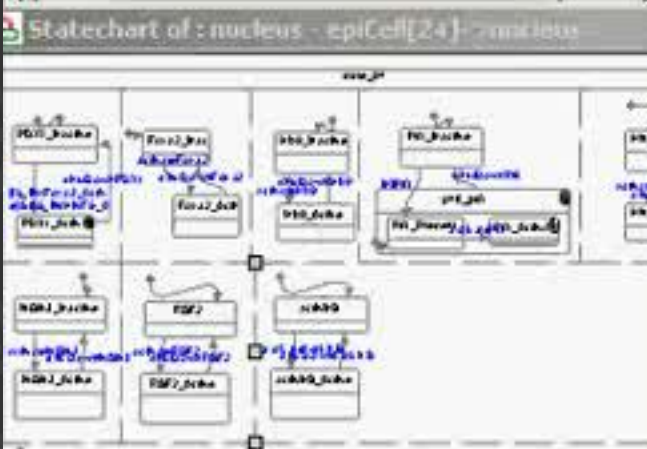
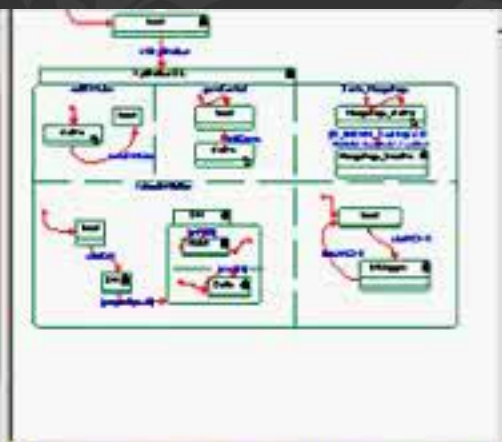
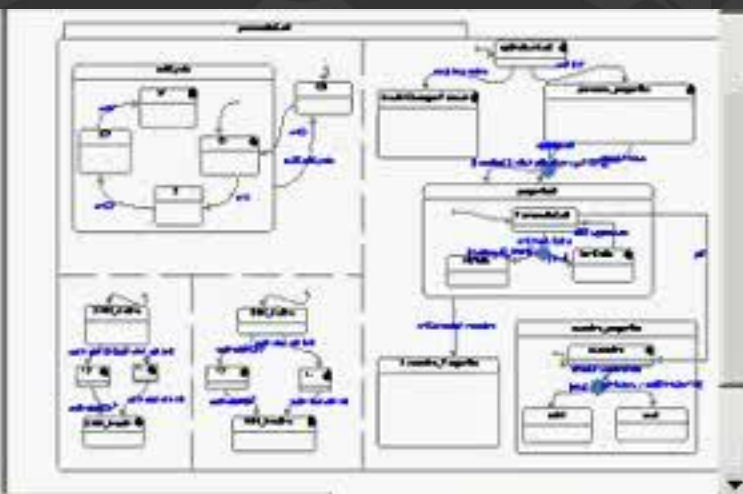
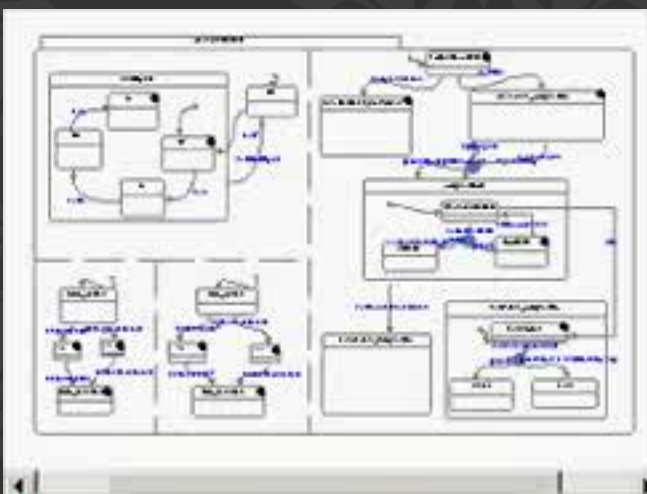
Notch



[0]

