

Approaching multiple sequence alignment from a phylogenetic perspective

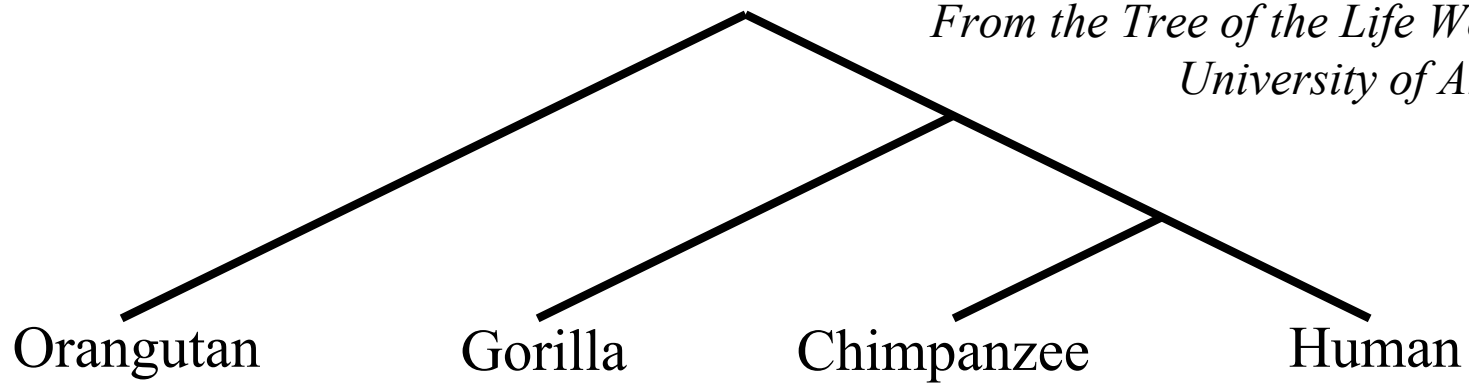
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The University of Texas at Austin

Species phylogeny

*From the Tree of the Life Website,
University of Arizona*



How did life evolve on earth?

An international effort to understand how life evolved on earth

Biomedical applications: drug design, protein structure and function prediction, biodiversity

**Phylogenetic estimation is a “Grand Challenge”:
millions of taxa, NP-hard optimization problems**



- Courtesy of the Tree of Life project

The CIPRES Project

(Cyber-Infrastructure for Phylogenetic Research)

www.phylo.org

This project is funded by the NSF under a Large ITR grant

- *ALGORITHMS and SOFTWARE: scaling to millions of sequences (open source, freely distributed)*
- *MATHEMATICS/PROBABILITY/STATISTICS: Obtaining better mathematical theory under complex models of evolution*
- *DATABASES: Producing new database technology for structured data, to enable scientific discoveries*
- *SIMULATIONS: The first million taxon simulation under realistically complex models*
- *OUTREACH: Museum partners, K-12, general scientific public*
- *PORTAL available to all researchers*

Step 1: Gather data

S1 = AGGCTATCACCTGACCTCCA

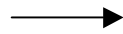
S2 = TAGCTATCACGACCGC

S3 = TAGCTGACCGC

S4 = TCACGACCGACA

Step 2: Multiple Sequence Alignment

S1 = AGGCTATCACCTGACCTCCA
S2 = TAGCTATCACGACCGC
S3 = TAGCTGACCGC
S4 = TCACGACCGACA



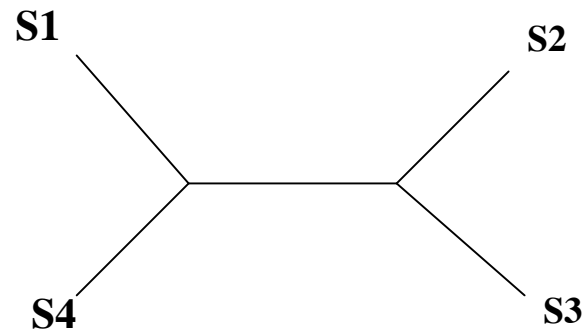
S1 = -AGGCTATCACCTGACCTCCA
S2 = TAG-CTATCAC--GACCGC--
S3 = TAG-CT-----GACCGC--
S4 = -----TCAC--GACCGACA

