



# From microbes to microbiota and back:

Using thousands of genomes to understand thousands of metagenomes

Curtis Huttenhower

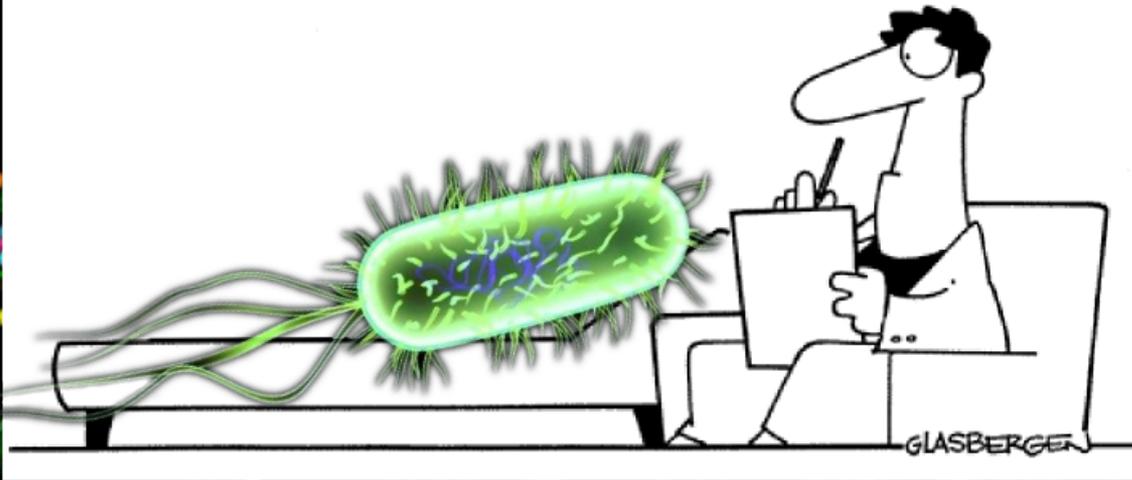


Harvard School of Public Health  
Department of Biostatistics



02-16-13

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"Wait! Wait! Listen to me! ... We don't have to be just commensal microbes!"

# Sequencing as a tool for microbial community analysis

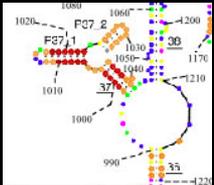


Lyse cells  
Extract DNA (and/or RNA)

**16S amplicons**

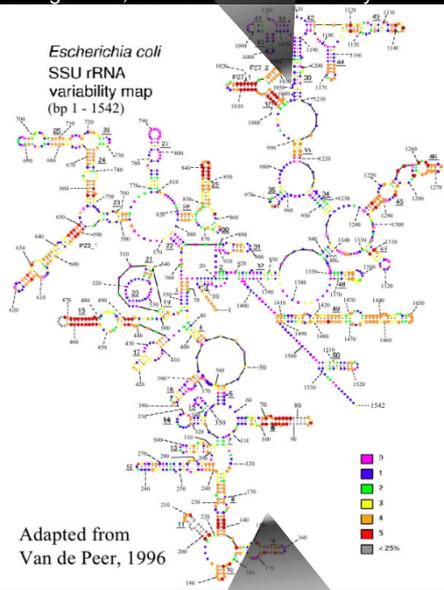
**Meta'omic**

V6

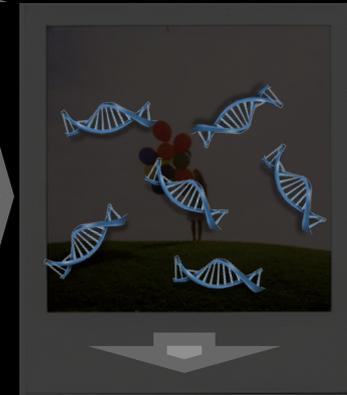


George Rice, Montana State University

PCR to amplify the single  
16S rRNA marker gene

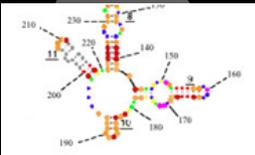


Hello  
my name is  
Classify sequence  
→ microbe

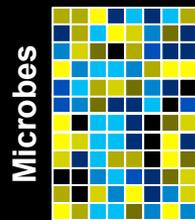


Genes,  
Genomes,  
Metabolic profiling,  
Relative abundances,  
Genetic variants...

V2



Samples



Microbes  
Relative  
abundances



# ChocoPhlAn: Cataloging core and unique marker gene sequences



Nicola Segata

- IMG alone now contains ~3,100 bacterial genomes
  - Plus ~100 archaeal, ~100 eukaryotic, and a few thousand viruses
  - About half final and half draft
- These comprise 1,222 bacterial species
  - 652 genera, 278 families, 130 orders, 66 classes, 33 phyla
  - 2,383 total clades
- **And roughly 12M genes**



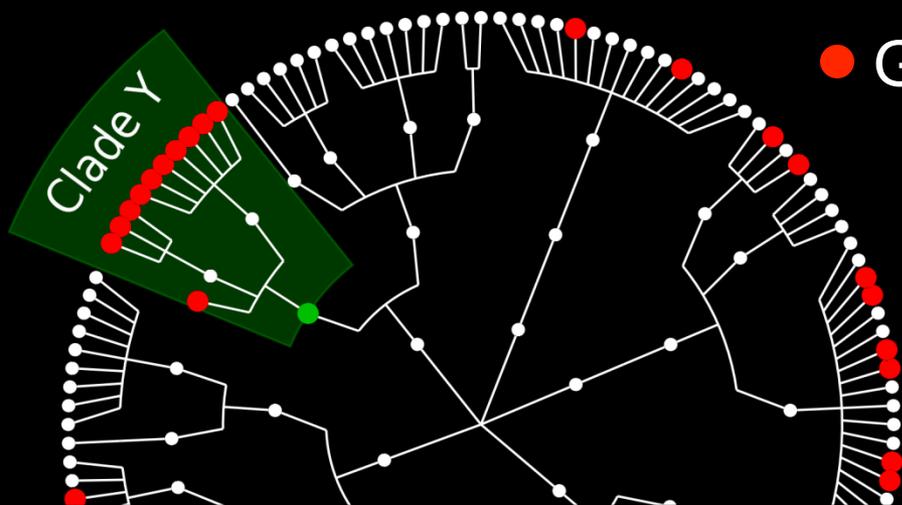
- These genes and genomes are a tremendous resource to:
  - *Identify **unique** markers that can be used to infer taxonomy*
  - *Identify **conserved** markers that can be used to infer phylogeny*
  - *Relate the microbial members of a community to their annotated **metagenomic functional potential***



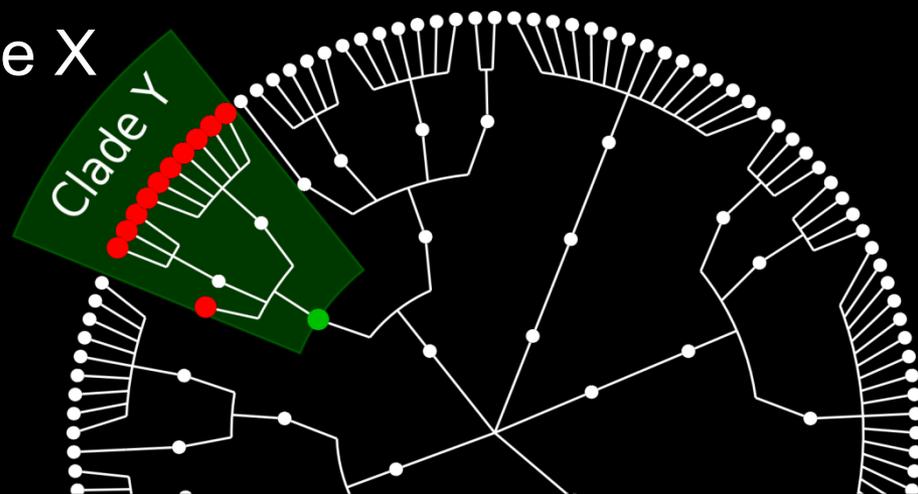
# MetaPhlAn: phylogenetically unique markers sequences for taxonomic profiling

X is a **core gene** for clade Y

X is a **unique marker gene** for clade Y



● Gene X



ChocoPhlAn (offline pipeline)

- Identify all **core genes** for all clades
- Screen core genes for **unique marker genes**
- Select most representative marker genes

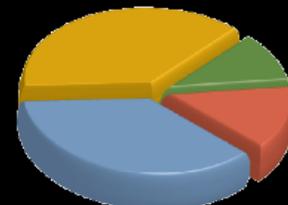
Unique  
marker  
genes DB

Available  
reference  
genomes

MetaPhlAn

Metagenome

- Blast reads against the marker genes
- Assign, count, normalize reads





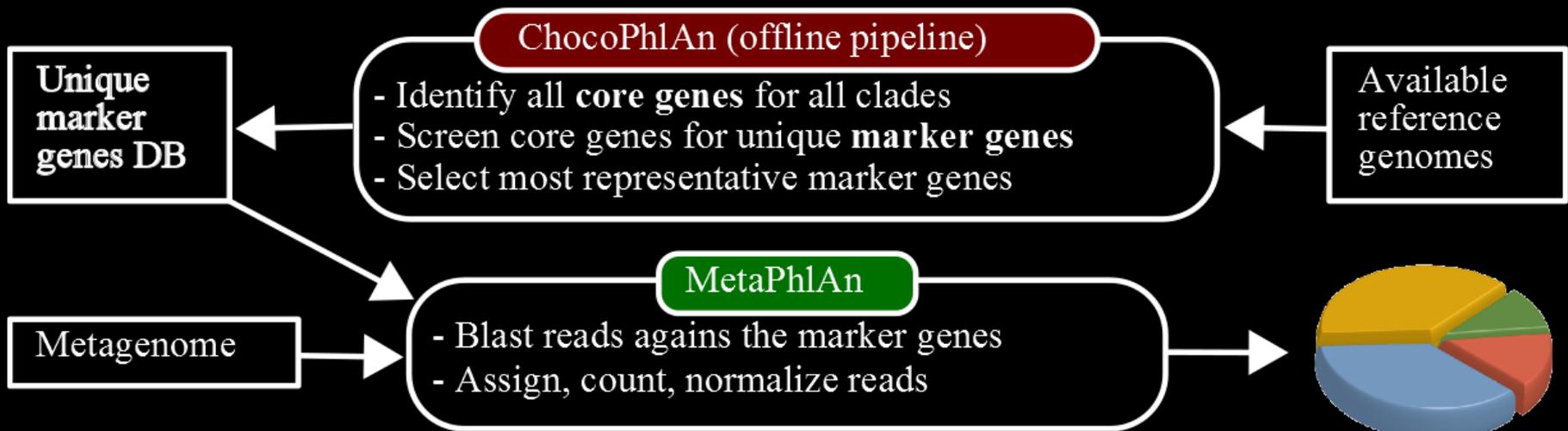
# MetaPhlAn: phylogenetically unique markers sequences for taxonomic profiling

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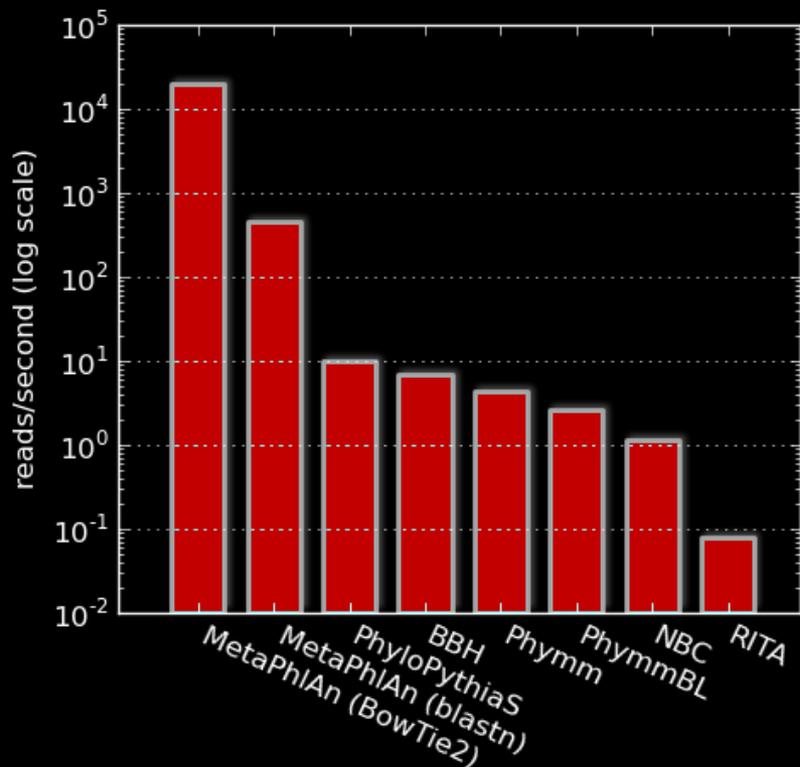
- ~2M total unique marker genes
  - Validated by whole-genome BLAST, not just annotated genes
- ~400k most representative markers used for identification
  - $231 \pm 107$  markers per species (350 fixed max)
  - Only 12 species with <15 markers (9 of which are *Brucella*)





# MetaPhlAn: inferring microbial abundances from metagenomic data using marker genes

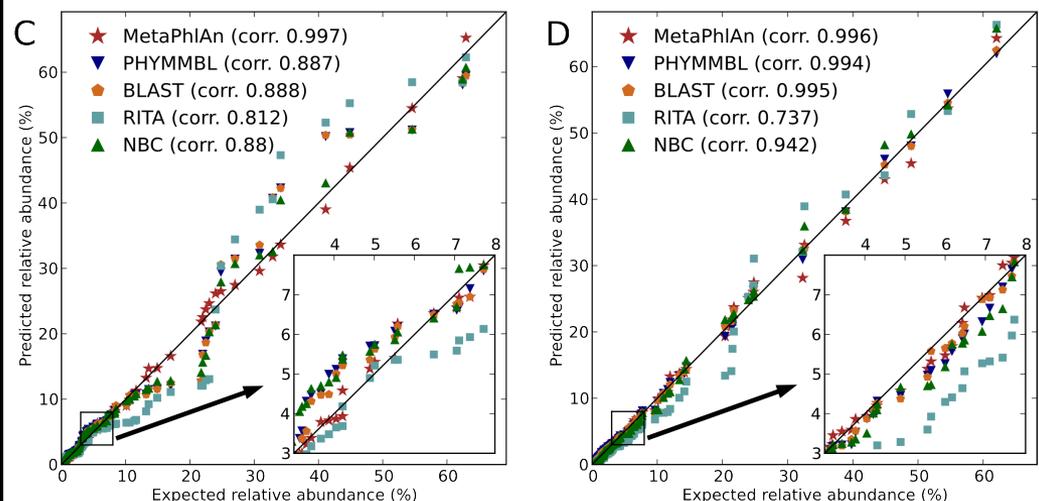
- Map metagenomic reads to marker genes to infer microbial abundances
  - Normalizing for copy number, gene length, etc.