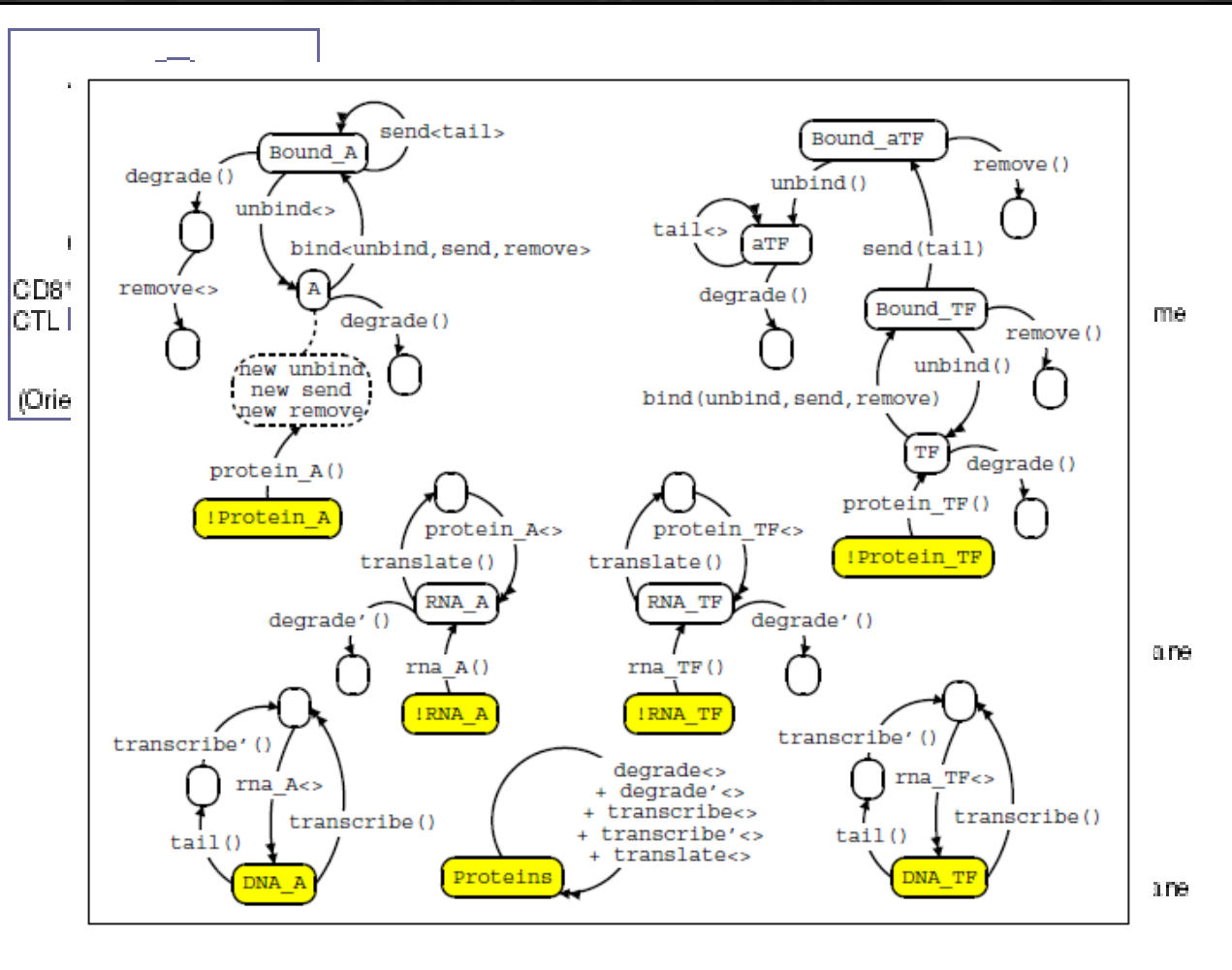
pairwise

neighborhood



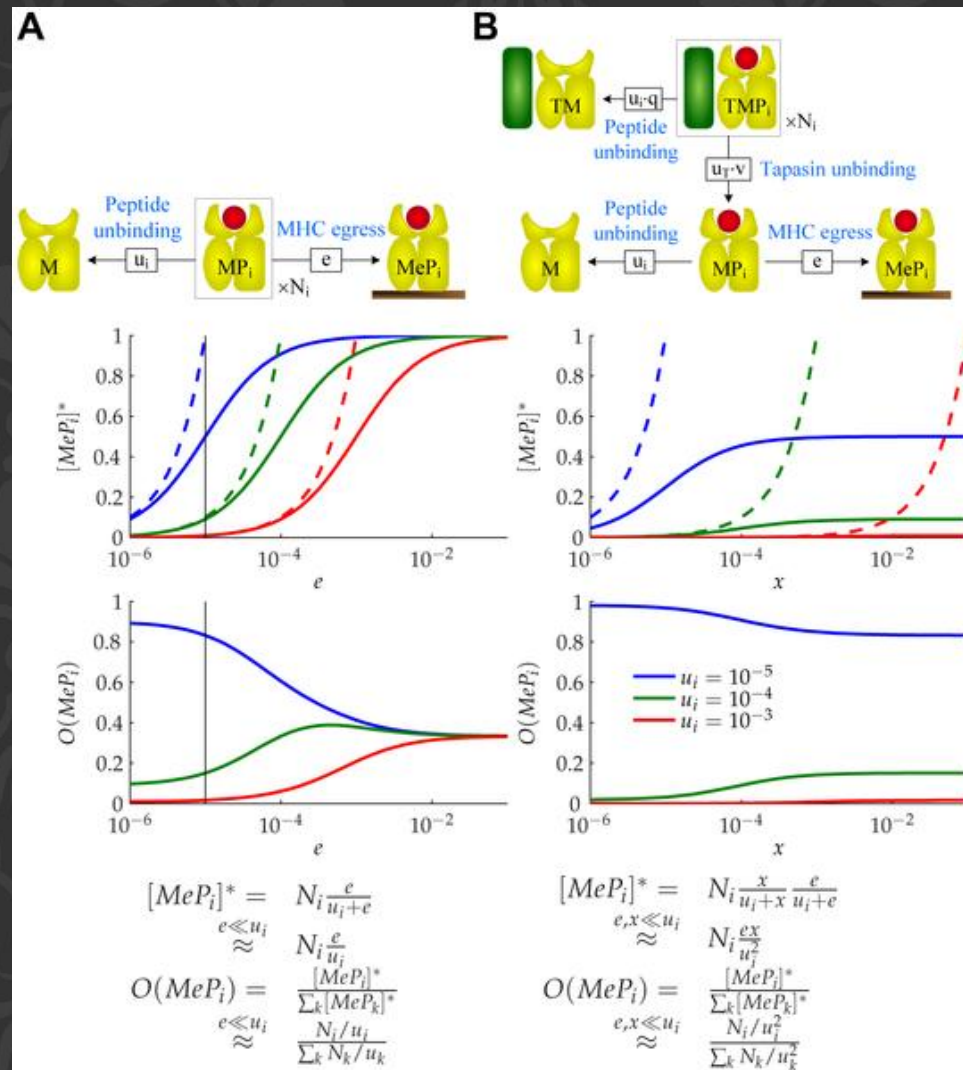
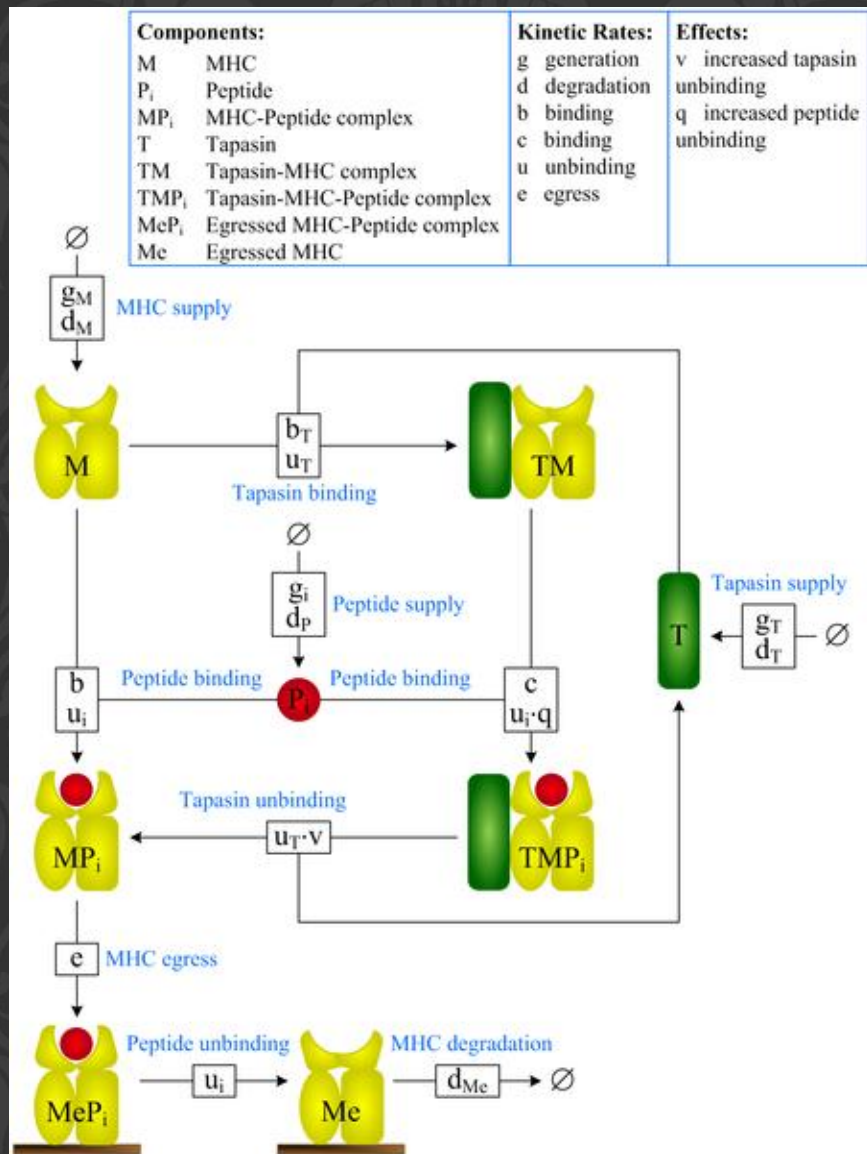
Setty et al., *PNAS* 105:51 (2008), 20374-20379

Modelling biochemical networks using Process Calculi

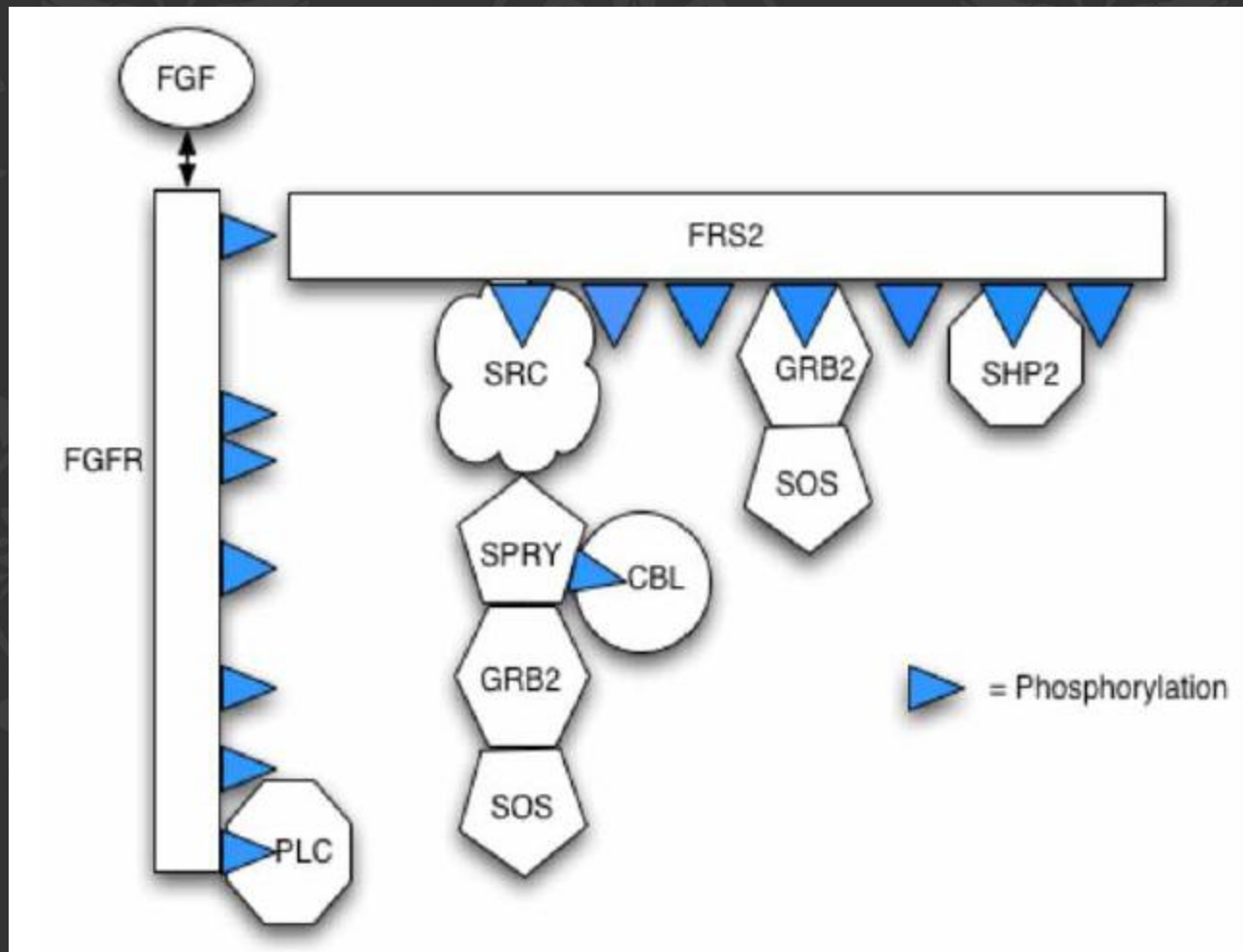


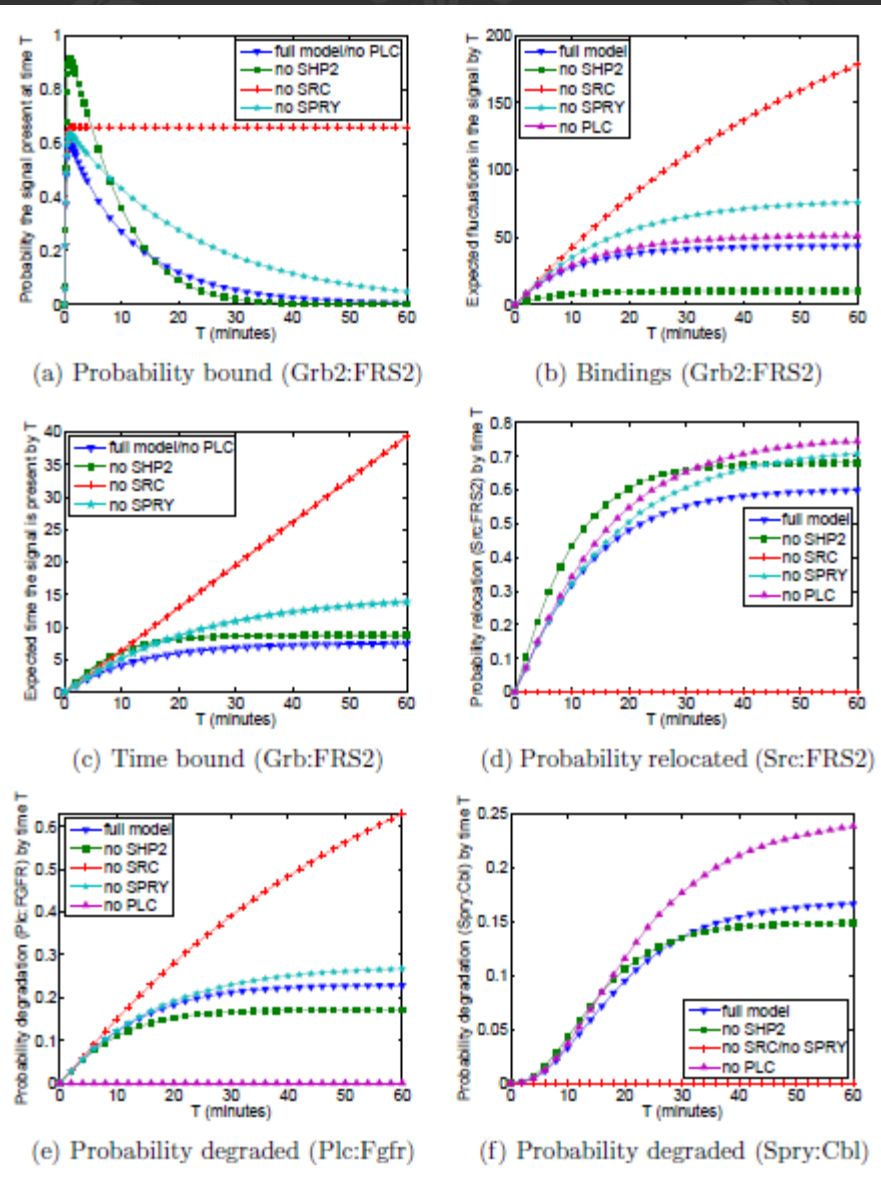
Modelling modulation of immune signalling