Step 3: Construct tree

S1 = AGGCTATCACCTGACCTCCA
S2 = TAGCTATCACGACCGC
S3 = TAGCTGACCGC
S4 = TCACGACCGACA

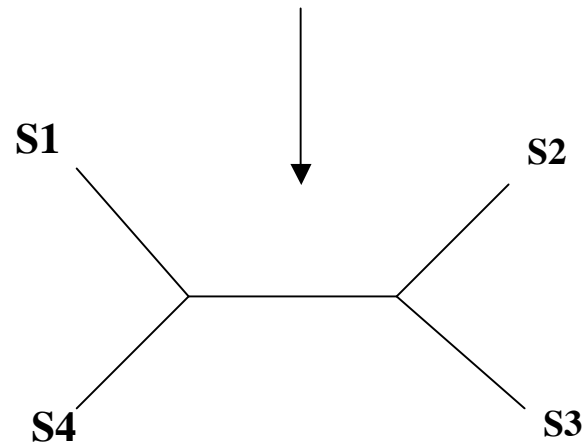


S1 = -AGGCTATCACCTGACCTCCA
S2 = TAG-CTATCAC--GACCGC--
S3 = TAG-CT-----GACCGC--
S4 = -----TCAC--GACCGACA



But molecular phylogenetics assumes the alignment is given

S1 = -AGGCTATCACCTGACCTCCA
S2 = TAG-CTATCAC--GACCGC--
S3 = TAG-CT-----GACCGC--
S4 = -----TCAC--GACCGACA



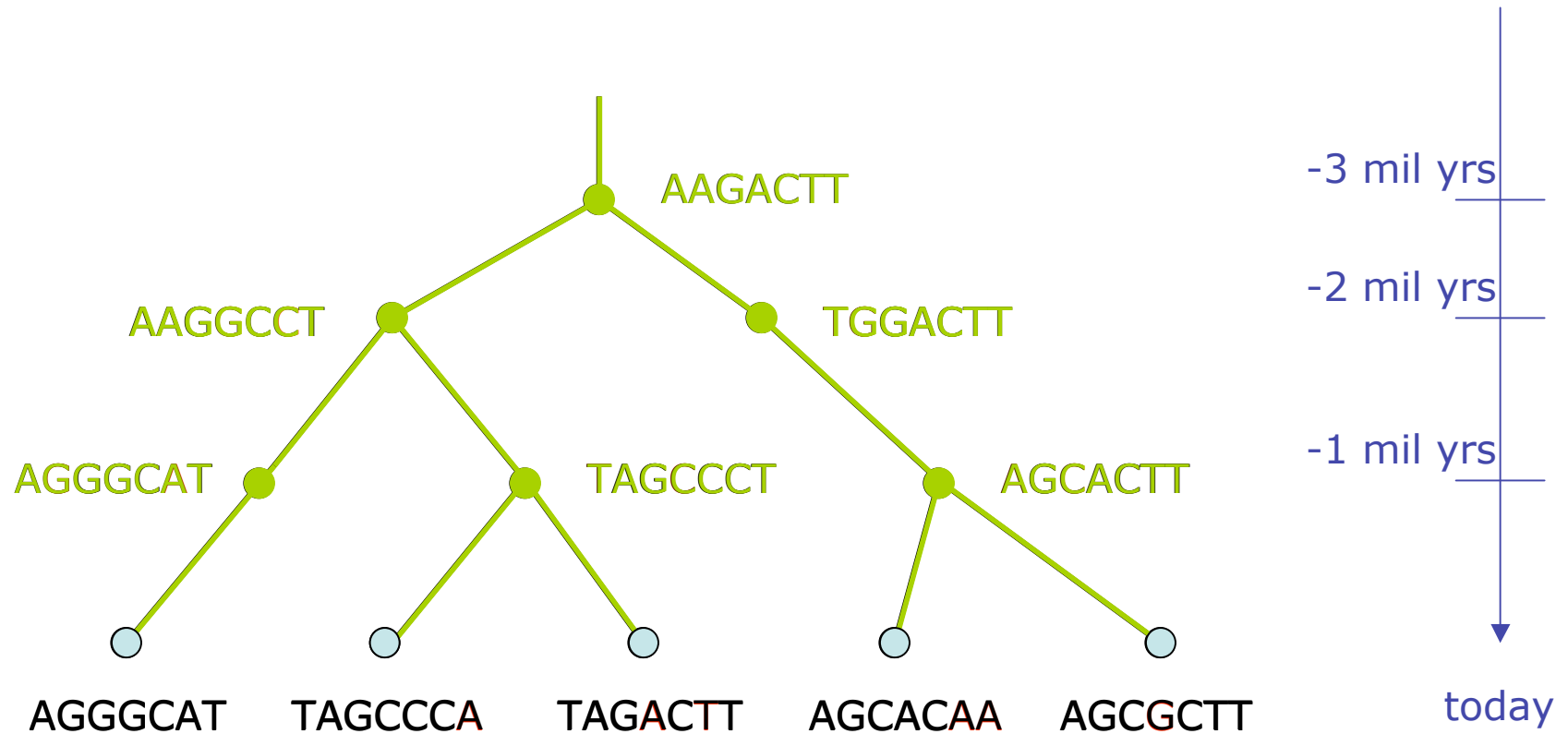
This talk

- **DCM-NJ**: Dramatic improvement in phylogeny estimation in terms of tree accuracy, and theoretical performance under Markov models of evolution
- **DCM-MP and DCM-ML**: Speeding up heuristics for large-scale phylogenetic estimation
- Simulation studies of two-phase methods (amino-acid and DNA sequences).
- **SATe**: A new technique for simultaneous estimation of trees and alignments

Performance criteria

- Estimated alignments are evaluated with respect to the *true alignment*. Studied both in simulation and on real data.
- Estimated trees are evaluated for “topological accuracy” with respect to the *true tree*. Typically studied in simulation.
- Methods for these problems can also be evaluated with respect to an optimization criterion (e.g., maximum likelihood score) as a function of running time. Typically studied on real data. (Reasonably valid for phylogeny but not yet for alignment.)

DNA Sequence Evolution

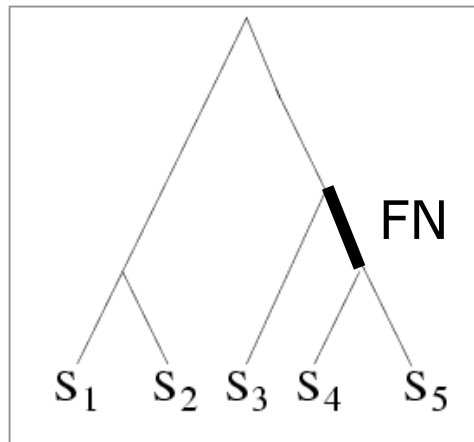


Markov models of single site evolution

Simplest (Jukes-Cantor):

- The model tree is a pair $(T, \{e, p(e)\})$, where T is a rooted binary tree, and $p(e)$ is the probability of a substitution on the edge e .
- The state at the root is random.
- If a site changes on an edge, it changes with equal probability to each of the remaining states.
- The evolutionary process is Markovian.