~1000x faster than previous approaches

Hours instead of weeks for Illumina samples with 100Gbs of sequence

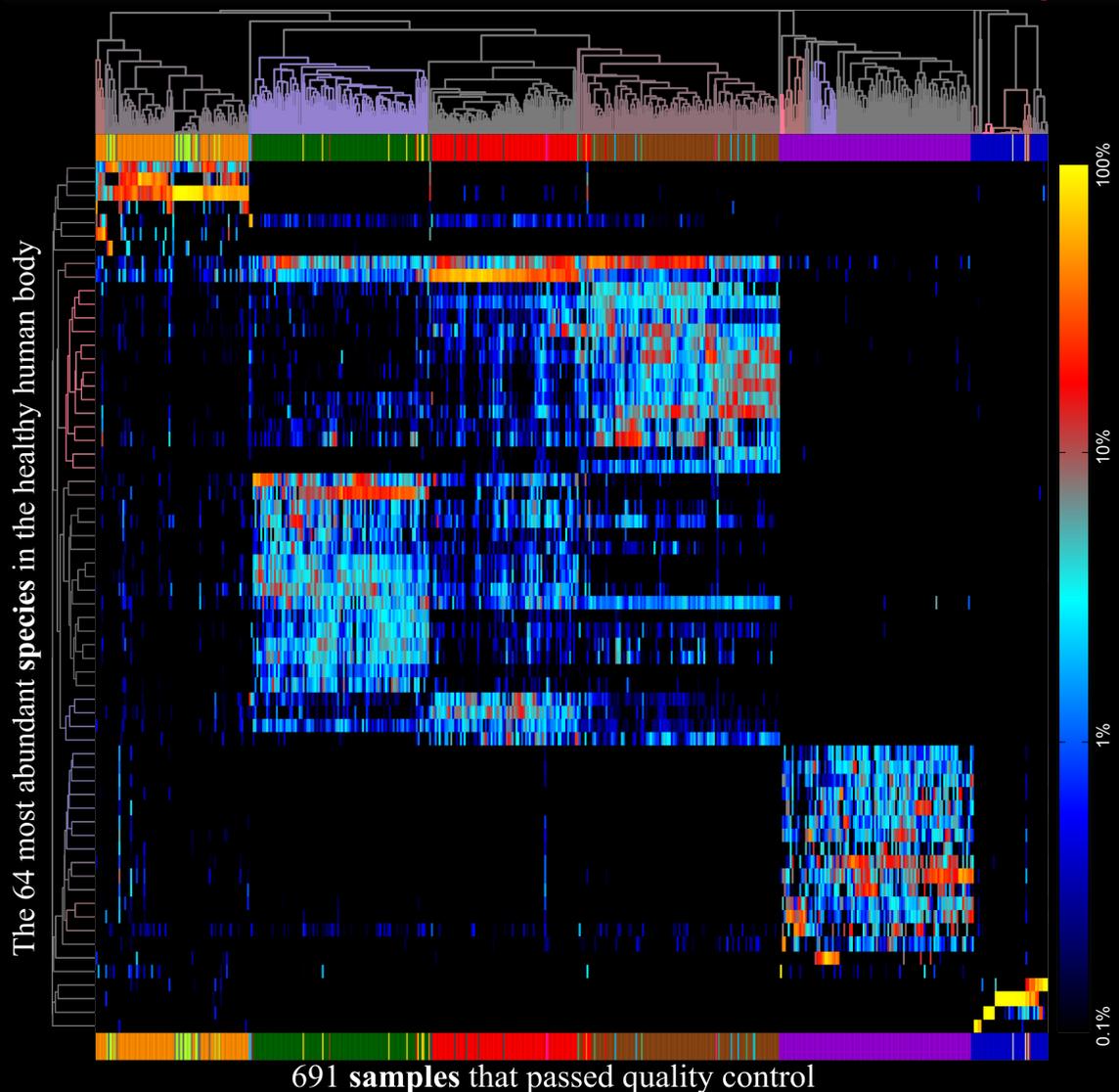
Eight log-normally distributed low-complexity (25 organisms each) synthetic metagenomes





# The HMP's human microbiome at species-level resolution

<http://hmpdacc.org/HMSMCP>



- |                               |                        |                                |                    |                     |
|-------------------------------|------------------------|--------------------------------|--------------------|---------------------|
| ■ anterior nares              | ■ throat               | ■ subgingival plaque           | ■ saliva           | ■ posterior fornix  |
| ■ right retroauricular crease | ■ buccal mucosa        | ■ tongue dorsum                | ■ palatine tonsils | ■ vaginal introitus |
| ■ left retroauricular crease  | ■ supragingival plaque | ■ attached keratinized gingiva | ■ stool            | ■ mid vagina        |

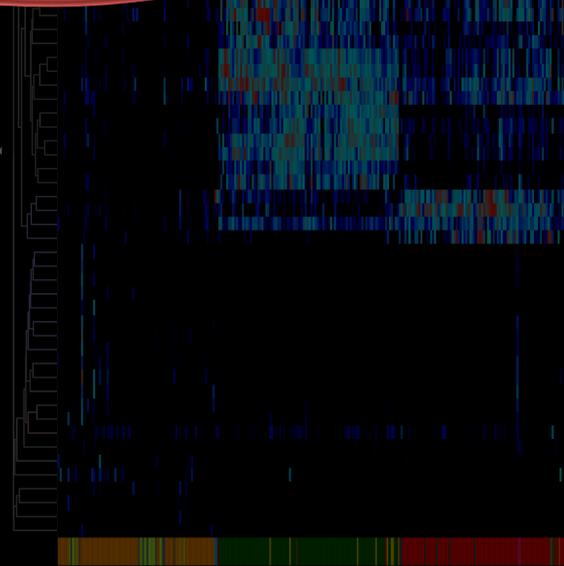


# The HMP's human microbiome at species-level resolution

<http://hmpdpc.org/HMSPCD>

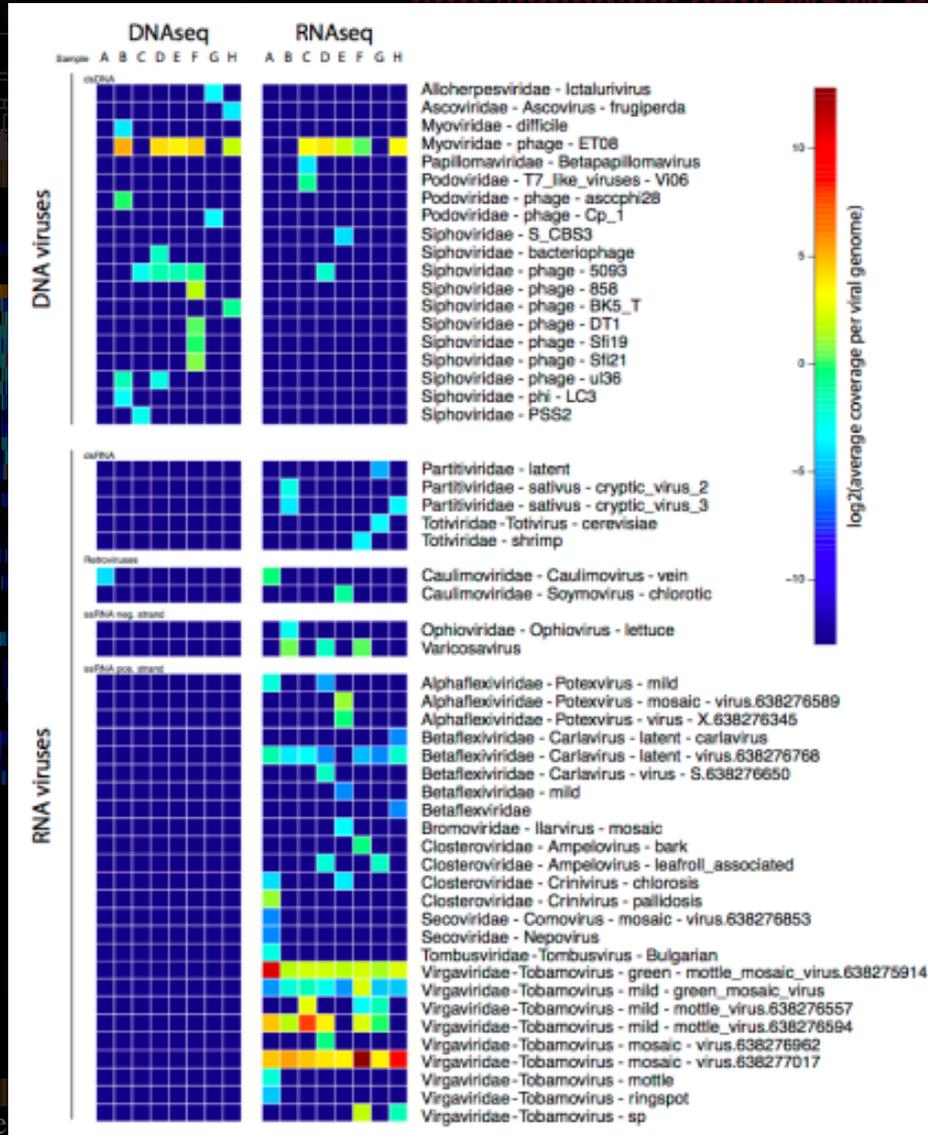
Watch for *MetaPhlan v2*:  
Additional archaea,  
DNA/RNA viruses, and  
eukaryotic microbes.

The 64 most abundant species in the



691 samples that passed

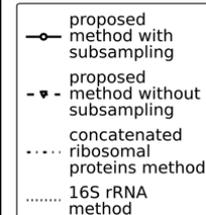
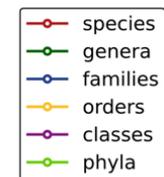
- anterior nares
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- subgingival plaque
- saliva
- posterior fornix
- right retroauricular crease
- buccal mucosa
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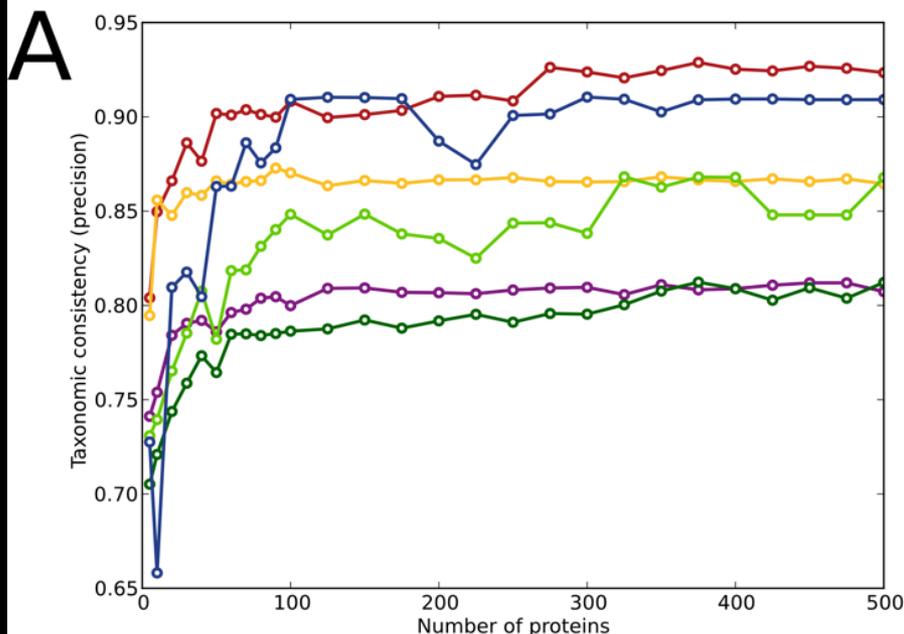