Dalchau, Phillips, Cardelli, et al. *PLoS Comp Biol* 7(10): e1002144 (2011)



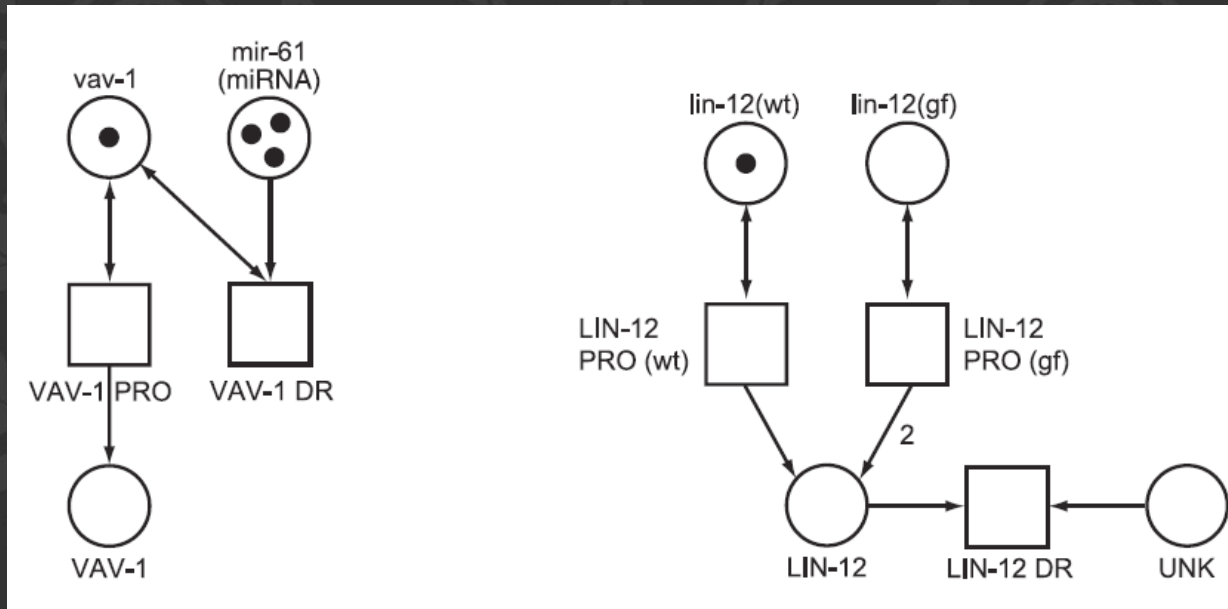
Probabilistic Model Checking of FGF Pathway



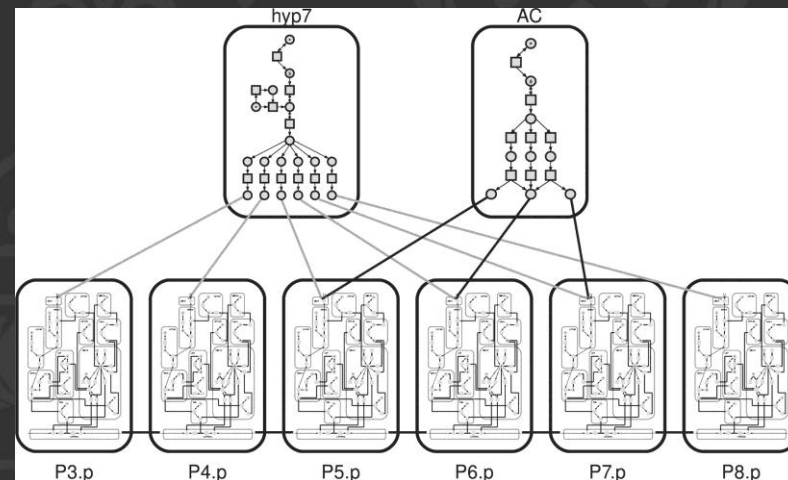
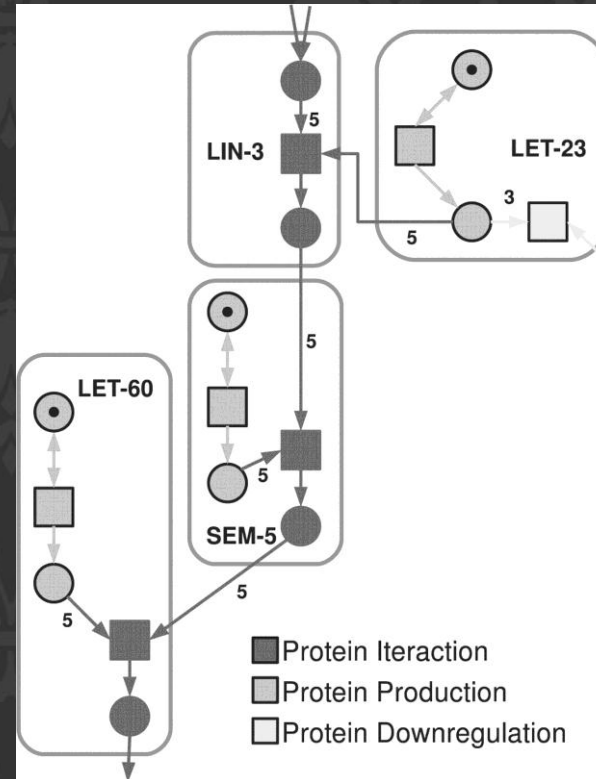
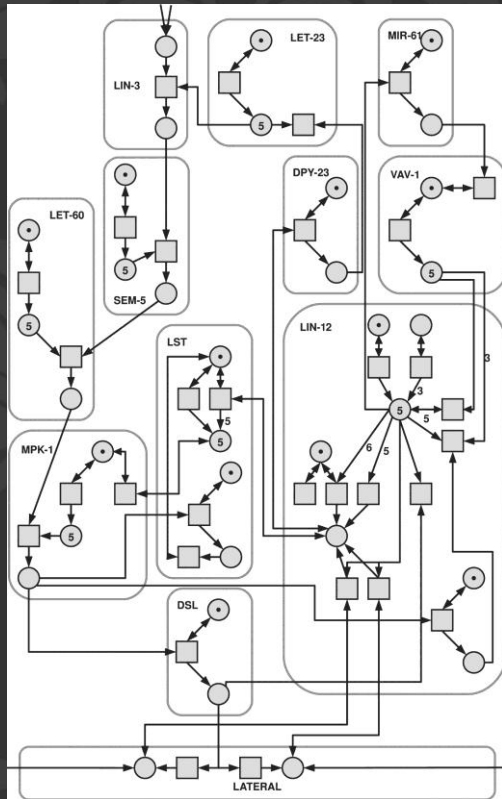
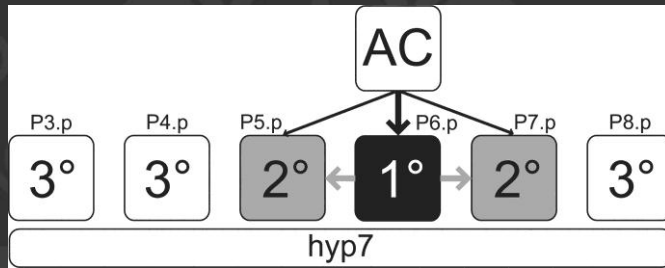