More complex models (such as the General Markov model) are also considered, often with little change to the theory.

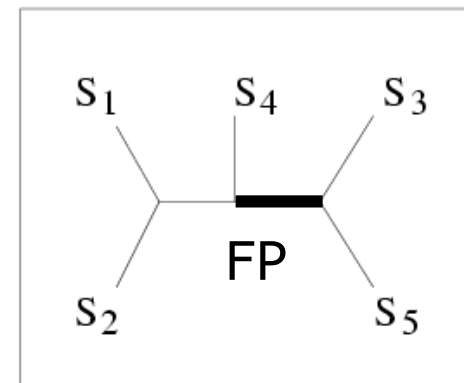


TRUE TREE



S_1	ACAATTAGAAC
S_2	ACCCTTAGAAC
S_3	ACCATTCCAAC
S_4	ACCAGACCAAC
S_5	ACCAGACCGGA

DNA SEQUENCES

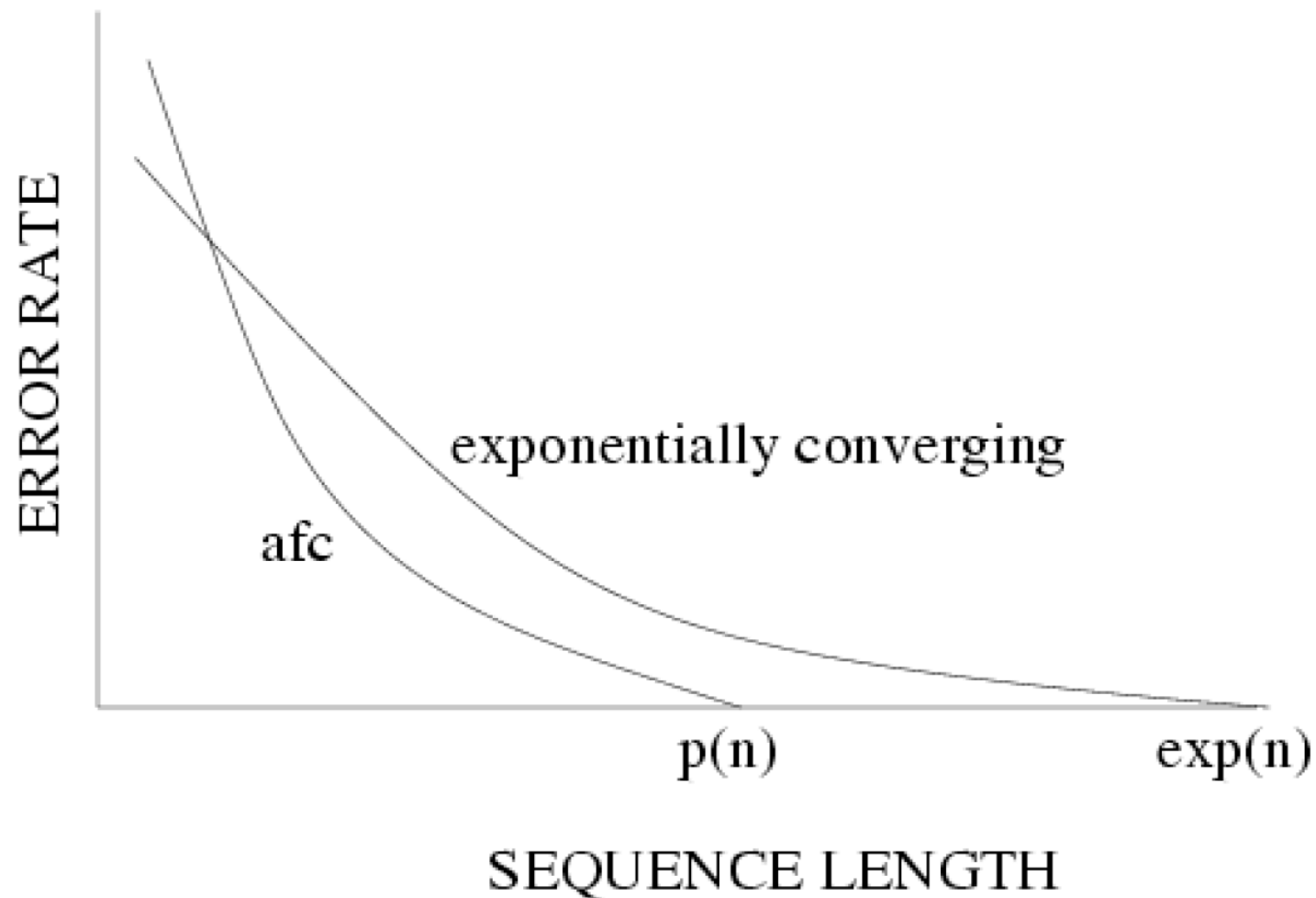


INFERRED TREE

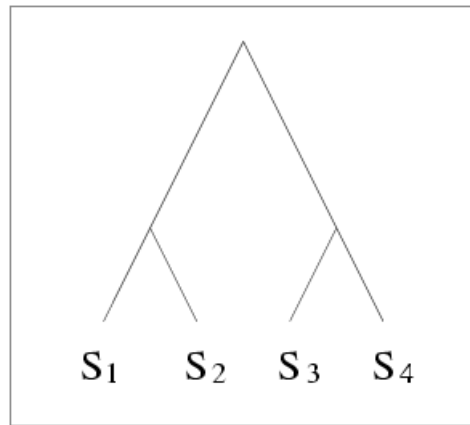
FN: false negative
(missing edge)
FP: false positive
(incorrect edge)

50% error rate

Statistical consistency, exponential convergence, and absolute fast convergence (afc)