# PhyloPhlAn: From markers for taxonomy to markers for phylogeny

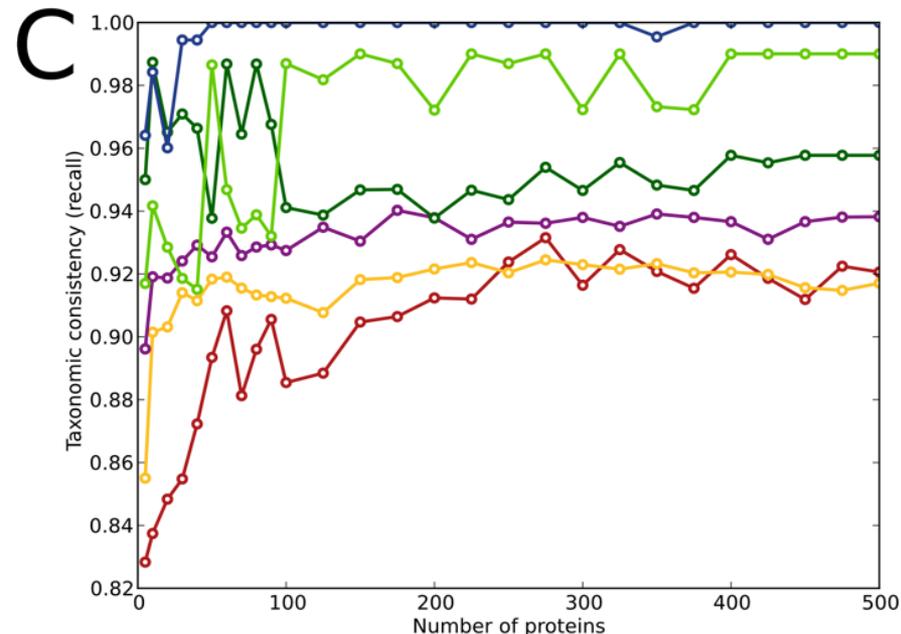
- Hundreds of unique markers per clade provide great taxonomic classification
- What if we use hundreds of conserved markers for phylogenetic classification?
  - PhyloPhlAn identifies the most informative residues of the most conserved 400 proteins
  - These can then be used for phylogenetic reconstruction, placement, and taxonomy



Taxonomic accuracy: precision



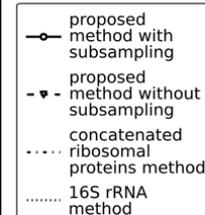
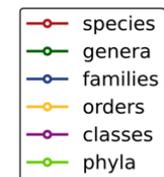
Taxonomic accuracy: recall



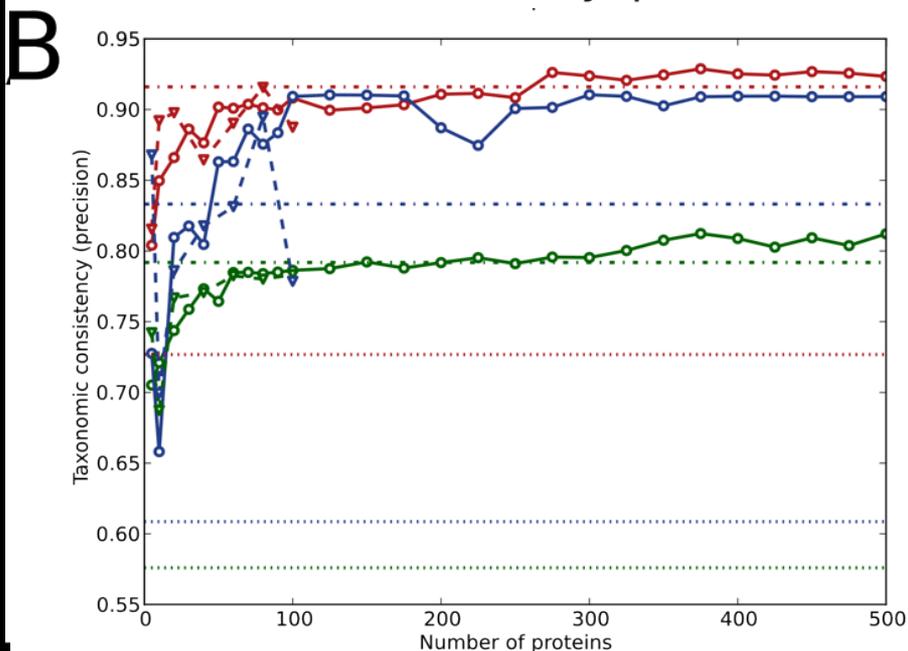


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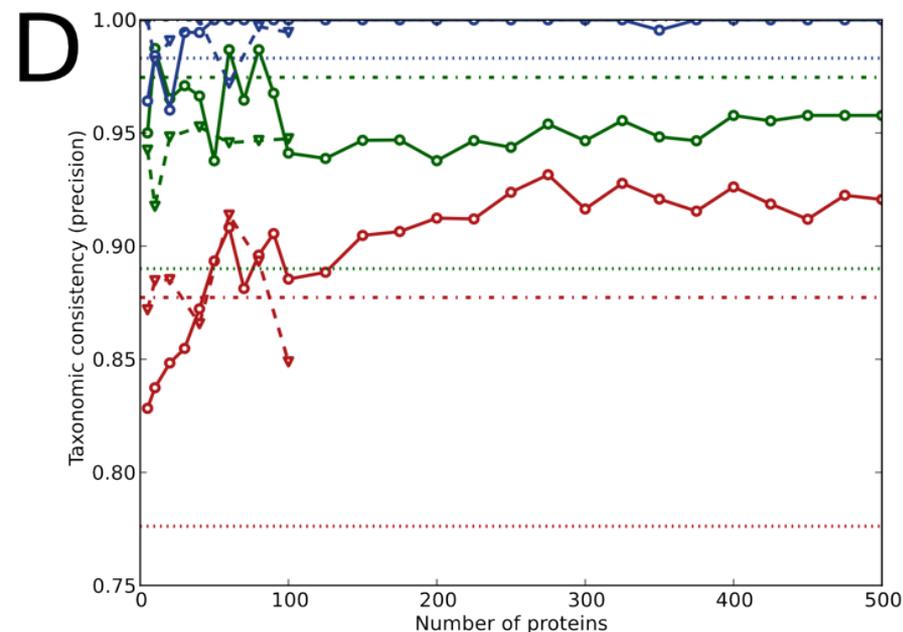
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Taxonomic accuracy: precision

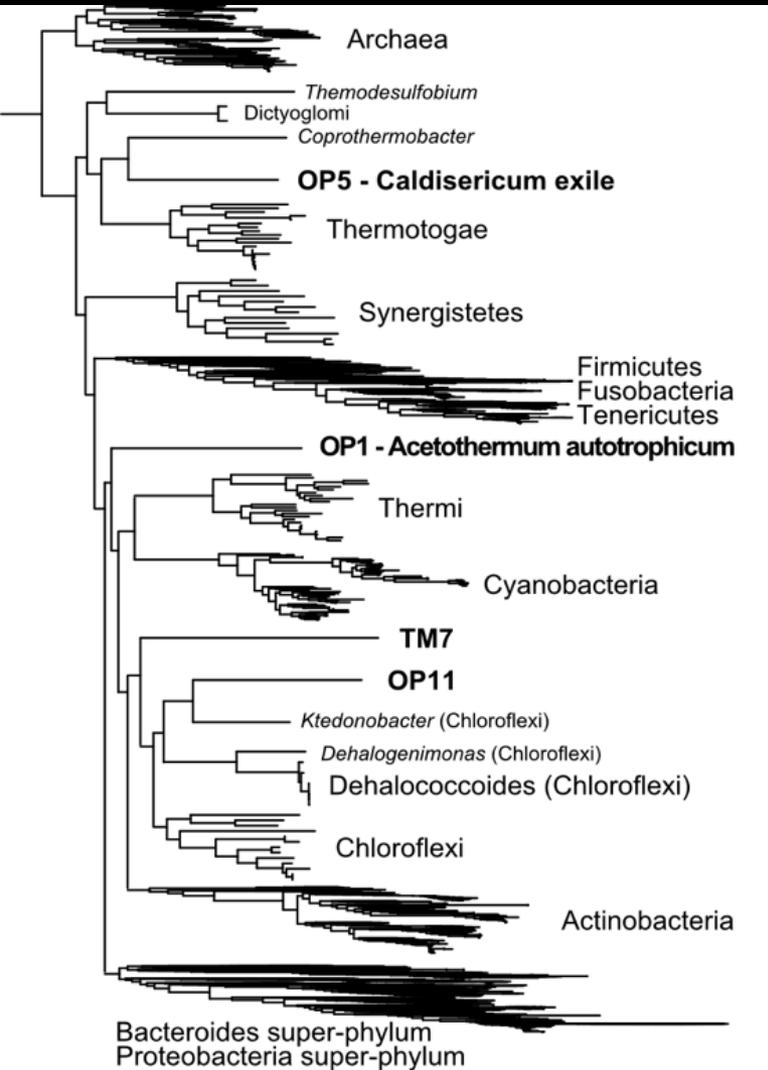


Taxonomic accuracy: recall



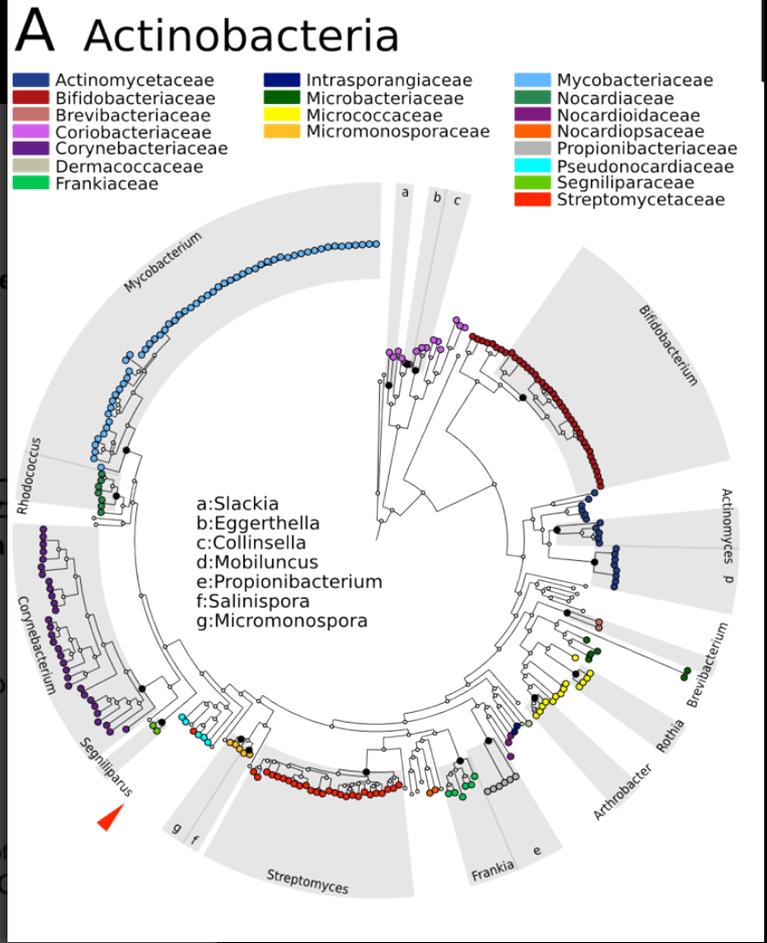
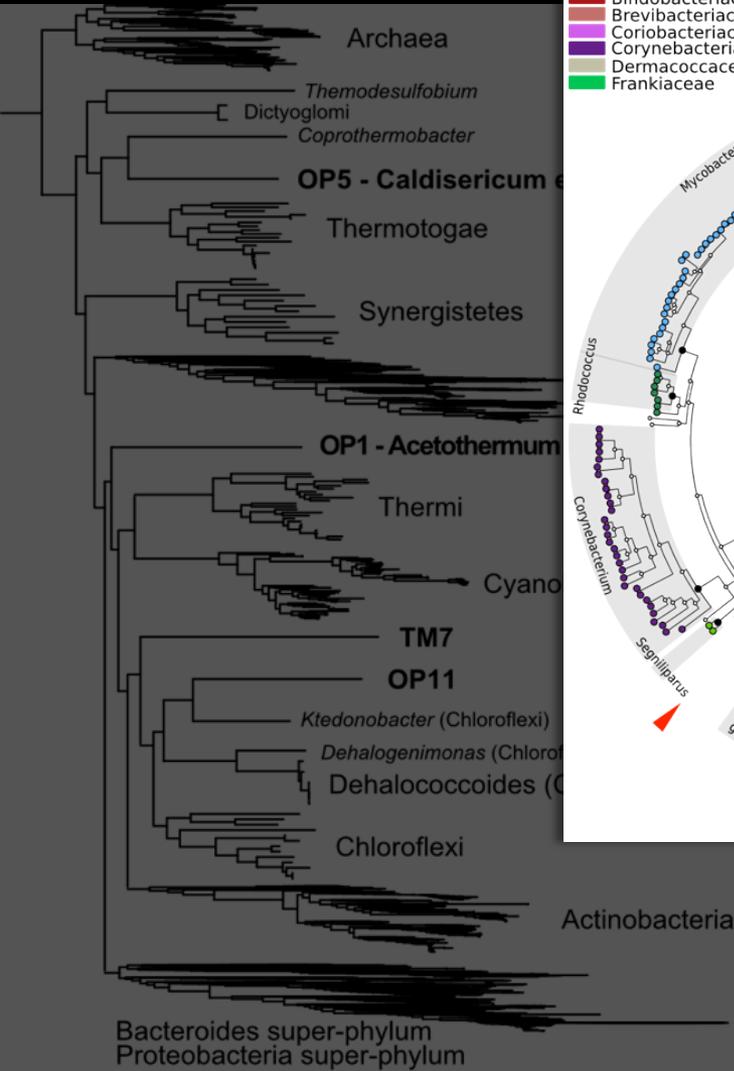