Modelling signal transduction pathways

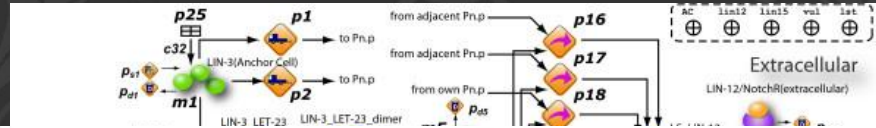
Using Petri Nets



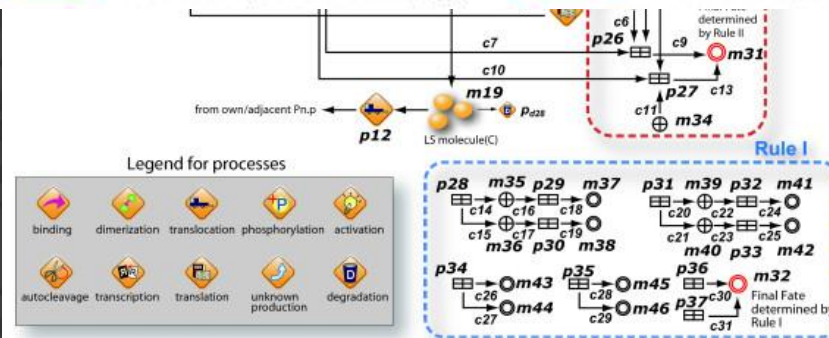
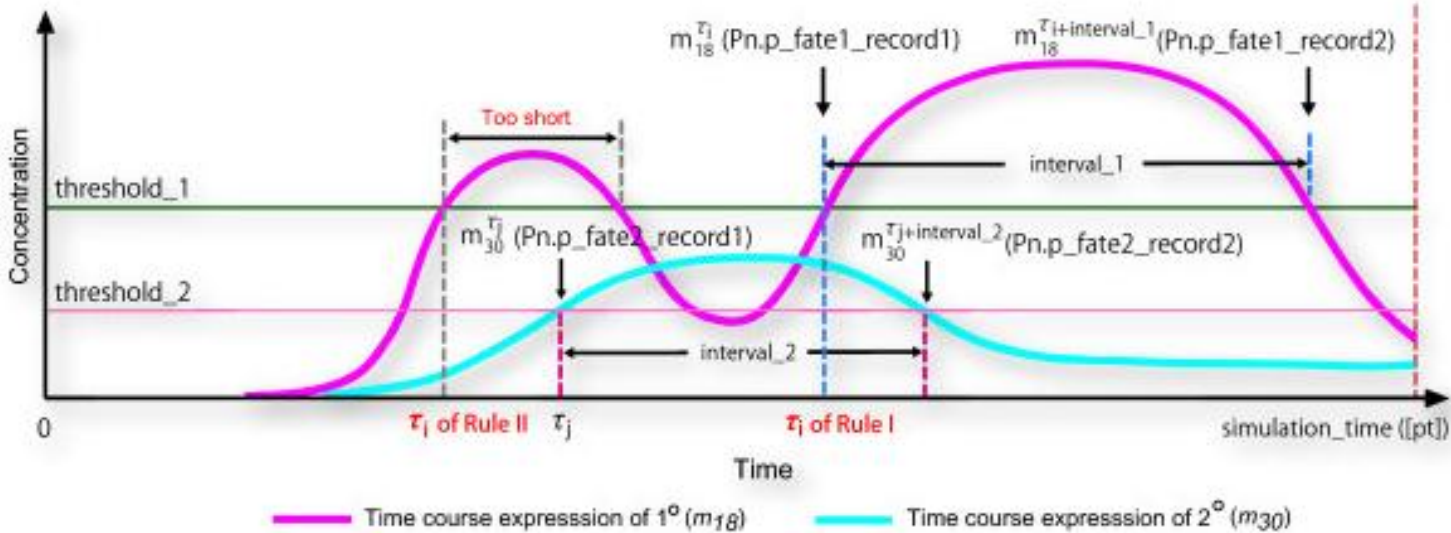
Modelling *C. elegans* vulval development with Petri nets



Modelling *C. elegans* vulval development with hybrid functional Petri nets

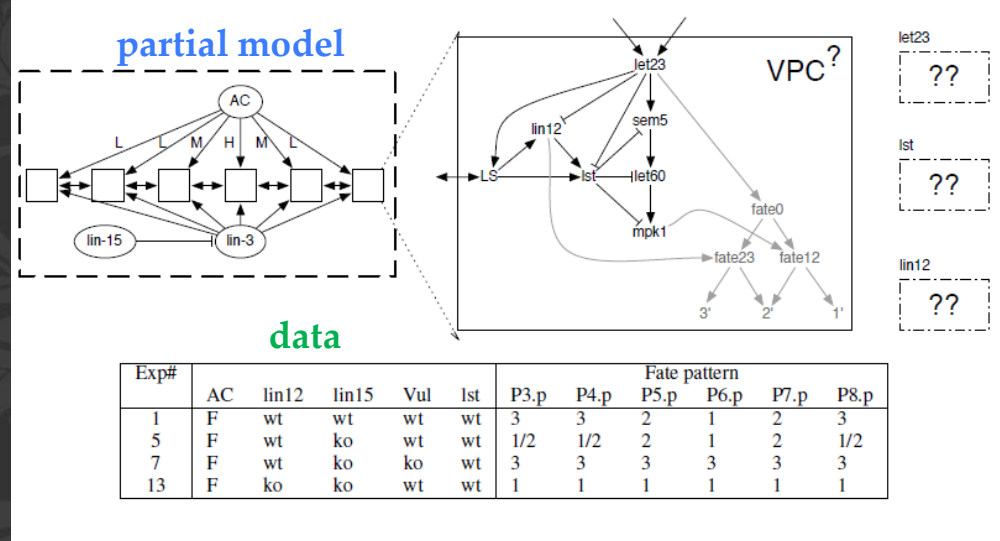
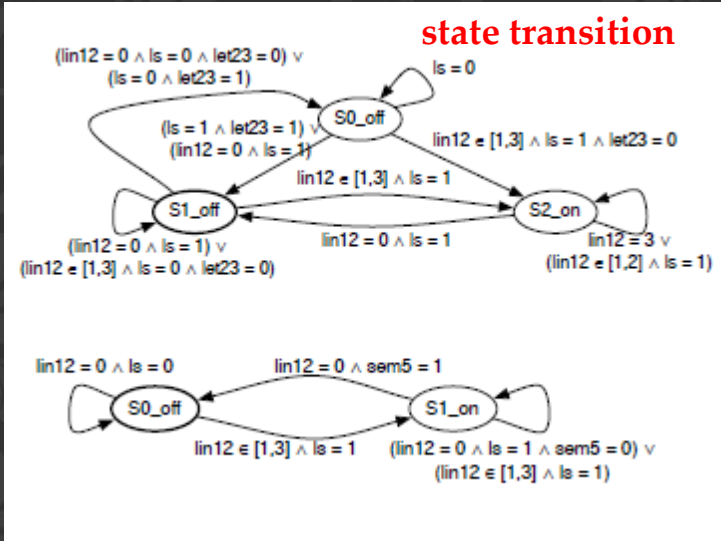


	Cell Illustrator			
	Original elements of HFPNe			Examples of biological images
Type	Discrete	Continuous	Generic	Discrete, Continuous, and Generic



Synthesis of Biological Models from Mutation Experiments

Koksal, Pu, Srivastava, Bodik et al. *POPL* '13



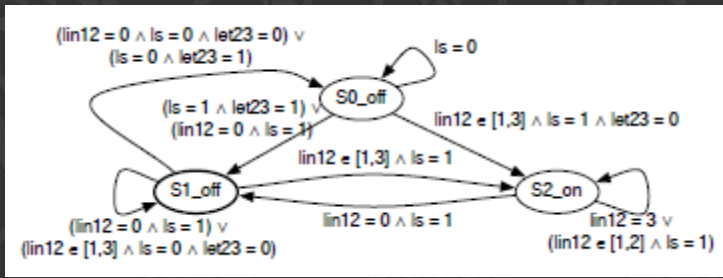
synthesis

Koksal, Pu, Srivastava, Bodik et al. *POPL* '13

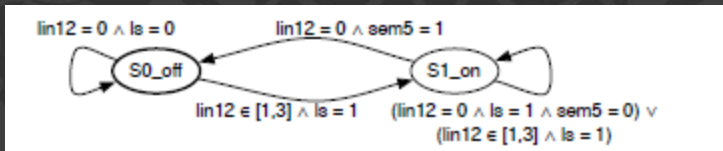
synthesis

synthesis

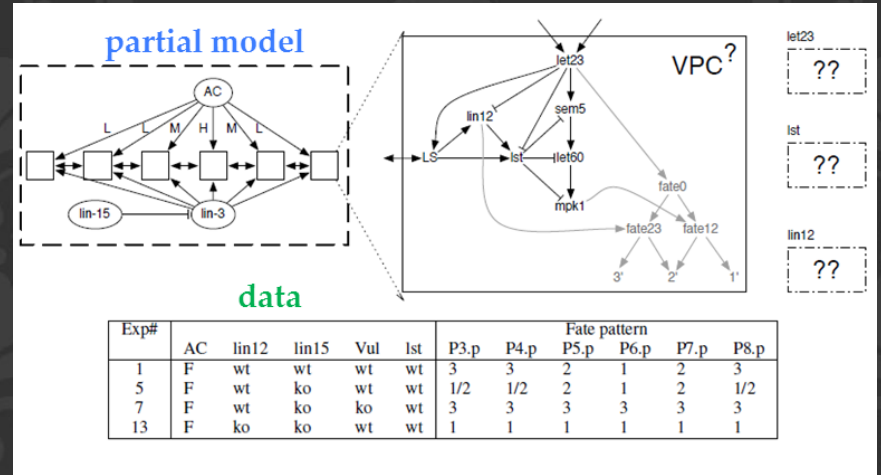
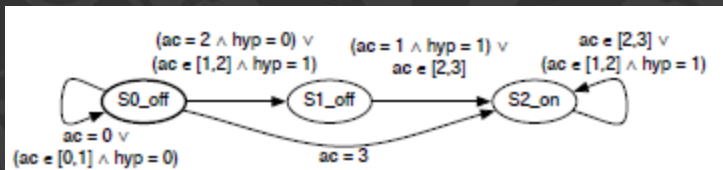
synthesis