Distance-based Phylogenetic Methods

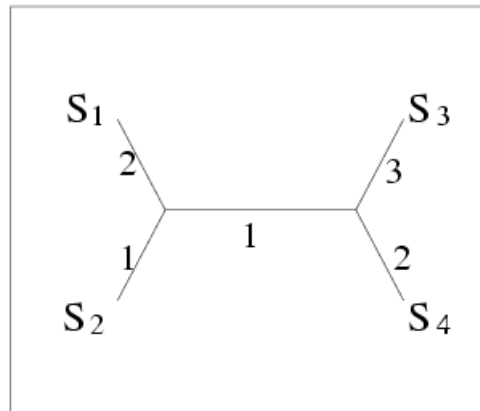


TRUE TREE

S₁ ACAATTAGAAC
S₂ ACCCTTAGAAC
S₃ ACCATTCCAAC
S₄ ACCAGACCAAC

DNA SEQUENCES

STATISTICAL
ESTIMATION
OF PAIRWISE
DISTANCES



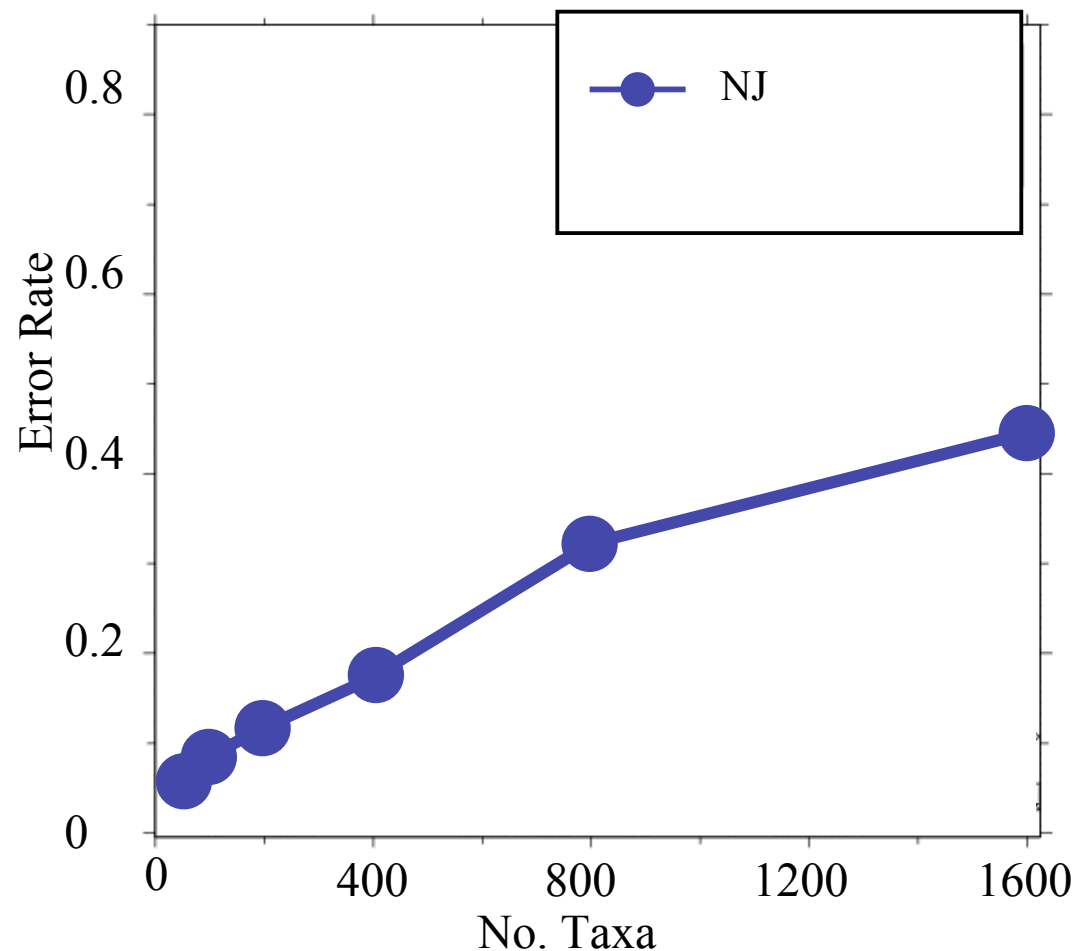
INFERRED TREE

METHODS
SUCH AS
NEIGHBOR
JOINING

	S ₁	S ₂	S ₃	S ₄
S ₁	0	3	6	5
S ₂		0	5	4
S ₃			0	5
S ₄				0

DISTANCE MATRIX

Neighbor joining (although statistically consistent)
has poor performance on large diameter trees
[Nakhleh et al. ISMB 2001]

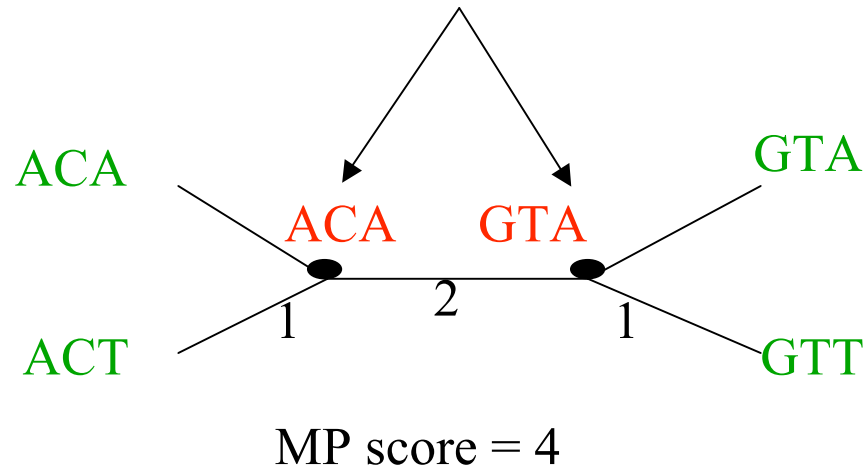


Simulation study based upon fixed edge lengths, K2P model of evolution, sequence lengths fixed to 1000 nucleotides.