# PhyloPhlAn: for phyla to subspecies



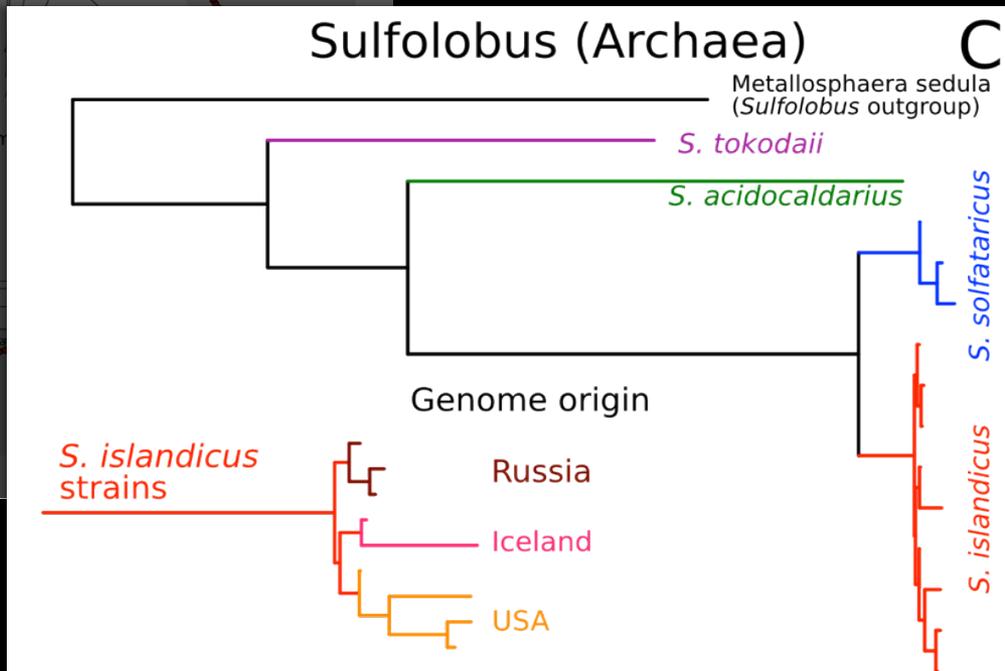
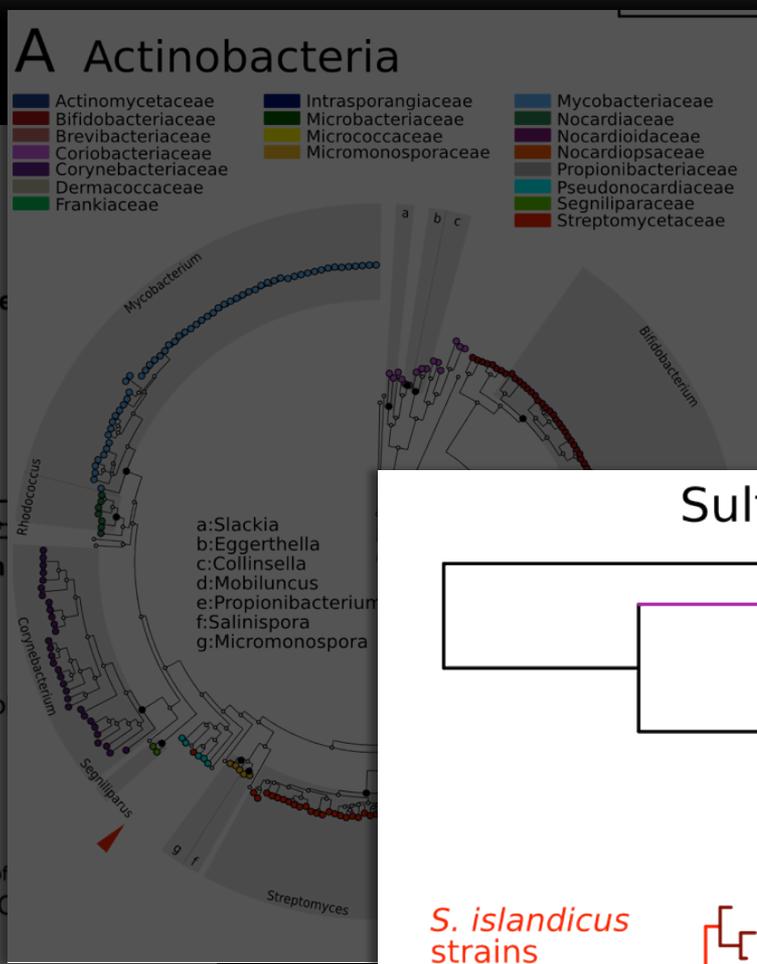
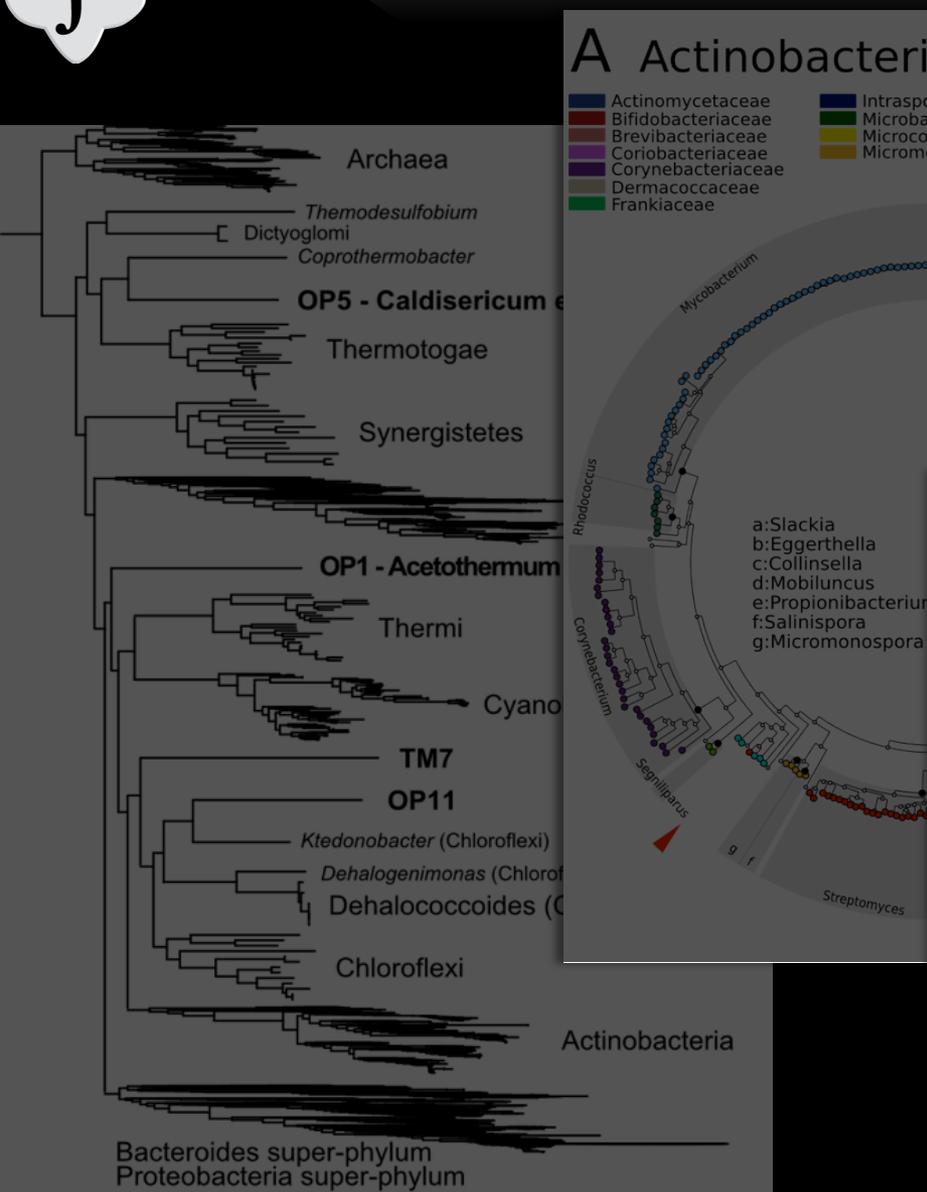


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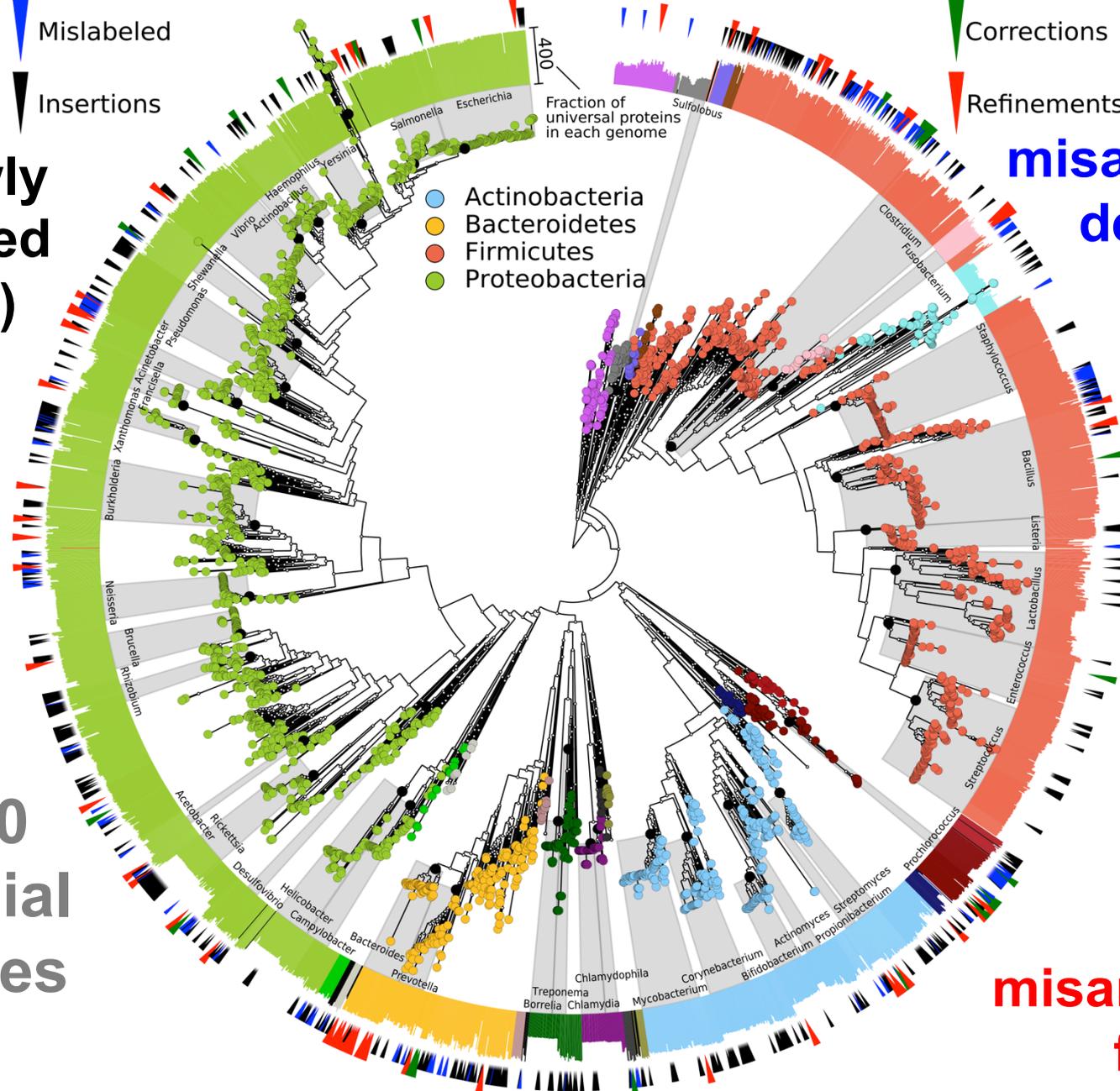


# PhyloPhlAn: for phyla to subspecies



**566 newly annotated (GEBA)**

**~3,700 microbial genomes**



Mislabeled  
Insertions

Corrections  
Refinements

Fraction of universal proteins in each genome

**111 misannotation detected**

**46 misannotation fixed**

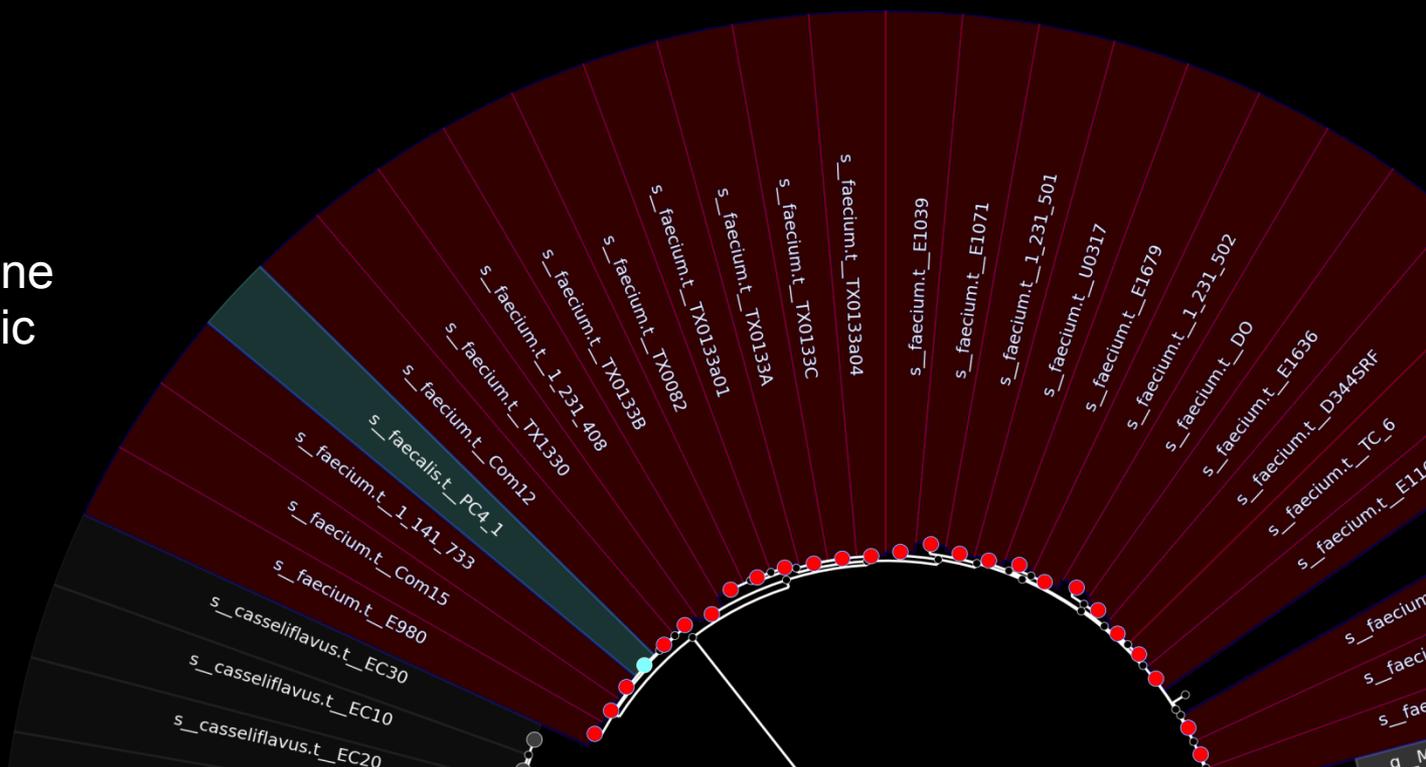
- Acidobacteria
- Chloroflexi
- Fusobacteria
- Tenericutes
- Other
- Aquificae
- Crenarchaeota
- Planctomycetes
- Thermi
- Chlamydiae
- Cyanobacteria
- Spirochaetes
- Thermotogae
- Chlorobi
- Euryarchaeota
- Synergistetes
- Verrucomicrobia