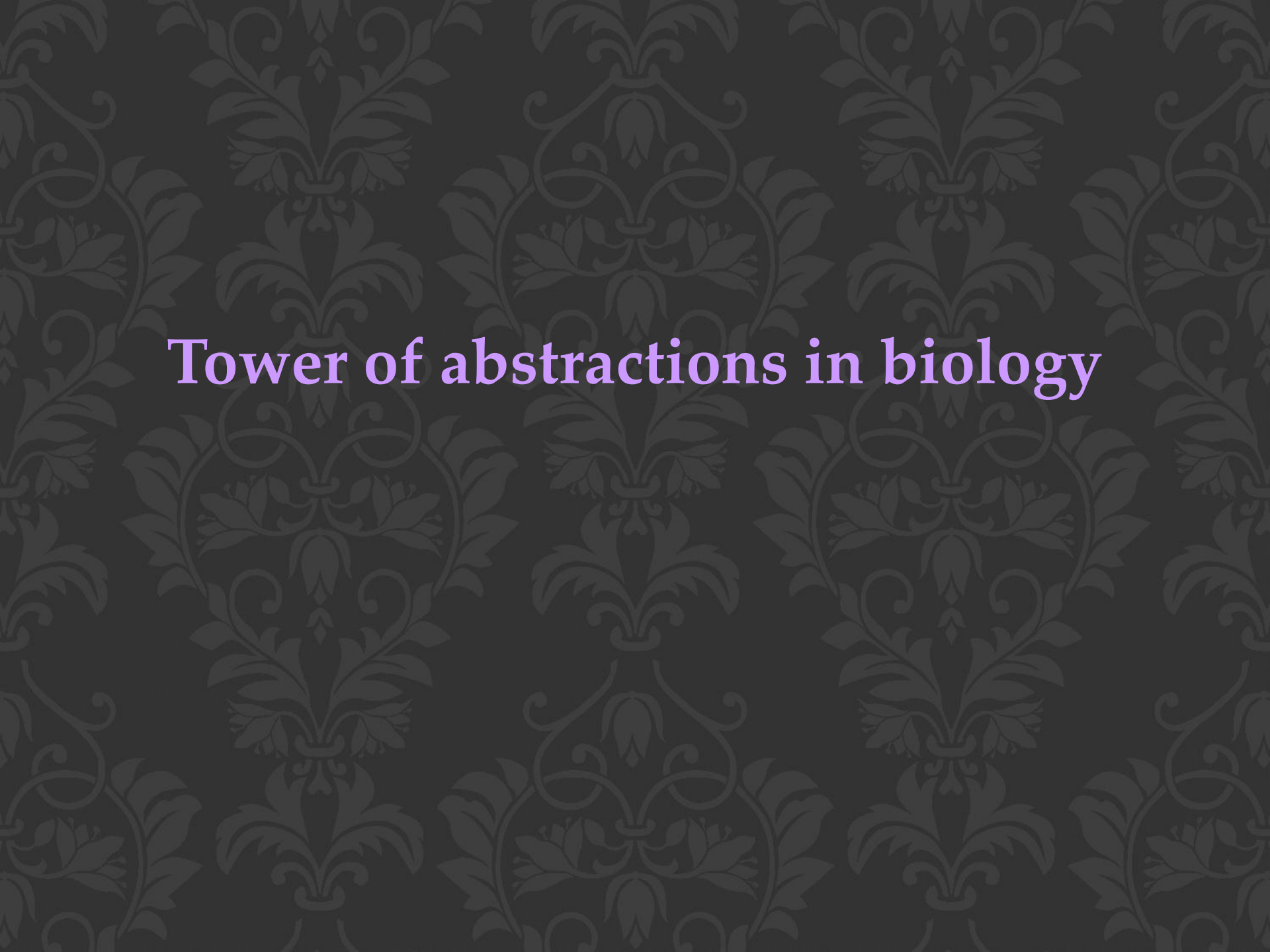
experiments



experiments

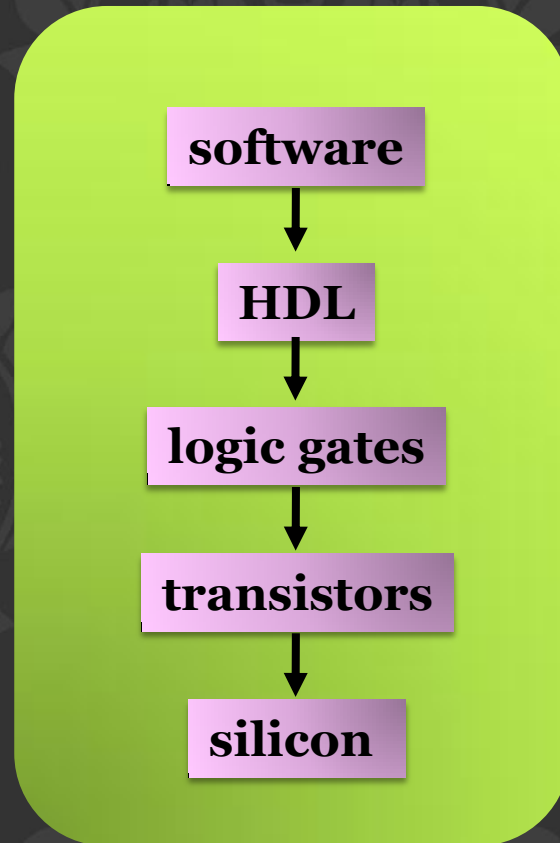


Challenges

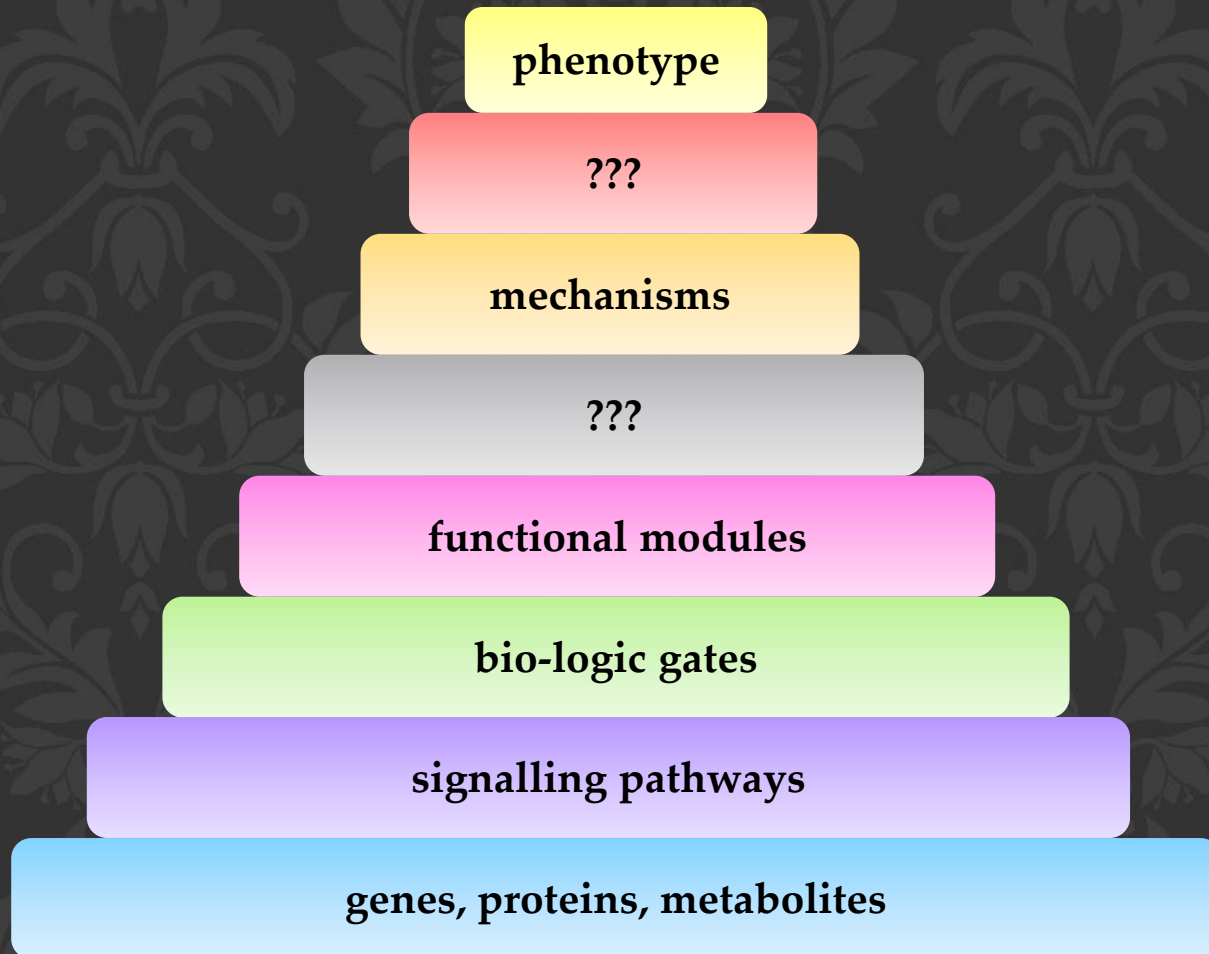


Tower of abstractions in biology

Tower of abstractions in hardware



Tower of abstractions in biology





User-friendly modelling

Executable Biology

```
graph TD; EB[Executable Biology] --> BQ((biological questions)); EB --> BN((biological 'needs' )); BQ --> ExpB[Experimental Biology]; BN --> CS[Computer Science]; ExpB <--> CS;
```