Error rates reflect proportion of incorrect edges in inferred trees.

Maximum Parsimony:

Optimal labeling on a fixed tree can be computed in linear time $O(nk)$



MP is not statistically consistent.
Finding the optimal MP tree is **NP-hard**.

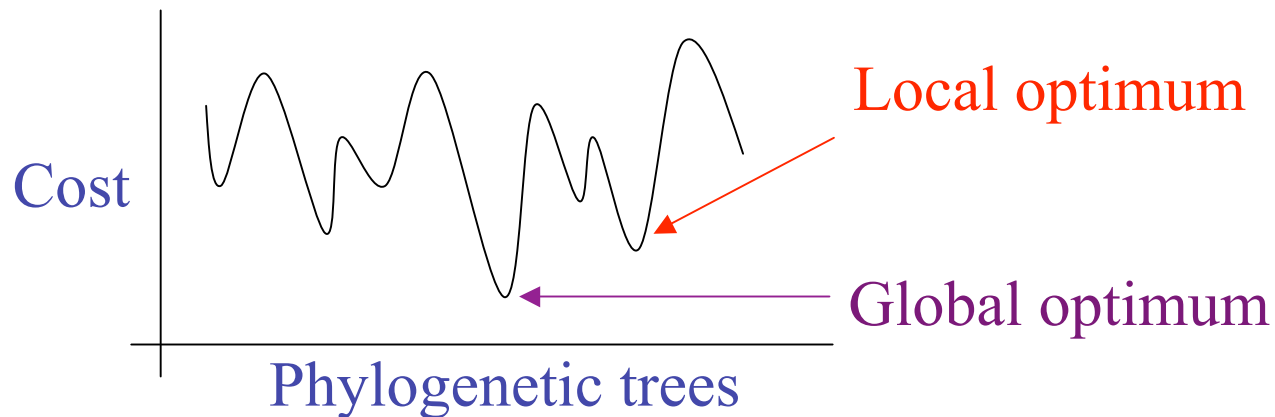
Maximum Likelihood (ML)

- Given: stochastic model of sequence evolution (e.g. Jukes-Cantor) and a set S of sequences
- Objective: Find tree T and parameter values so as to maximize the probability of the data.

NP-hard, but statistically consistent. Preferred by many systematists, but even harder than MP in practice. (Exponential sequence lengths suffice for accuracy with high probability.)

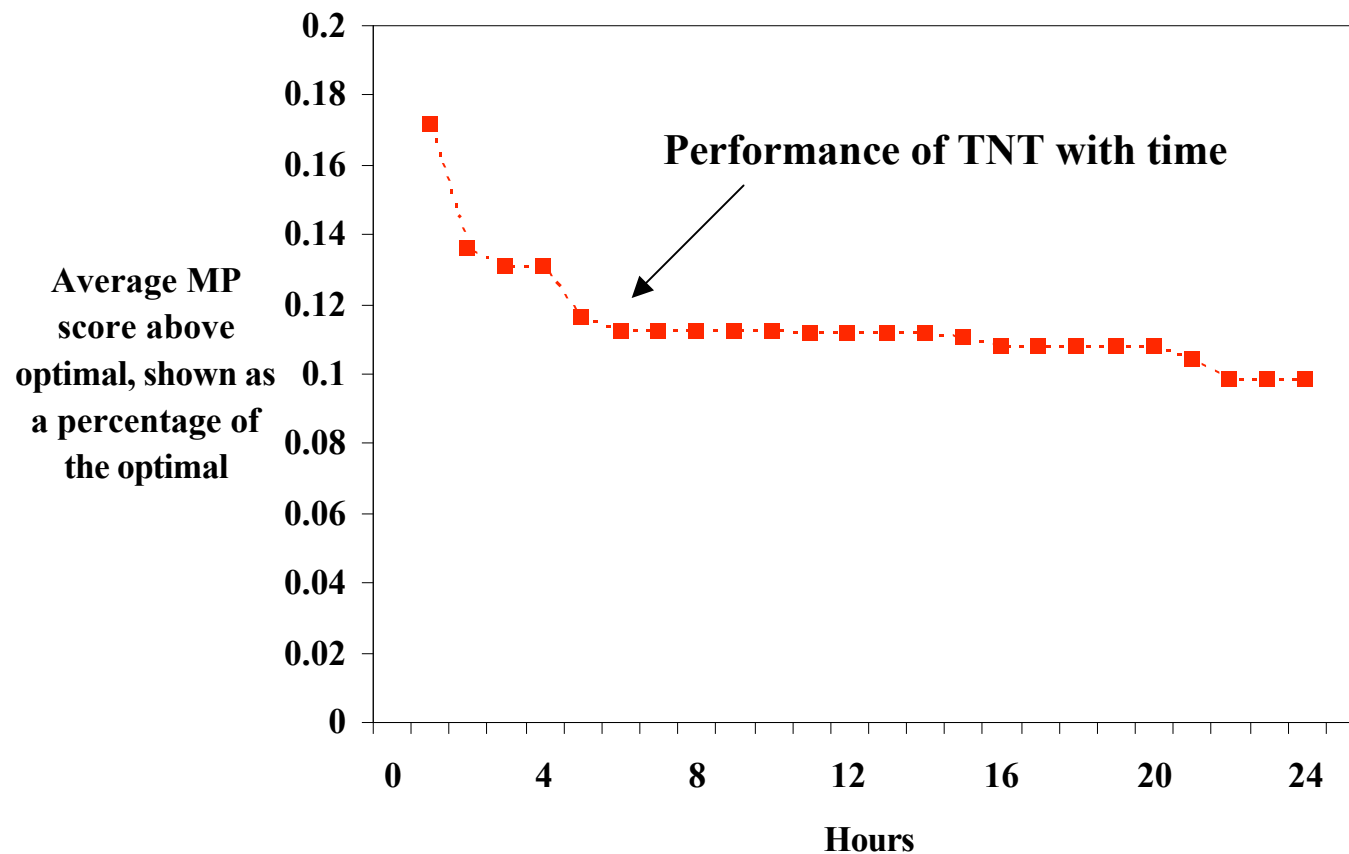
Approaches for “solving” MP (and other NP-hard problems in phylogeny)