# PhyloPhlAn: Taxonomic curation and reannotation

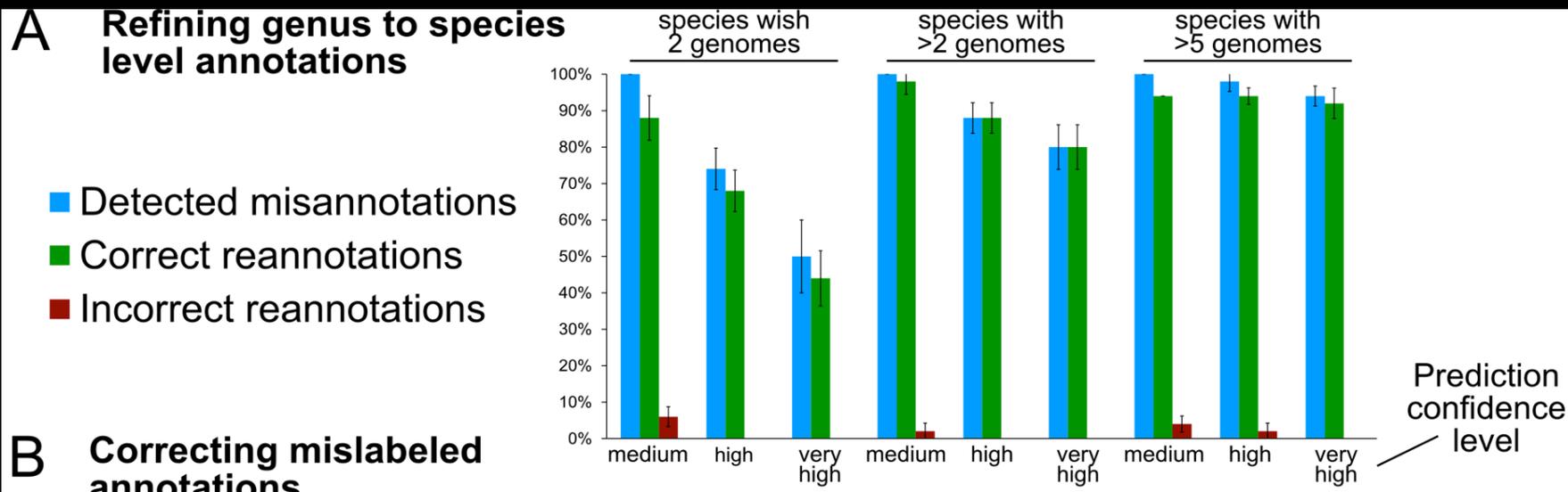
- Taxa with at least one 'unknown' taxonomic level: 445
- Additional taxa we detected as suspicious: 111



	Example	Very high confidence	High confidence	Medium confidence
Corrected	A B C→A B D	26	3	26
Refined	A B ?→A B C	67	25	75
Removed	A B C→A B ?	11	1	1
Incomplete	A ? ?→A ? ?	224	10	66



# PhyloPhlAn: Automatically assigning precise taxonomy using precise phylogeny

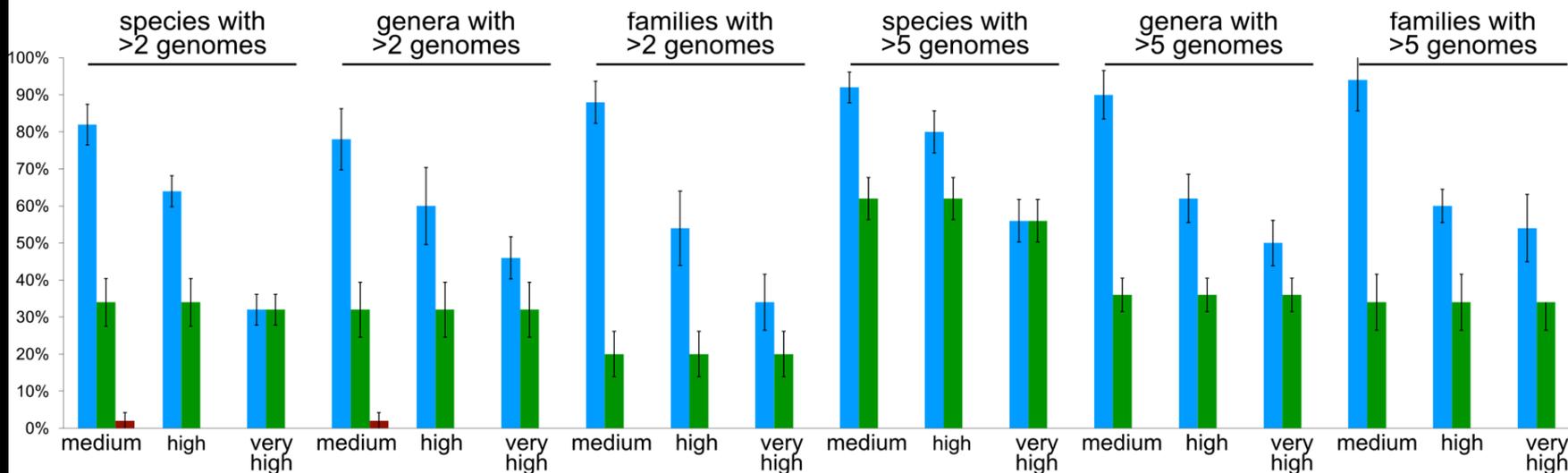




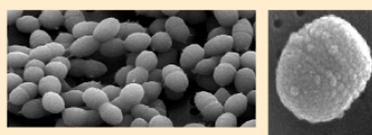
# PhyloPhlAn: Automatically assigning precise taxonomy using precise phylogeny

- Detected misannotations
- Correct reannotations
- Incorrect reannotations

## B Correcting mislabeled annotations



# A map of diversity in the human microbiome



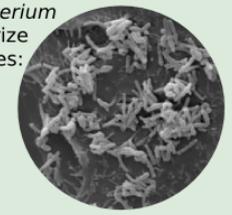
*Streptococcus* dominates the oral cavity with *S. mitis* > 75% in the **cheek**

*Propionibacterium acnes* lives on the skin and **nose** of most people



Many *Corynebacterium* species characterize different body sites:

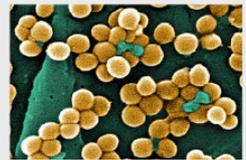
- C. matruchoti* the **plaque**
- C. accolens* the **nose**
- C. croppenstedtii* the **skin**



*Lactobacillus* species (*L. gasseri*, *L. jensenii*, *L. crispatus*, *L. iners*) are predominant but mutually exclusive in the **vagina**



*Staphylococcus epidermidis* colonizes external body sites



- Commensal microbes
- ☆ Potential pathogens

## The four most abundant phyla

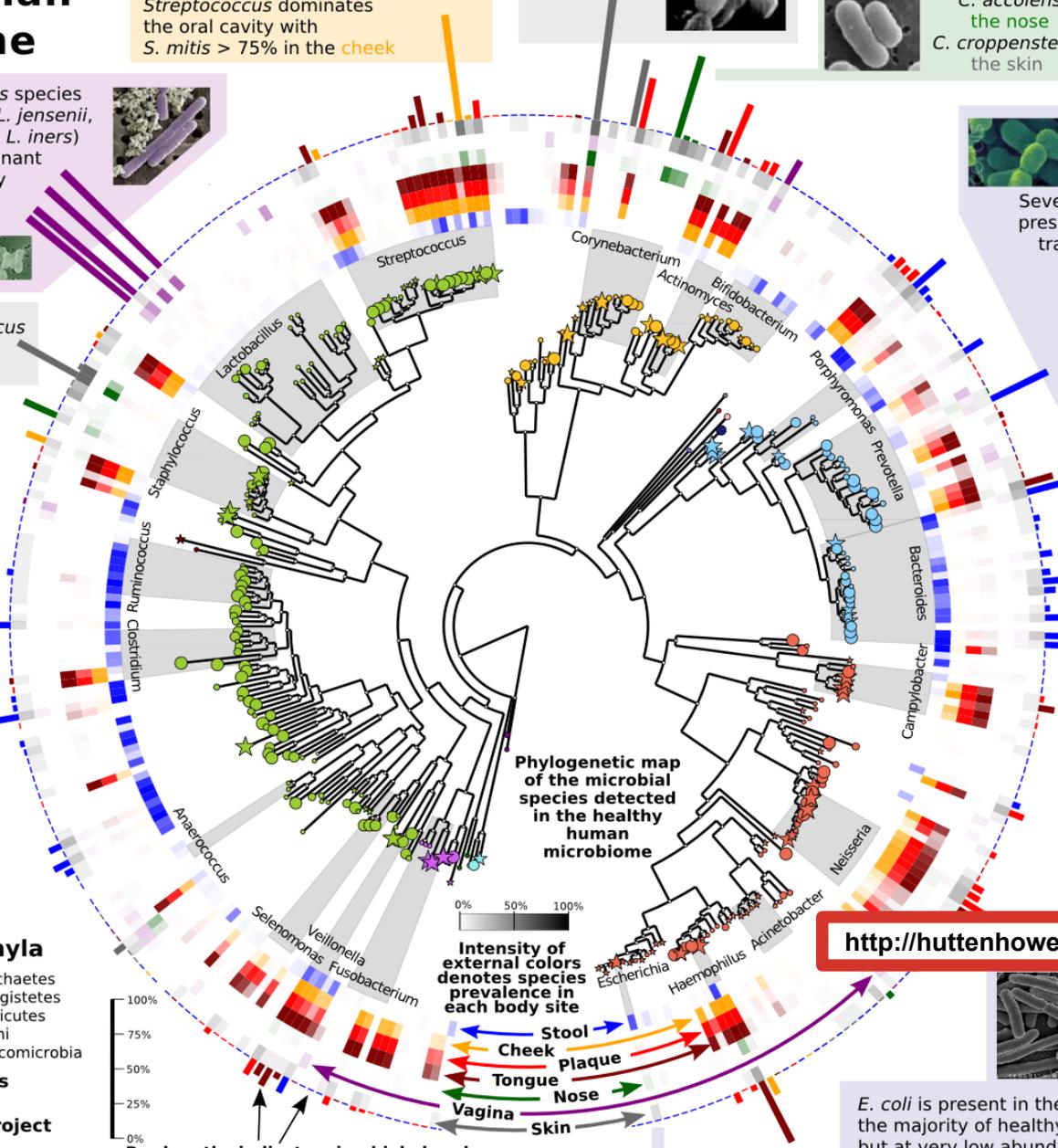
- Actinobacteria
- Bacteroidetes
- Firmicutes
- Proteobacteria

## Low abundance phyla

- Chloroflexi
- Cyanobacteria
- Euryarchaeota
- Fusobacteria
- Lentisphaerae
- Spirochaetes
- Synergistetes
- Tenericutes
- Thermi
- Verrucomicrobia

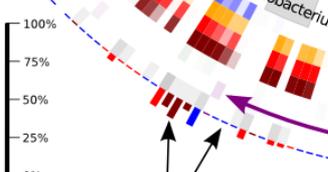
National Institutes of Health  
Human Microbiome Project

N. Segata & C. Huttenhower  
<http://huttenhower.sph.harvard.edu>  
(generated using GCLiDER and mOTU from MetaPhlan analysis)

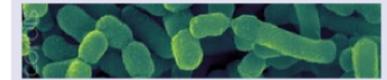


Phylogenetic map of the microbial species detected in the healthy human microbiome

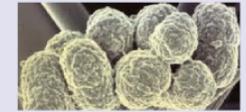
Intensity of external colors denotes species prevalence in each body site



Bar lengths indicate microbial abundance (colored by body site of greatest prevalence)



Several *Prevotella* species are present in the gastrointestinal tract. *P. copri* is present in 19% of the subjects and dominates the **intestinal** flora when present



Microscopy from <http://bacmap.wishartlab.com>

*Bacteroides* is the most abundant genus in the **gut** of almost all healthy subjects



*Campylobacter* includes opportunistic pathogens, but members live in the oral cavities of most healthy people in the cohort