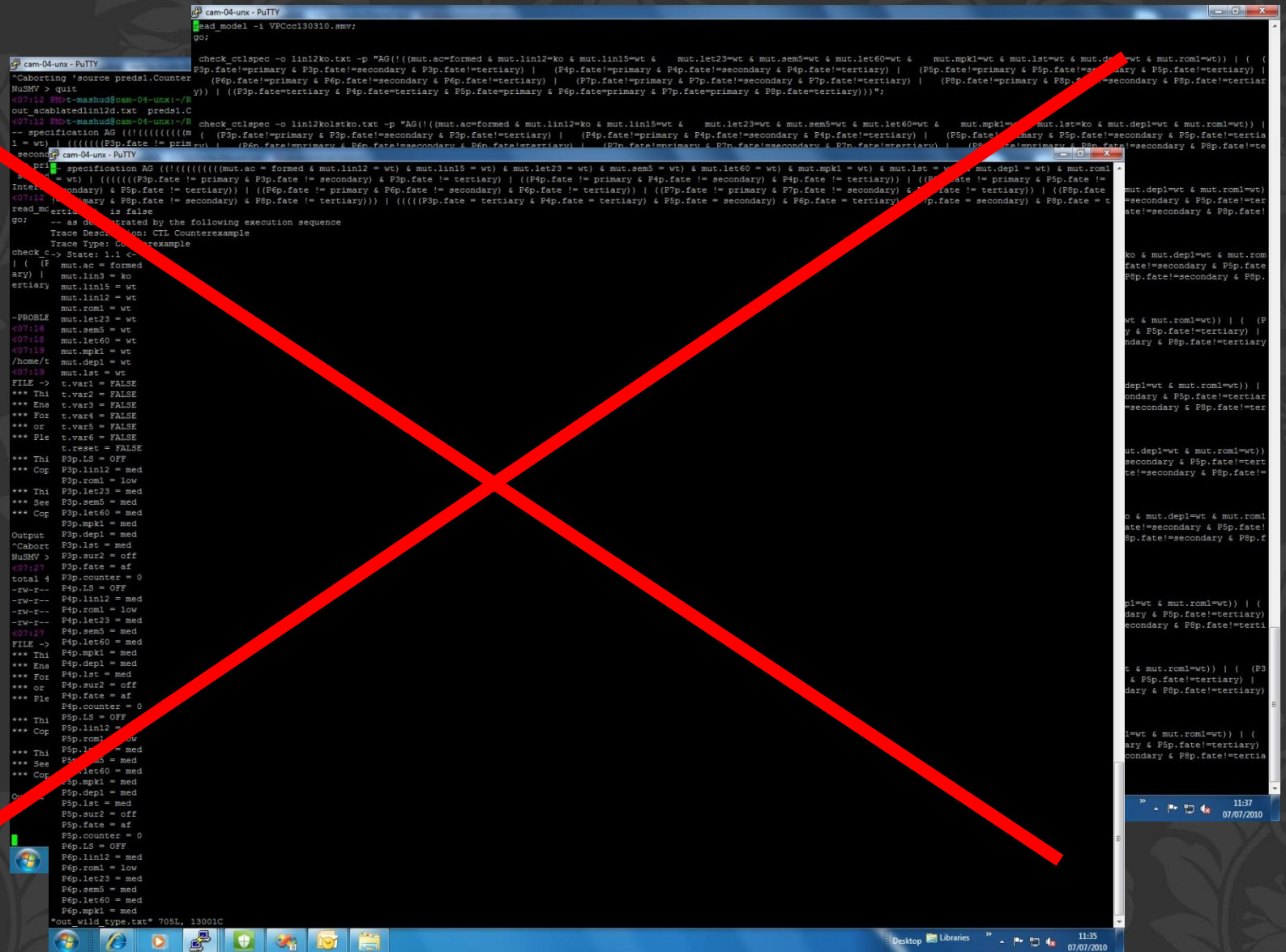
The diagram illustrates the workflow of Executable Biology. At the top, a pink banner labeled 'Executable Biology' has two arrows pointing down to 'biological questions' (a green oval) and 'biological 'needs'' (a blue oval). From 'biological questions', a green arrow points to 'Experimental Biology' (a green hatched rectangle). From 'biological 'needs'', a blue arrow points to 'Computer Science' (a blue dotted rectangle). At the bottom, a green arrow points from 'Experimental Biology' to 'Computer Science', and a blue arrow points from 'Computer Science' back to 'Experimental Biology', indicating a bidirectional relationship.

biological
questions

biological
'needs'

Experimental Biology

Computer Science



Not talking about GUI...

Mocha: Exploiting Modularity in Model Checking

File Edit Simulate Check Options

Project: main.rm

- types
- modules
- invariants
 - wild_type
 - sanity
- judgments
 - ☒ check_sanity
 - ☒ check_wild_type

Simulation Result

main_counter = 12
main_counter = 12
mpk1_state = med2_
main_state = tertiary
mpk1_state = low_i
main_state = secondary
mpk1_state = low_i
lin3 = med
dep1_state = med_i
l1t_state = med_i
sur2_state = off_i
main_counter = 12
let23_state = med4_i
l1t_state = high1_i
mpk1_state = med2_
let23_state = med4_i
main_state = secondary
mpk1_state = high_i
let23_state = high_i
main_state = primary
main_counter = 12
sur2_state = off_i
main_state = secondary
l1t_state = high1_i
main_state = secondary
sur2_state = off_i
thereset = off
thereset = on
thereset = off

Objects Files

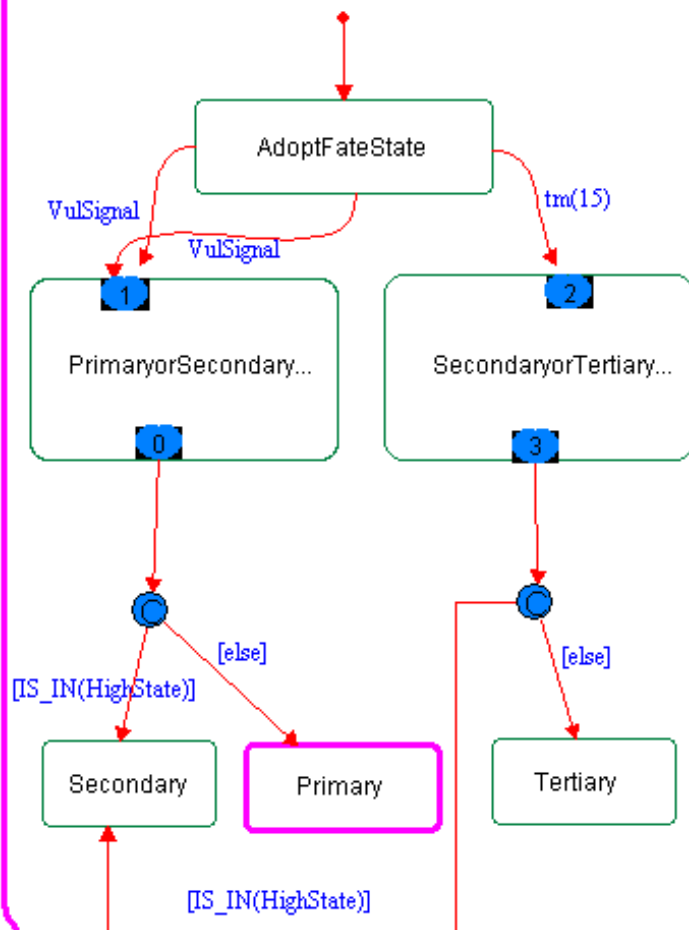
main.rm Check Result: check_wild_type Simulation Result

Invariant disproved.

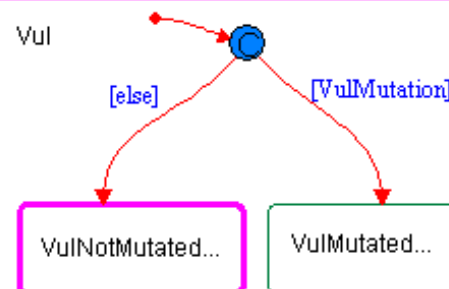
Type checking...