1. Hill-climbing heuristics (which can get stuck in local optima)
2. Randomized algorithms for getting out of local optima
3. Approximation algorithms for MP (based upon Steiner Tree approximation algorithms).



Problems with current techniques for MP and ML

Shown here is the performance of a very good heuristic (TNT) for maximum parsimony analysis on a real dataset of almost 14,000 sequences. (“Optimal” here means *best score to date*, using any method for any amount of time.) Acceptable error is below 0.01%.



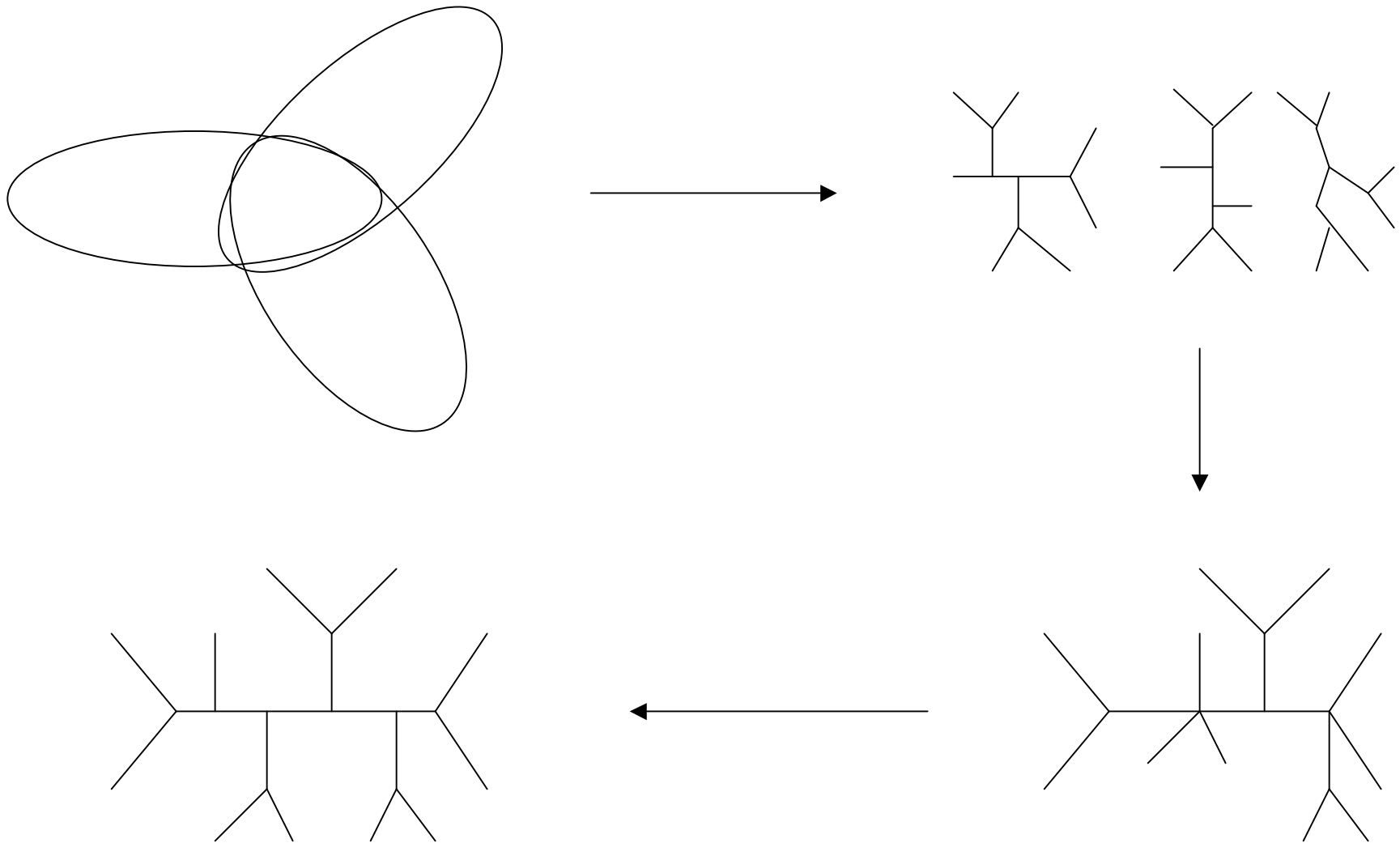
Problems with existing phylogeny reconstruction methods

- Polynomial time methods (generally based upon distances) have poor accuracy with large diameter datasets.
- Heuristics for NP-hard optimization problems take too long (months to reach acceptable local optima).