<http://huttenhower.sph.harvard.edu/graphlan>

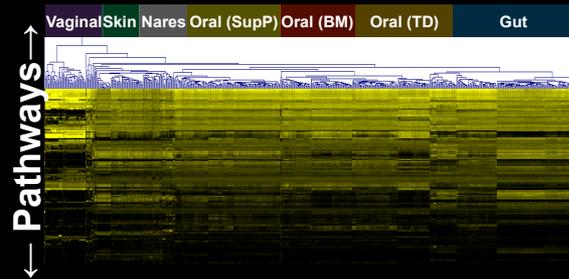


*E. coli* is present in the **gut** of the majority of healthy subjects but at very low abundance





# HUMAnN: Metabolic profiling for microbial communities

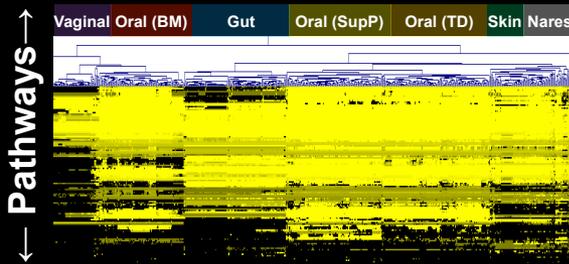


← Samples →



**Pathway abundance**

**Pathway coverage**



← Samples →



**100 subjects**  
 1-3 visits/subject  
 ~7 body sites/visit  
 10-200M reads/sample  
 100bp reads

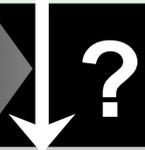


BLAST

**Functional seq.**  
 KEGG + MetaCyc  
 CAZy, TCDB,  
 VFDB, MEROPS...



Metagenomic reads



Enzymes and pathways

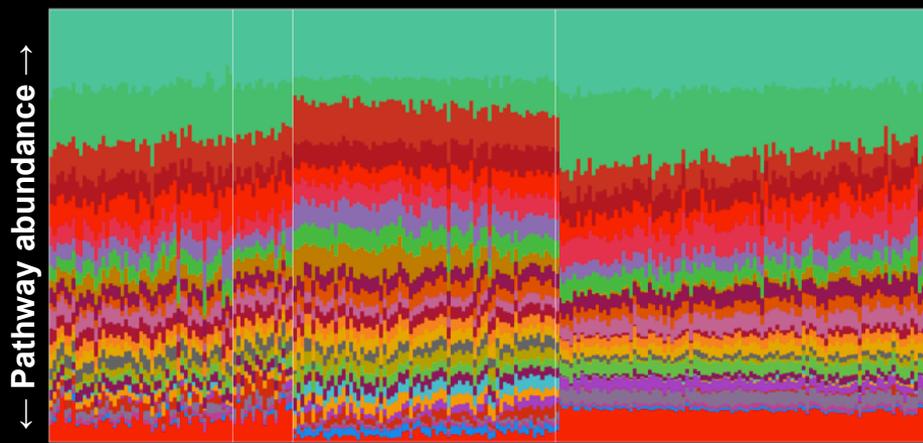
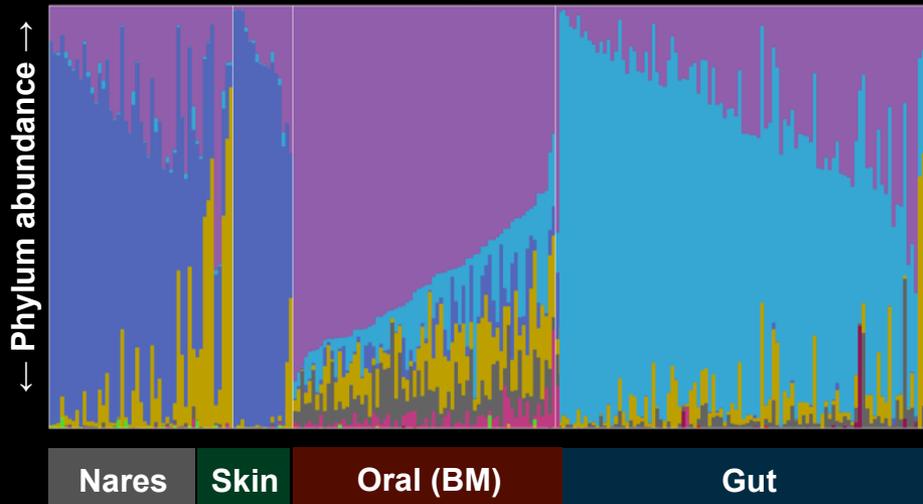
**HUMAnN**  
 HMP Unified Metabolic  
 Analysis Network  
<http://huttenhower.sph.harvard.edu/humann>



# The “core” human microbiome consists of genes, not bugs.

<http://hmpdacc.org/HMSMCP>

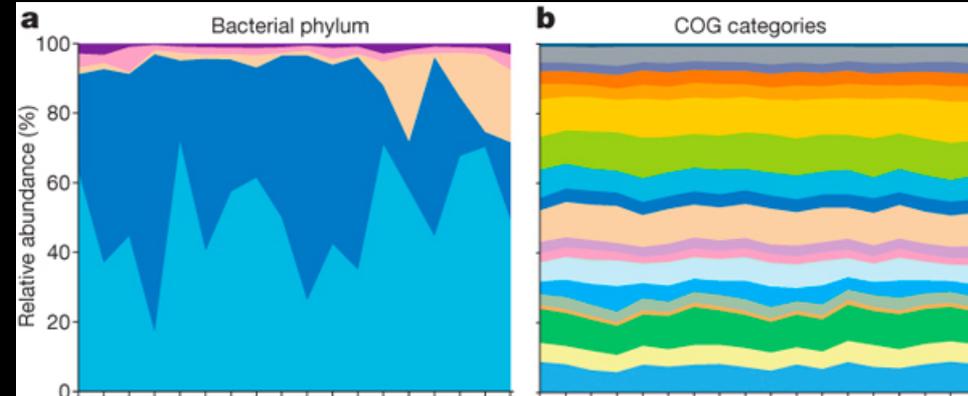
← Subjects →



← Subjects →

<http://hmpdacc.org/HMMRC>

Turnbaugh 2009



## Bacterial community assembly based on functional genes rather than species

Catherine Burke<sup>ab</sup>, Peter Steinberg<sup>cd</sup>, Doug Rusch<sup>e</sup>, Staffan Kjelleberg<sup>af</sup>, and Torsten Thomas<sup>ab,1</sup>

<sup>a</sup>School of Biotechnology and Biomolecular Sciences, <sup>b</sup>School of Biological, Earth and Environmental Sciences, Centre for Environmental and Estuarine Science, <sup>c</sup>University of Technology, Ultimo, New South Wales, Sydney, New South Wales 2052, Australia; <sup>d</sup>The J. Craig Venter Institute, University of Technology, Ultimo, New South Wales 2052, Australia; <sup>e</sup>Sydney Institute of Marine Science, Mosman, New South Wales 2088, Australia; <sup>f</sup>The J. Craig Venter Institute, Rockville, Maryland 20850, USA; <sup>1</sup>Environmental Life Sciences Engineering, Nanyang Technological University, Singapore

Edited by W. Ford Doolittle, Dalhousie University, Halifax, Canada, and approved July 14, 2011 (received May 11, 2011)



## The convergence of carbohydrate active gene repertoires in human gut microbes

Catherine A. Lozupone<sup>af</sup>, Micah Hamady<sup>g</sup>, Brandi L. Cantarel<sup>af</sup>, Pedro M. Coutinho<sup>af</sup>, Bernard Henrissat<sup>af</sup>, Jeffrey I. Gordon<sup>h</sup>, and Rob Knight<sup>af,1</sup>

<sup>a</sup>Department of Chemistry and Biochemistry, University of Colorado, Boulder, CO 80309; <sup>b</sup>Center for Genome Sciences, Washington University School of Medicine, St. Louis, MO 63108; <sup>c</sup>Department of Computer Science, University of Colorado, Boulder, CO 80309; and <sup>d</sup>Centre National de la Recherche Scientifique, Unite Mixte de Recherche 6098, <sup>e</sup>Universit  Aix-Marseille I and II, Marseille 13284, France

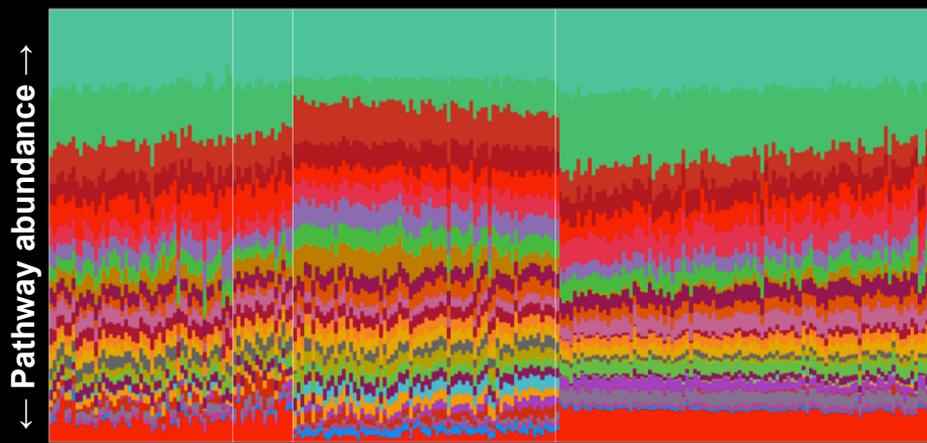
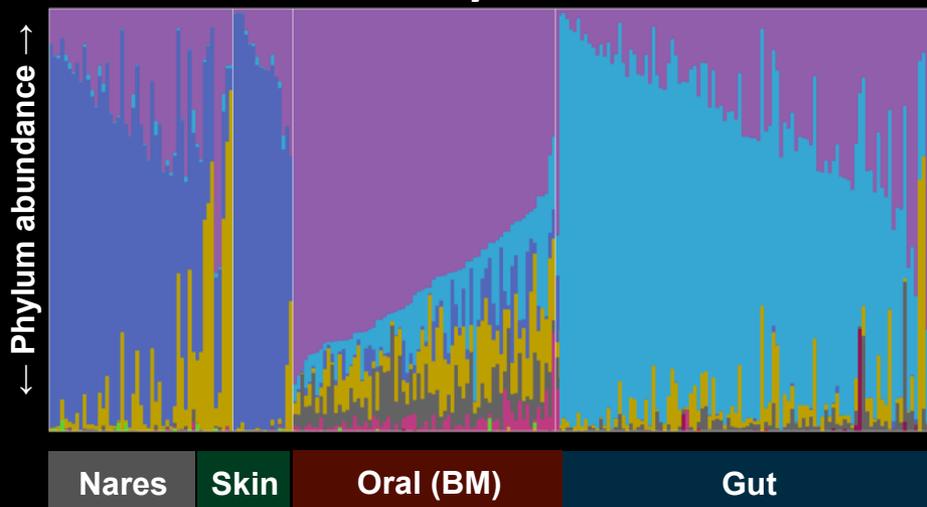
Contributed by Jeffrey I. Gordon, July 31, 2008 (sent for review June 13, 2008)



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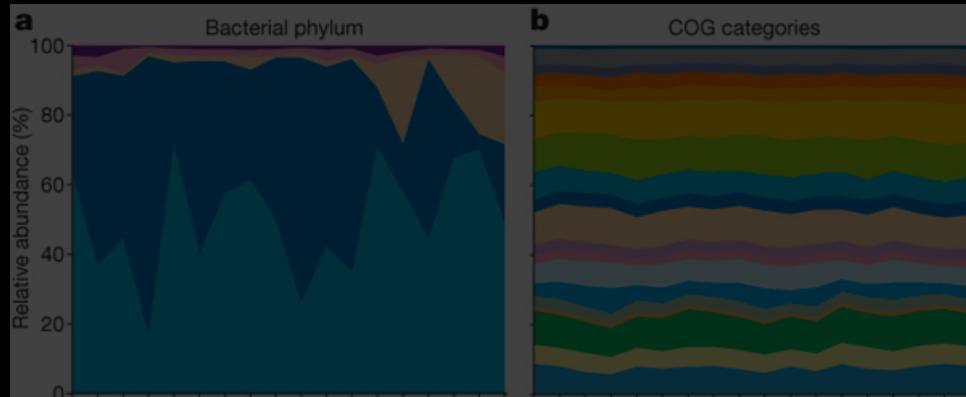
← Subjects →



← Subjects →

<http://hmpdacc.org/HMMRC>

Turnbaugh 2009



↙ This is the “core” human microbiome,  
↖ Not this.