notAblated>

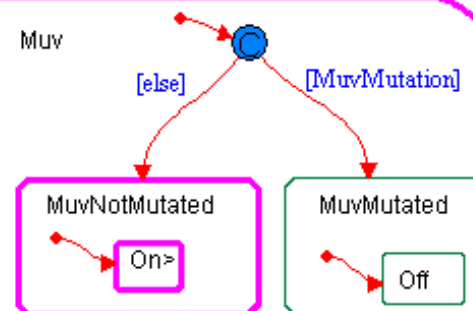
Main



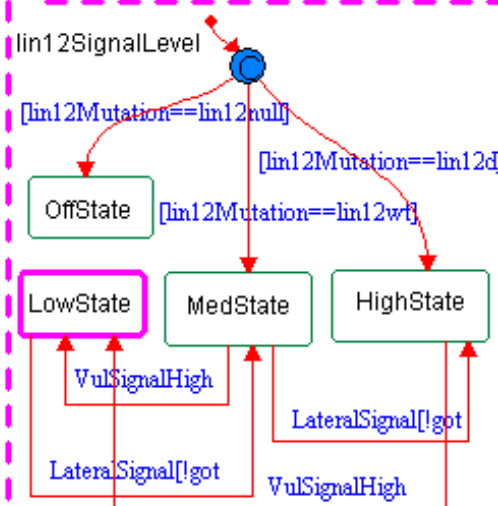
Vul



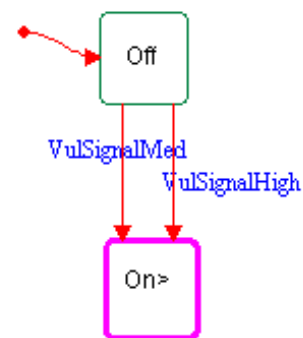
Muv



lin12SignalLevel



LateralSignalState



Bio Model Analyzer

Proving Stabilization of Biological Systems



Byron Cook
MSRC



Alex Taylor
MSRC



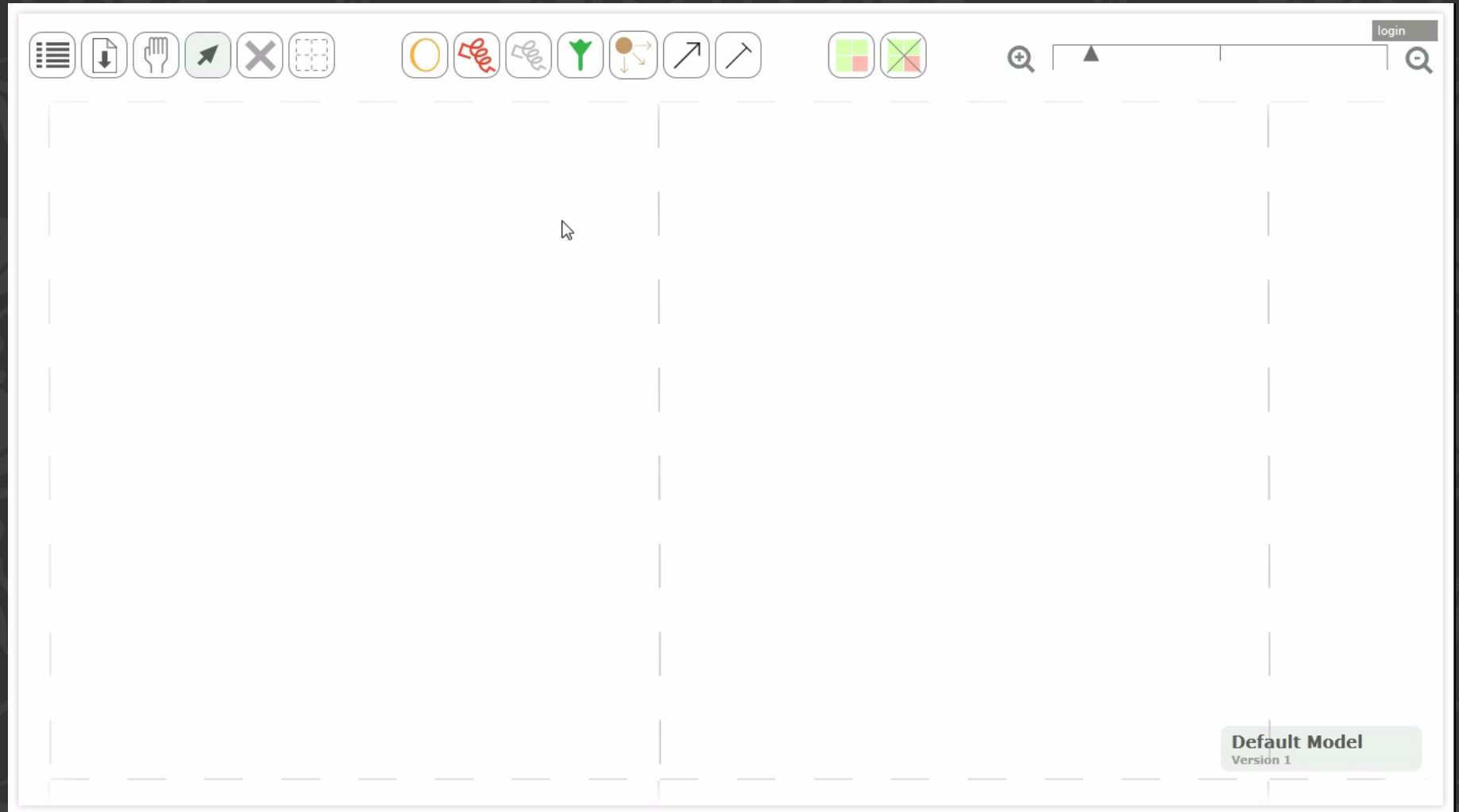
Samin Ishtiaq
MSRC



Nir Piterman
Univ. of Leicester

VMCAI 2011, CAV 2012

<http://biomodelanalyzer.research.microsoft.com/>



Proof visualization

The screenshot displays a software interface for proof visualization. At the top, there is a toolbar with icons for file operations (list, download, close, zoom, grid), drawing tools (circle, line, arrow, fork, arrow, hammer), and a color palette (green, red, green, red). The main workspace contains a network diagram with nodes and directed edges. The nodes are: **ConstA 4** (green), **Roc1 1** (green), **Roc2 1** (green), **VarA 0** (green), **VarB 2** (green), **VarC 1** (red), **VarD 1** (grey), **VarE 1** (grey), and **ProtF 1** (grey). A large orange oval encircles the nodes **Roc1 1**, **Roc2 1**, **VarA 0**, and **VarB 2**. The edges represent interactions: **ConstA 4** to **Roc1 1** and **Roc2 1**; **Roc1 1** to **VarA 0** and **VarB 2**; **Roc2 1** to **VarA 0**; **VarA 0** to **VarC 1**; **VarB 2** to **VarC 1**; **VarC 1** to **VarD 1**; **VarD 1** to **VarE 1**; and **VarE 1** to **ProtF 1**. On the right side, a **PROOF REPORT** window is open, displaying the following text:

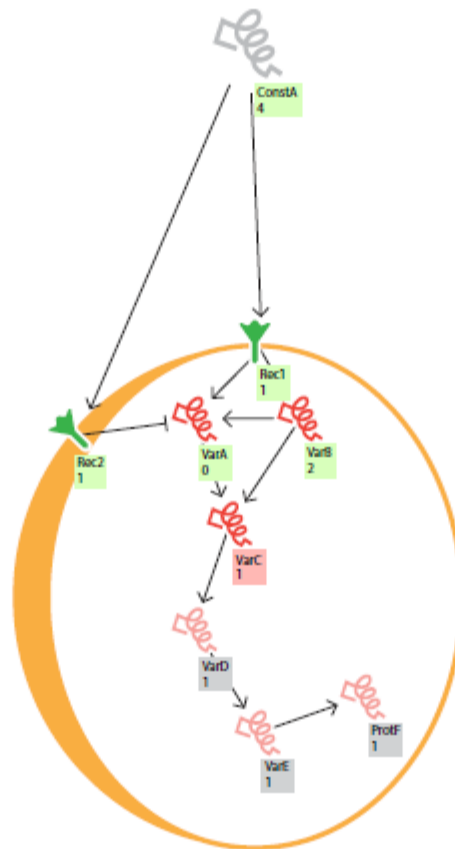
Analysis
[model name] failed to stabilize.
After stepping through [number of steps] separate interactions in the model, the analysis eventually timed out (after [time] seconds) and failed to determine a final stable state.

Results
The number of steps taken in the analysis and [time] second time out suggests this model may require [minor/ major] modifications before stabilization is achieved.

[list variables] were identified as key in the model's failure to stabilize. A change to these variables or their connections could have an impact on the progression of the analysis and, potentially, the results could be impacted if ranges for [list variables] were [narrowed/adjusted], activations were [introduced/removed] between [list pairings of variables], or inhibitions were [introduced/removed] between [list pairings of variables].

Although the model failed to stabilize, [two] potentially helpful counter examples were identified.

At the bottom of the interface, there is a footer with the **BMA** logo, a link to **About - Help - Terms of Use**, and the **Microsoft Research** logo.