Warnow et al.: Meta-algorithms for phylogenetics

- Basic technique: determine the conditions under which a phylogeny reconstruction method does well (or poorly), and design a divide-and-conquer strategy to improve the performance
- The divide-and-conquer technique is specific to the method.
- Warnow et al. developed a class of divide-and-conquer methods, collectively called DCMs (**Disk-Covering Methods**).

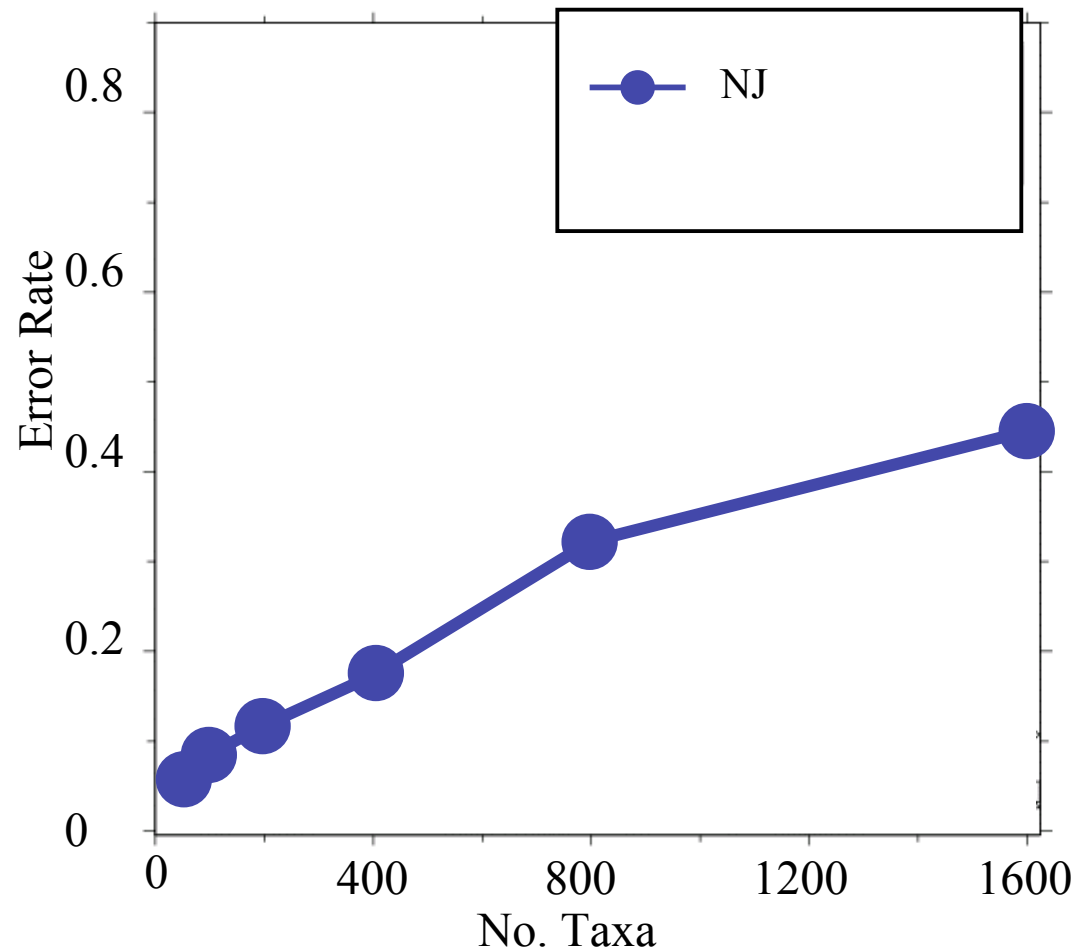
Disk-Covering Method (DCM)



Improving phylogeny reconstruction methods using DCMs

- Improving the theoretical convergence rate and performance of polynomial time distance-based methods using DCM1
- Speeding up heuristics for NP-hard optimization problems (Maximum Parsimony and Maximum Likelihood) using Rec-I-DCM3

Neighbor joining (although statistically consistent)
has poor performance on large diameter trees
[Nakhleh et al. ISMB 2001]