- Over 2/3 of its genes are uncharacterized, more than almost any single bacterial genome
- We don't know how its “cell types” communicate
- We don't know their physical structure or lineages

## The convergence of carbohydrate active gene repertoires in human gut microbes

Catherine A. Lozupone<sup>a\*</sup>, Micah Hamady<sup>a</sup>, Brandi L. Cantarel<sup>b,c</sup>, Pedro M. Coutinho<sup>b,c</sup>, Bernard Henrissat<sup>b,c</sup>, Jeffrey I. Gordon<sup>a</sup>, and Rob Knight<sup>a\*</sup>

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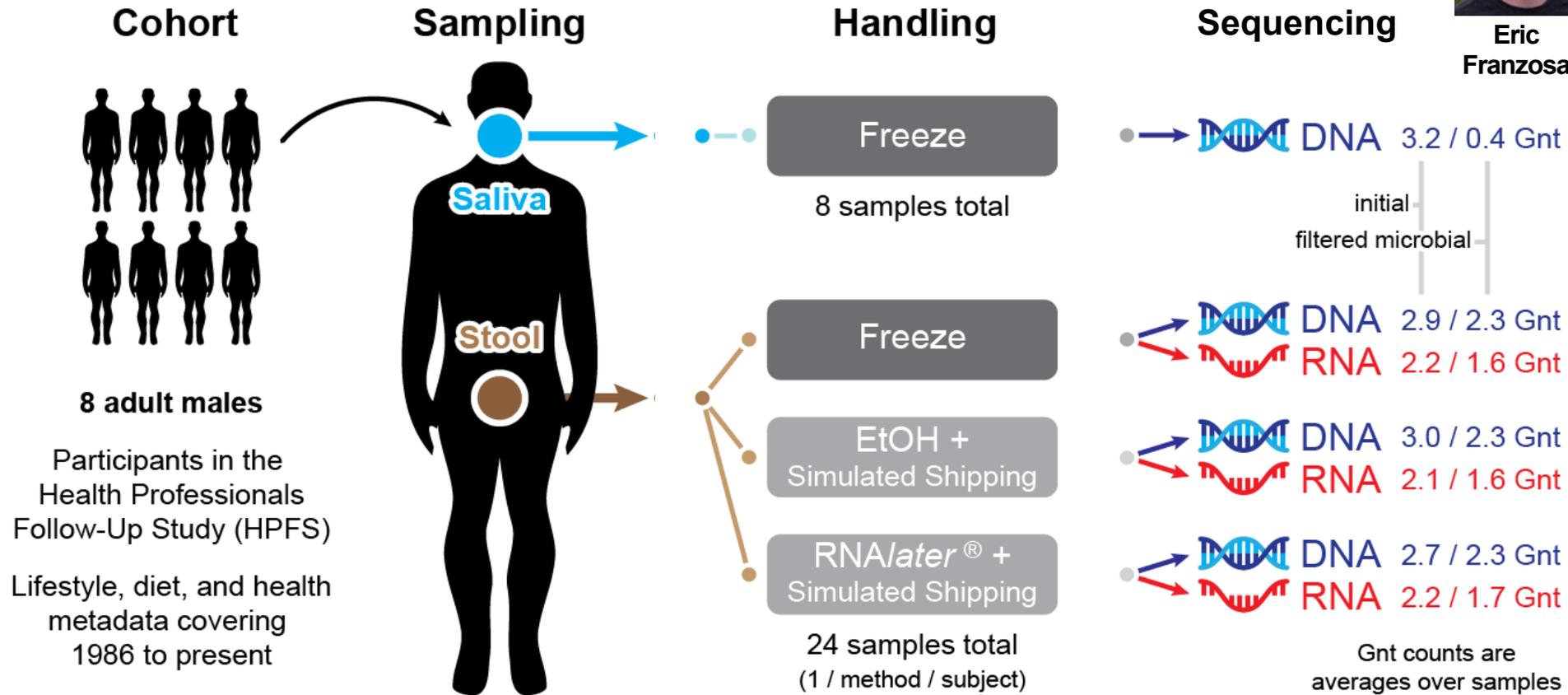
Contributed by Jeffrey I. Gordon, July 31, 2008 (sent for review June 13, 2008)

# HPFS Pilot Project: Overview



Eric Franzosa

With Jacques Izard, Andy Chan, Wendy Garrett



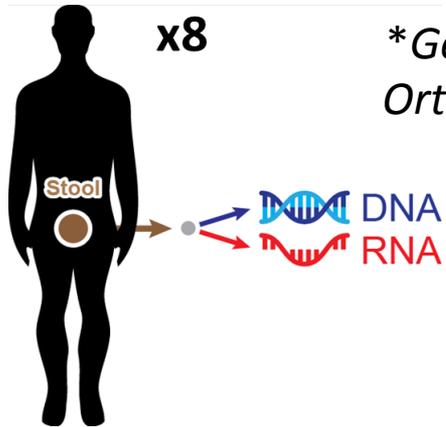
0) Investigate links between the mouth and gut microbiomes

1) Evaluate stability of meta'omic samples under subject-shipped conditions

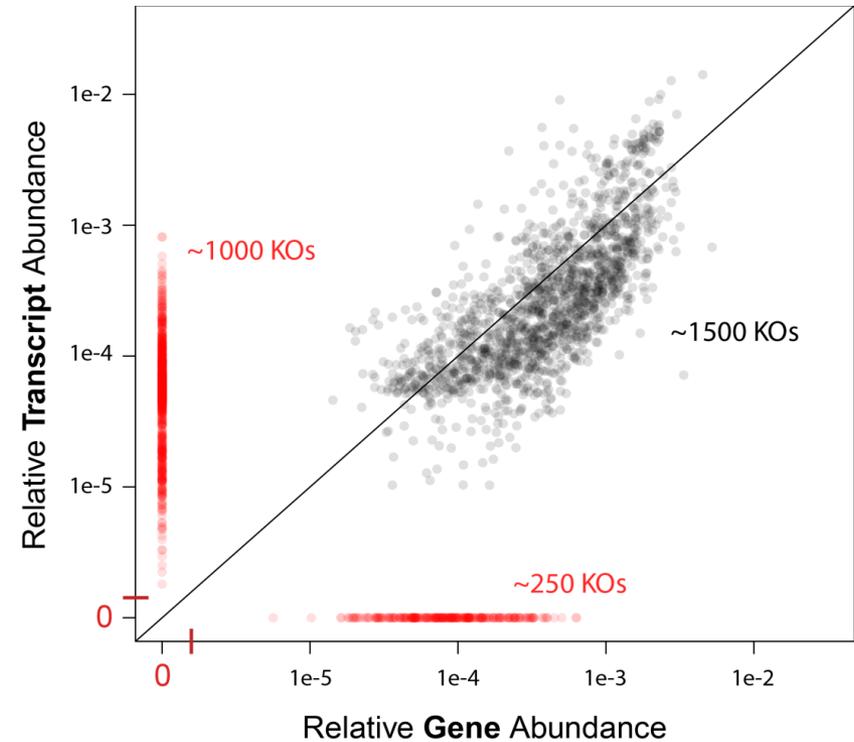
2) Relate the gut metagenome and metatranscriptome

## 2) Relating the gut metagenome and metatranscriptome

A large portion of genes  
(~50%) correlate well at the  
DNA and RNA levels



\*Genes are KEGG  
Orthogroups, KOs

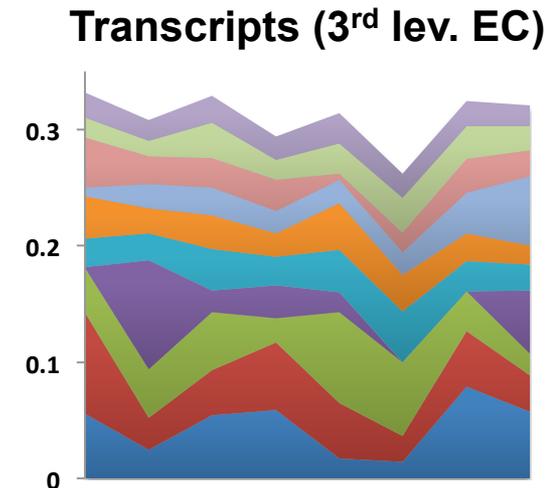
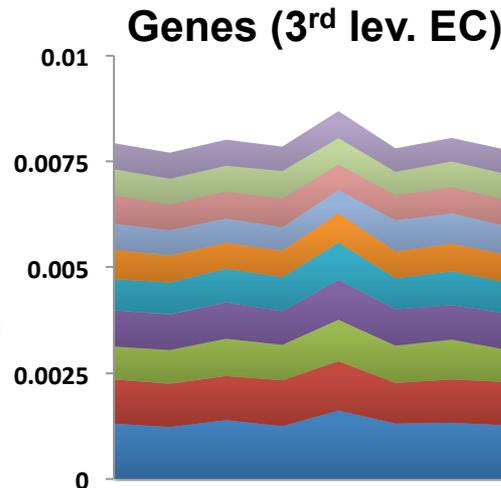
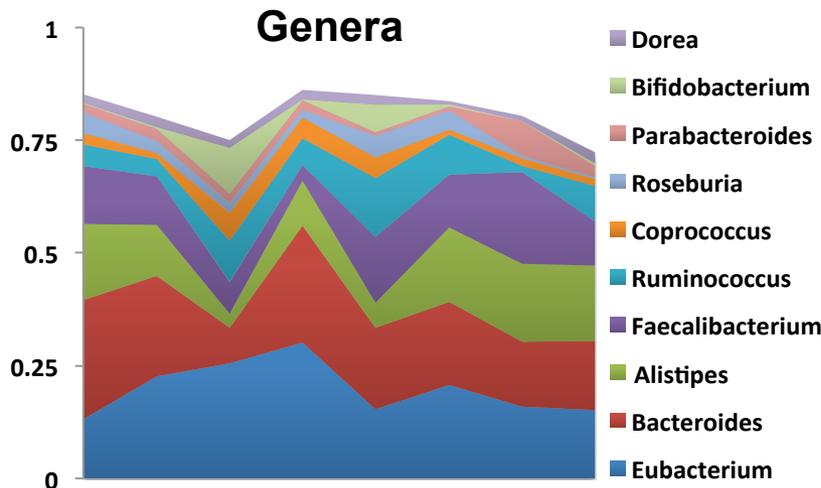


## 2) Relating the gut metagenome and metatranscriptome

- Microbial membership varies.
  - Early colonization? Genetics?
- Over time, the community “solves” for a habitat-specific metagenome.
- It then differentially regulates that metagenome.
  - These two types of regulation differ *at least* in time scale.



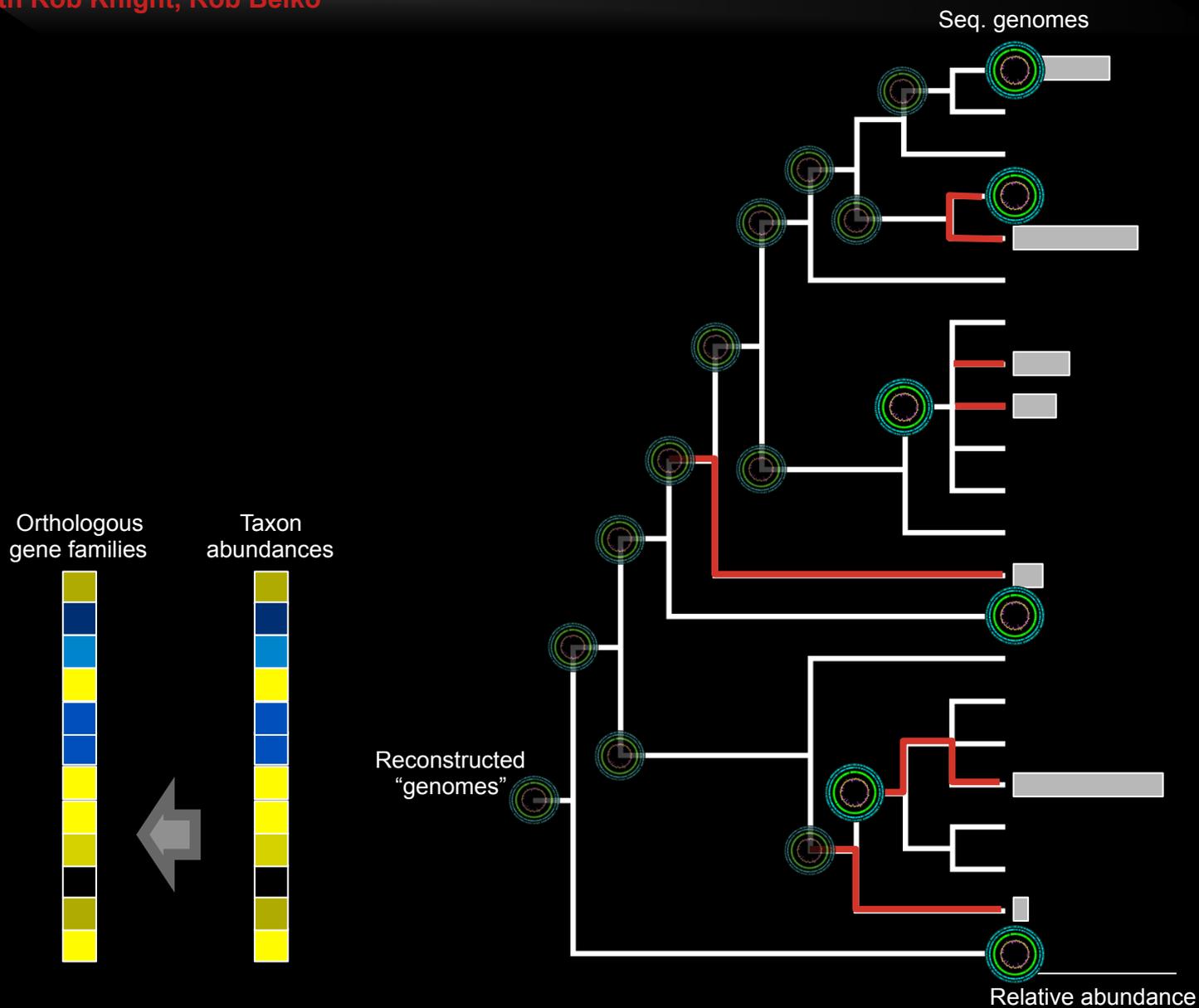
How consistent are the top ten...