PROOF REPORT



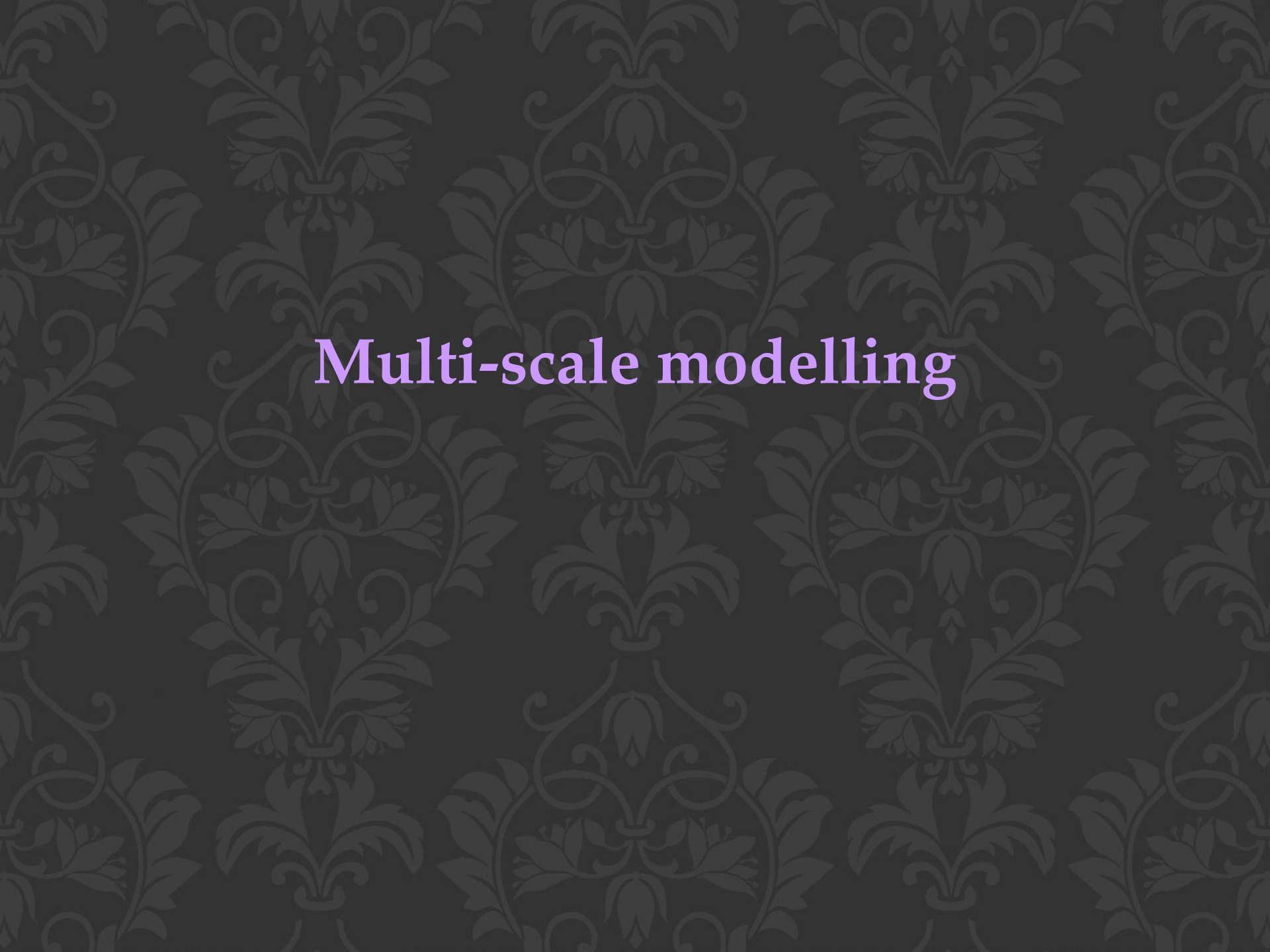
Results Table

	EXP. 1	EXP. 2	EXP. 3	EXP. 4
var A	0-1	2		
var B	3-5			
var C	4	4-5		
var D	0-n	n		

Proof Progression

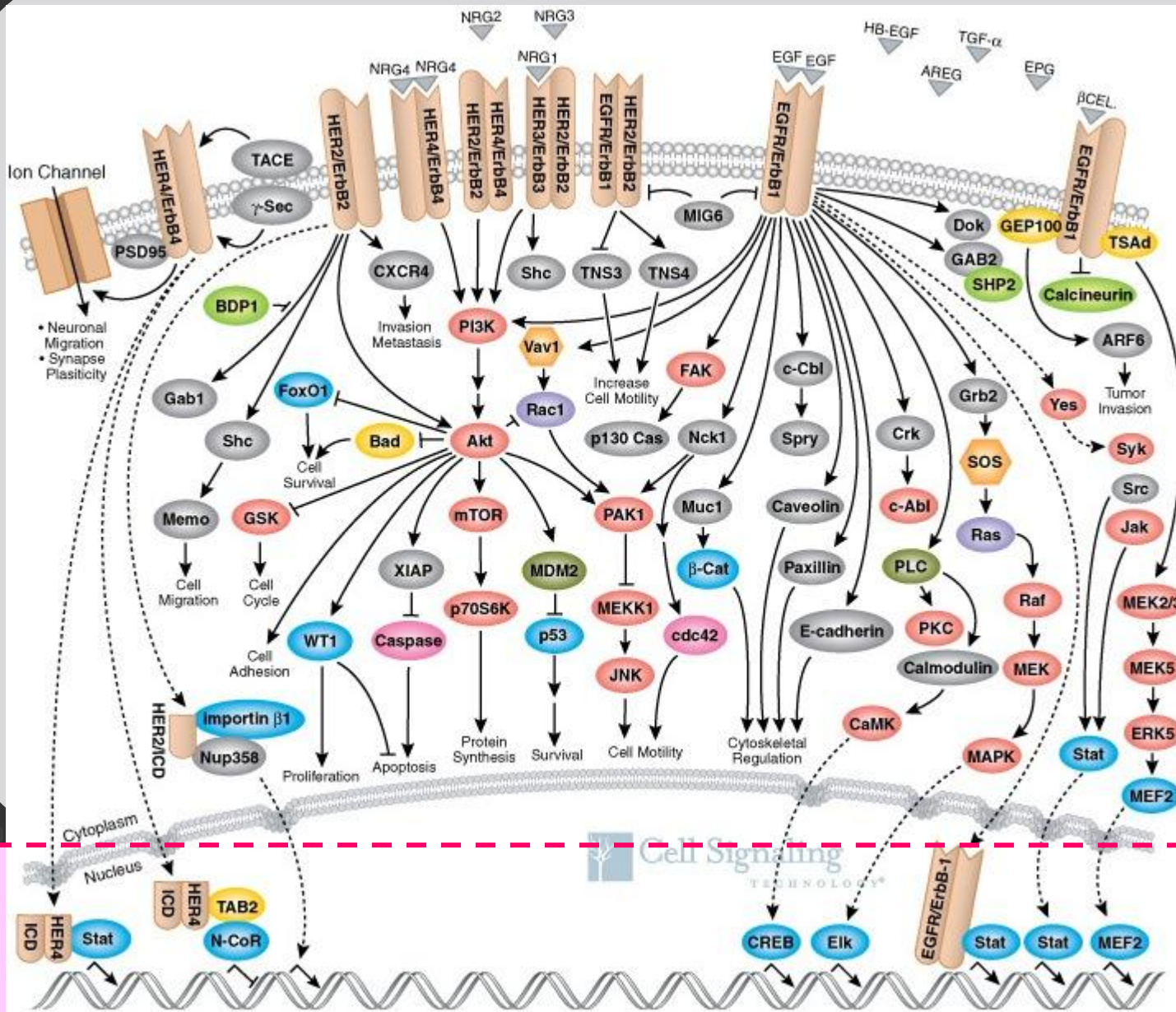
	T=1	T=2	T=3	T=4
var A	1-3	2	2	2
var B	0-4	0-3	0-1	0
var C	0-4	0-2	0-1	1
var D	0-2	1-2	2	2
var N	1-3	2	2	2





Multi-scale modelling

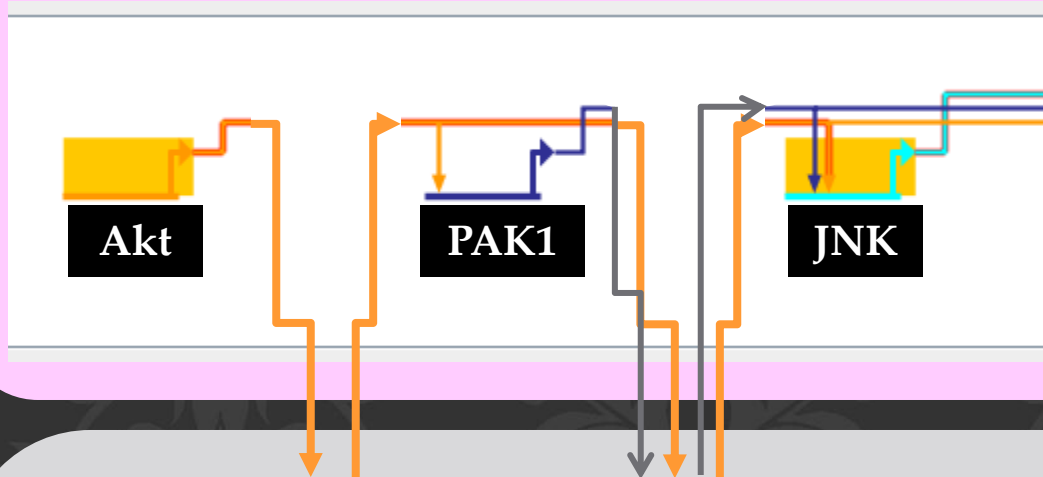
Different time scales and space



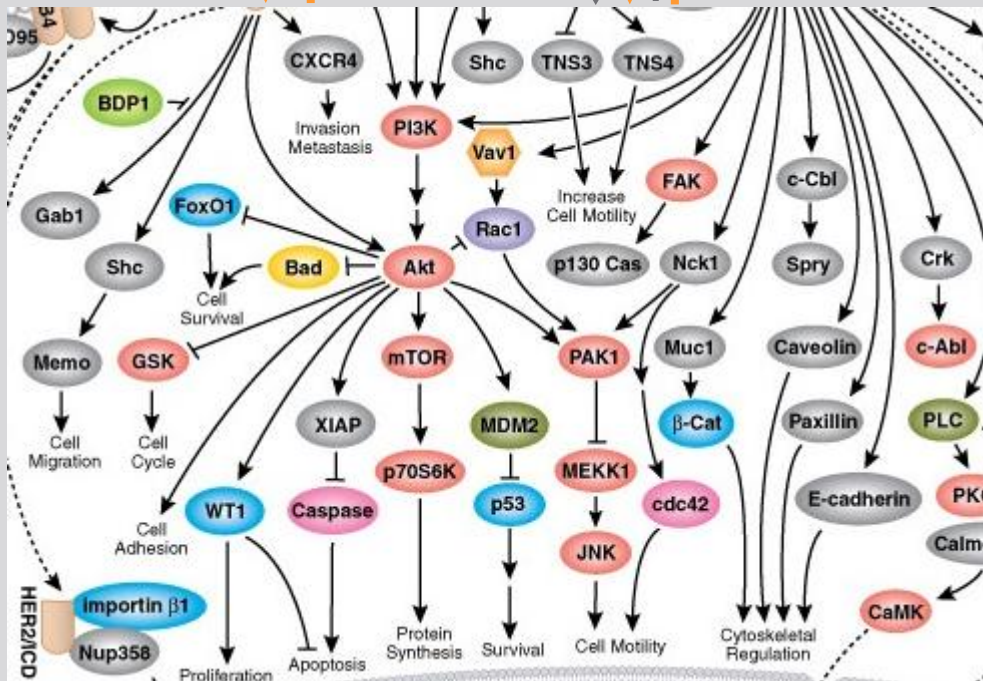
Cell Signalling

Gene Regulation

Feedback between the levels

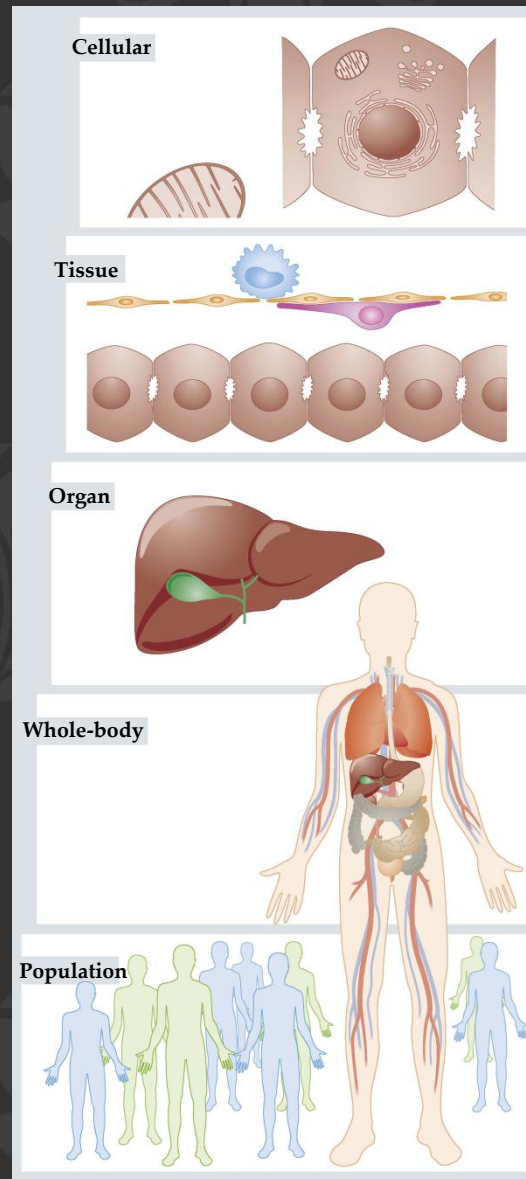


Gene Regulation



Cell Signalling

A multi-scale presentation of human physiology



BIOLOGY
“ROCKS”

Thanks!...