# PICRUSt: Inferring community metagenomic potential from marker gene sequencing

With Rob Knight, Rob Beiko

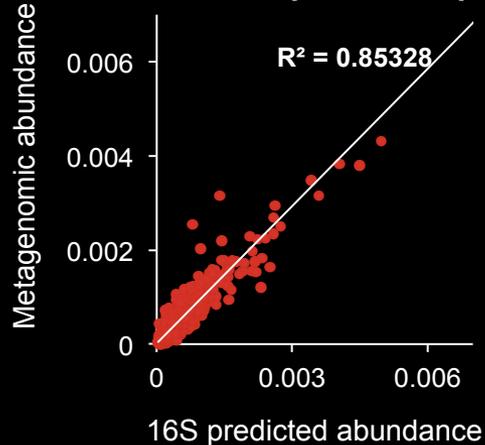




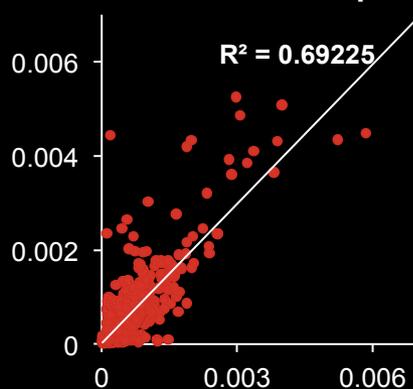
# PICRUSt: Inferring community metagenomic potential from marker gene sequencing

With Rob Knight, Rob Beiko

Gene families in one HMP hard palate sample



HMP stool sample



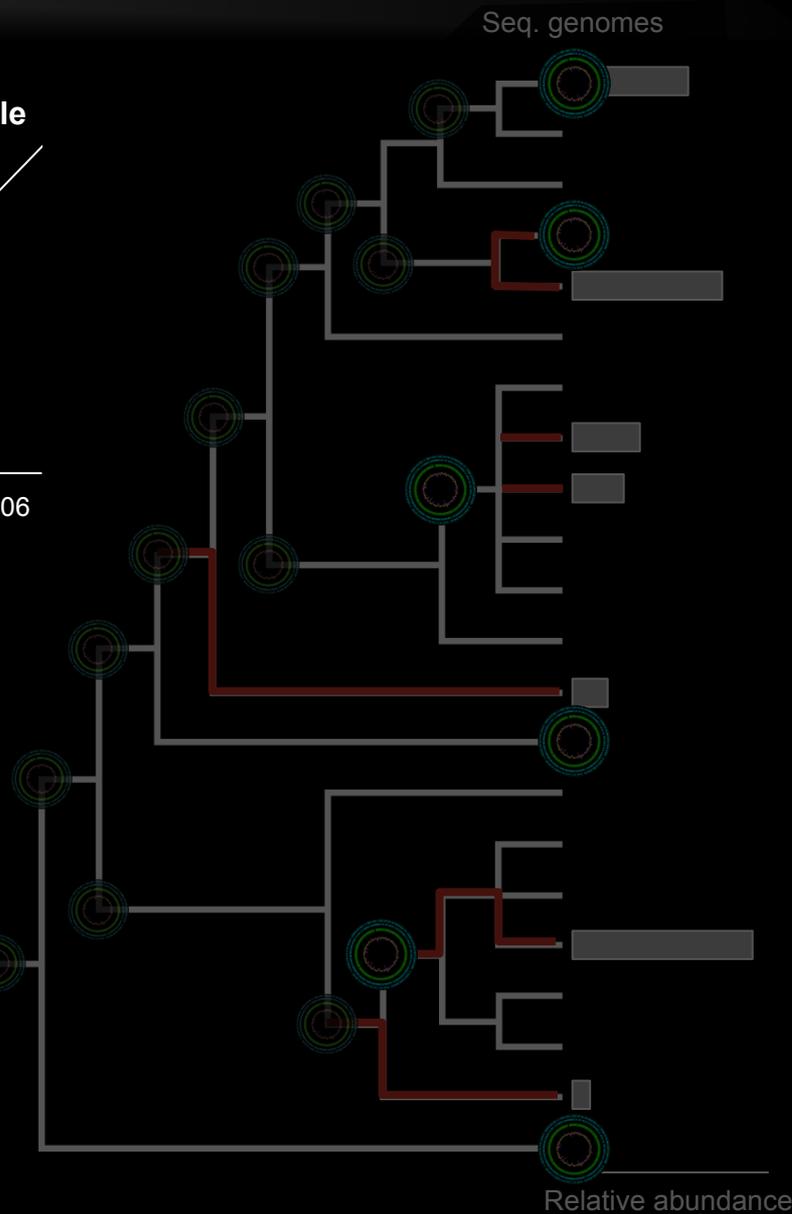
Orthologous gene families



Taxon abundances



Reconstructed "genomes"





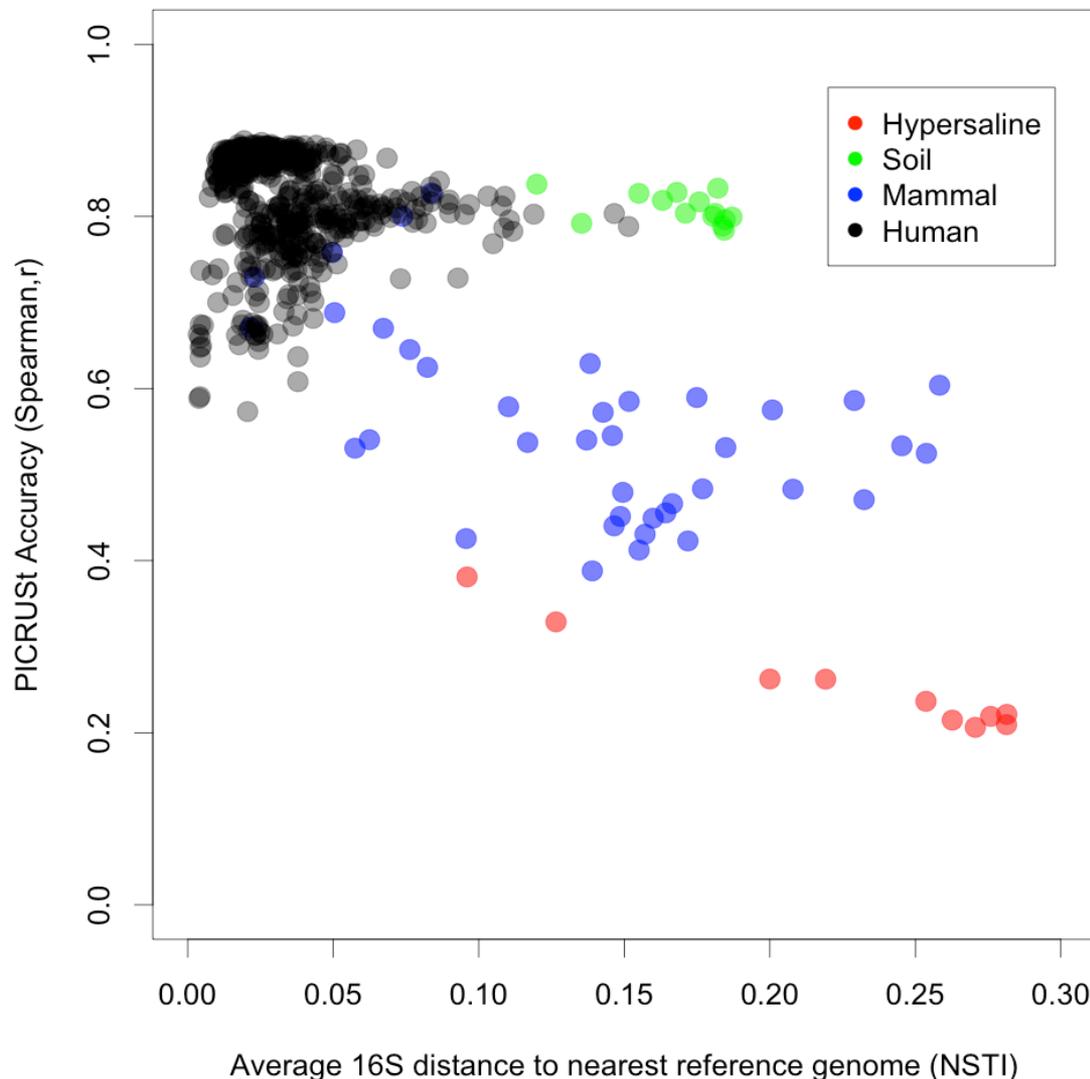
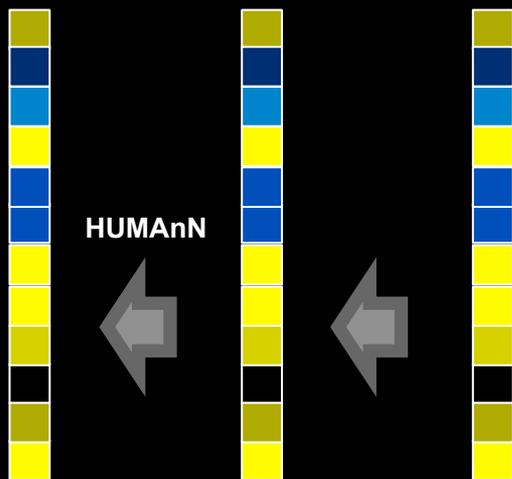
# PICRUSt: Inferring community metagenomic potential from marker gene sequencing

With Rob Knight, Rob Beiko

One can recover general community function with reasonable accuracy from 16S profiles.

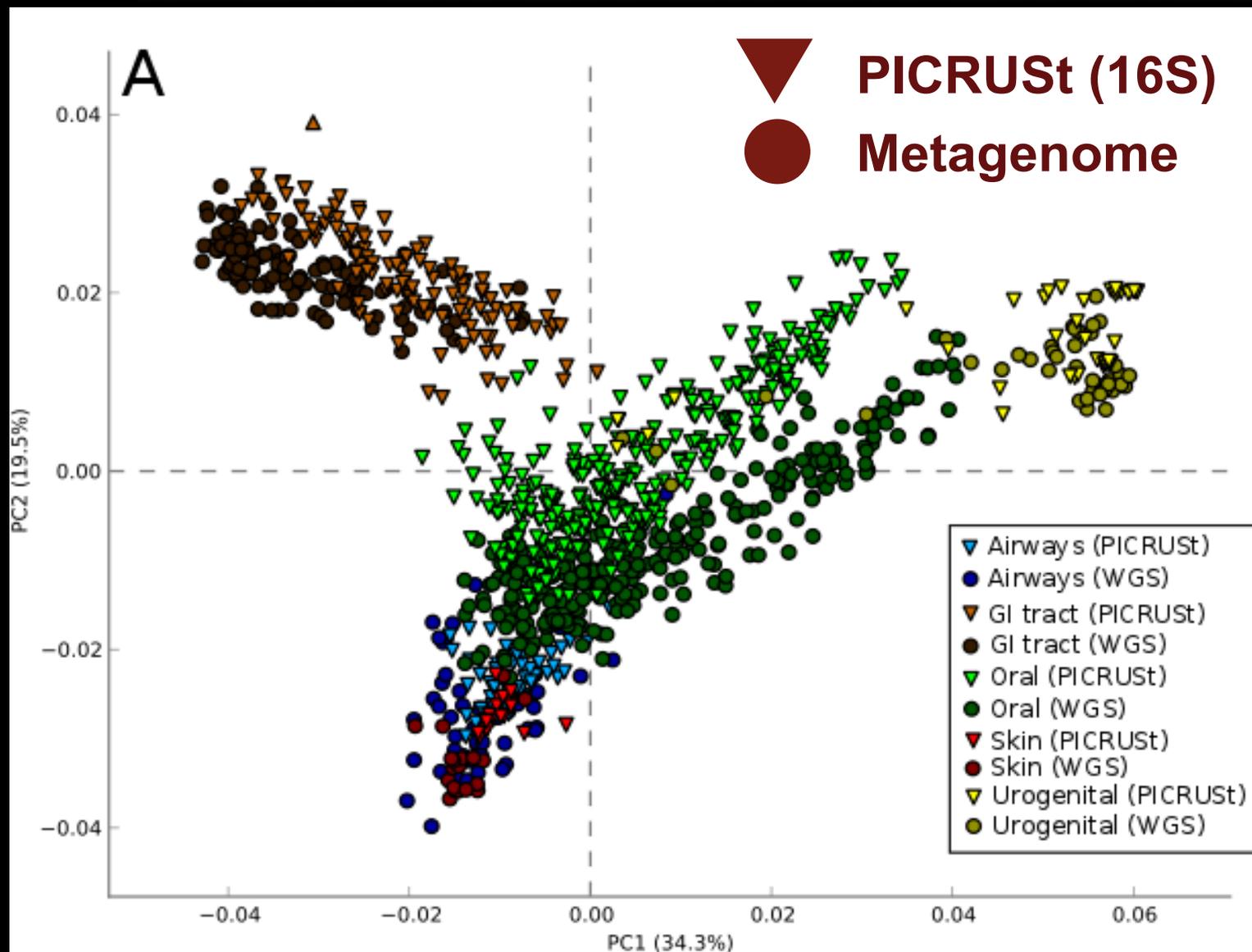
<http://picrust.github.com>

Pathways and modules      Orthologous gene families      Taxon abundances





It's not shotgun sequencing,  
but it's not too shabby, either



Ask *both* what you can do for your microbiome  
*and* what your microbiome can do for you





# Thanks!



## Human Microbiome Project

Owen White	Sahar Abubucker
Joe Petrosino	Brandi Cantarel
George Weinstock	Alyx Schubert
Karen Nelson	Mathangi Thiagarajan
Lita Proctor	Beltran Rodriguez-Mueller
Erica Sodergren	Makedonka Mitreva
Anthony Fodor	Yuzhen Ye
Marty Blaser	Mihai Pop
Jacques Ravel	Larry Forney
Pat Schloss	Barbara Methe

Bruce Birren Mark Daly  
Doyle Ward Ashlee Earl



Dirk Gevers  
Kat Huang



Ramnik Xavier  
Harry Sokol  
Dan Knights  
Moran Yassour



Daniela  
Boernigen



Vagheesh  
Narasimhan

Emma  
Schwager

Eric Franzosa



Joseph Moon

Jim Kaminski

Craig Bielski



Brian Palmer

Ren Lu

Hufeng Zhou



 Wendy Garrett  
Michelle Rooks



 Rob Beiko  
Morgan Langille



 Rob Knight  
Greg Caporaso  
Jesse Zaneveld



 Mark Silverberg  
Boyko Kabakchiev  
Andrea Tyler

 Ruth Ley  
Omry Koren

 Jacques Izard  
Katherine Lemon

 Bruce Sands