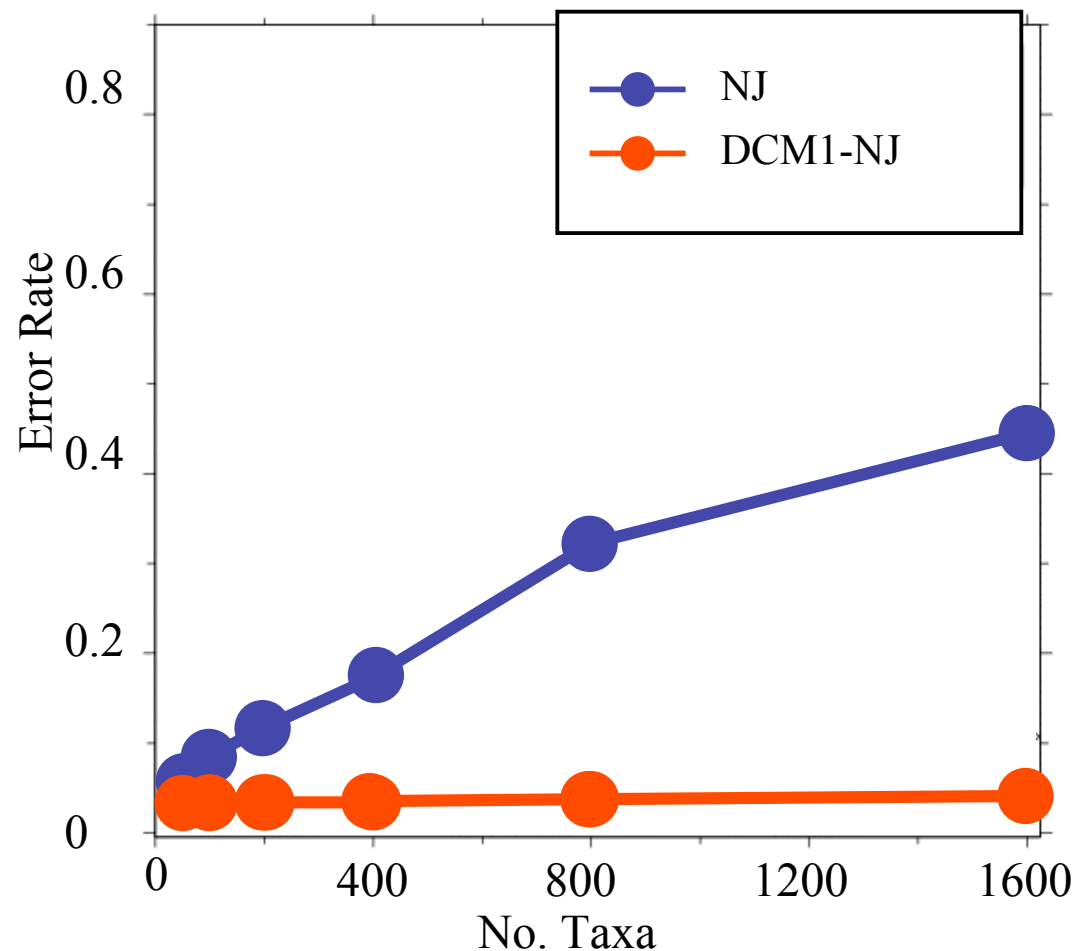


Simulation study based upon fixed edge lengths, K2P model of evolution, sequence lengths fixed to 1000 nucleotides.

Error rates reflect proportion of incorrect edges in inferred trees.

DCM1-boosting distance-based methods

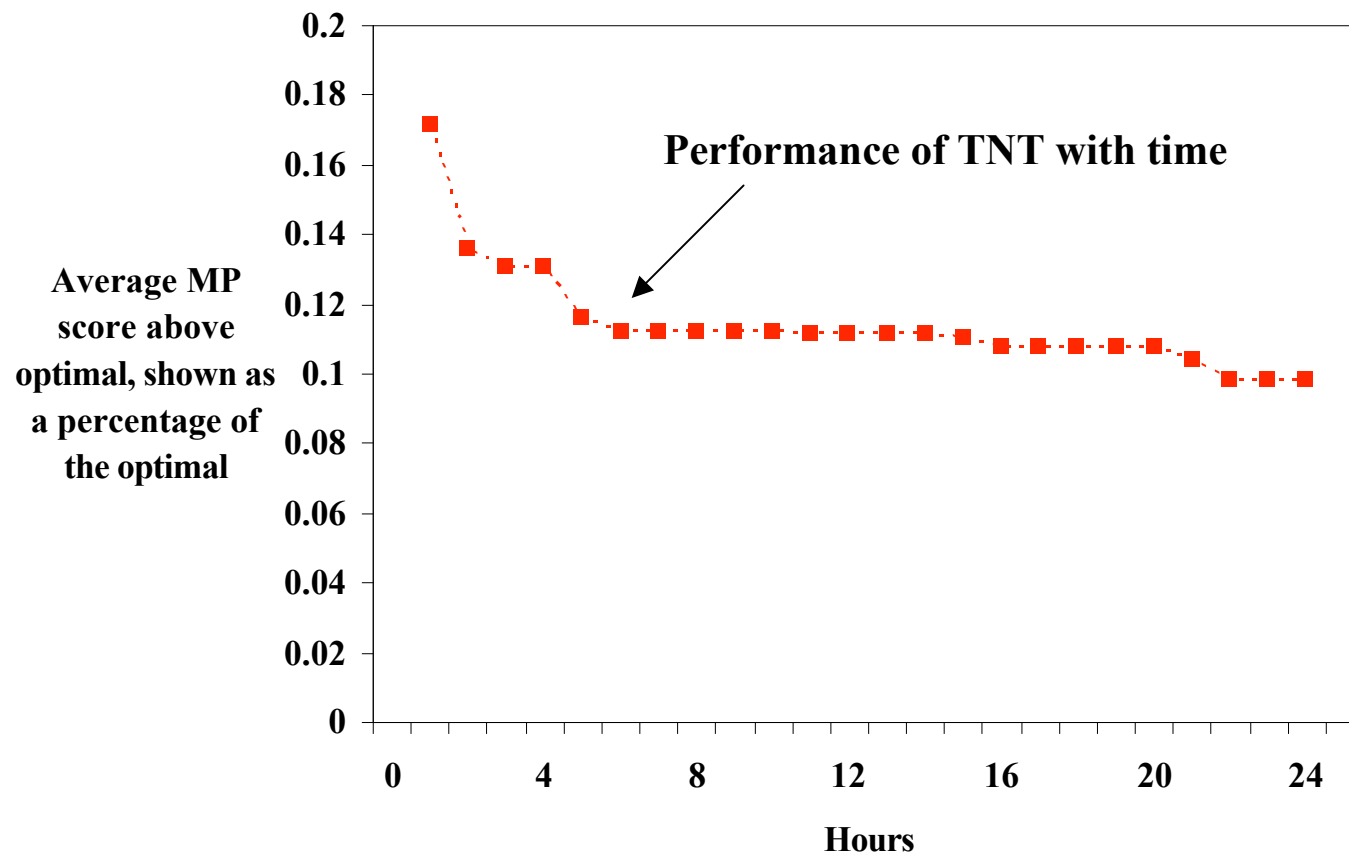
[Nakhleh et al. ISMB 2001]



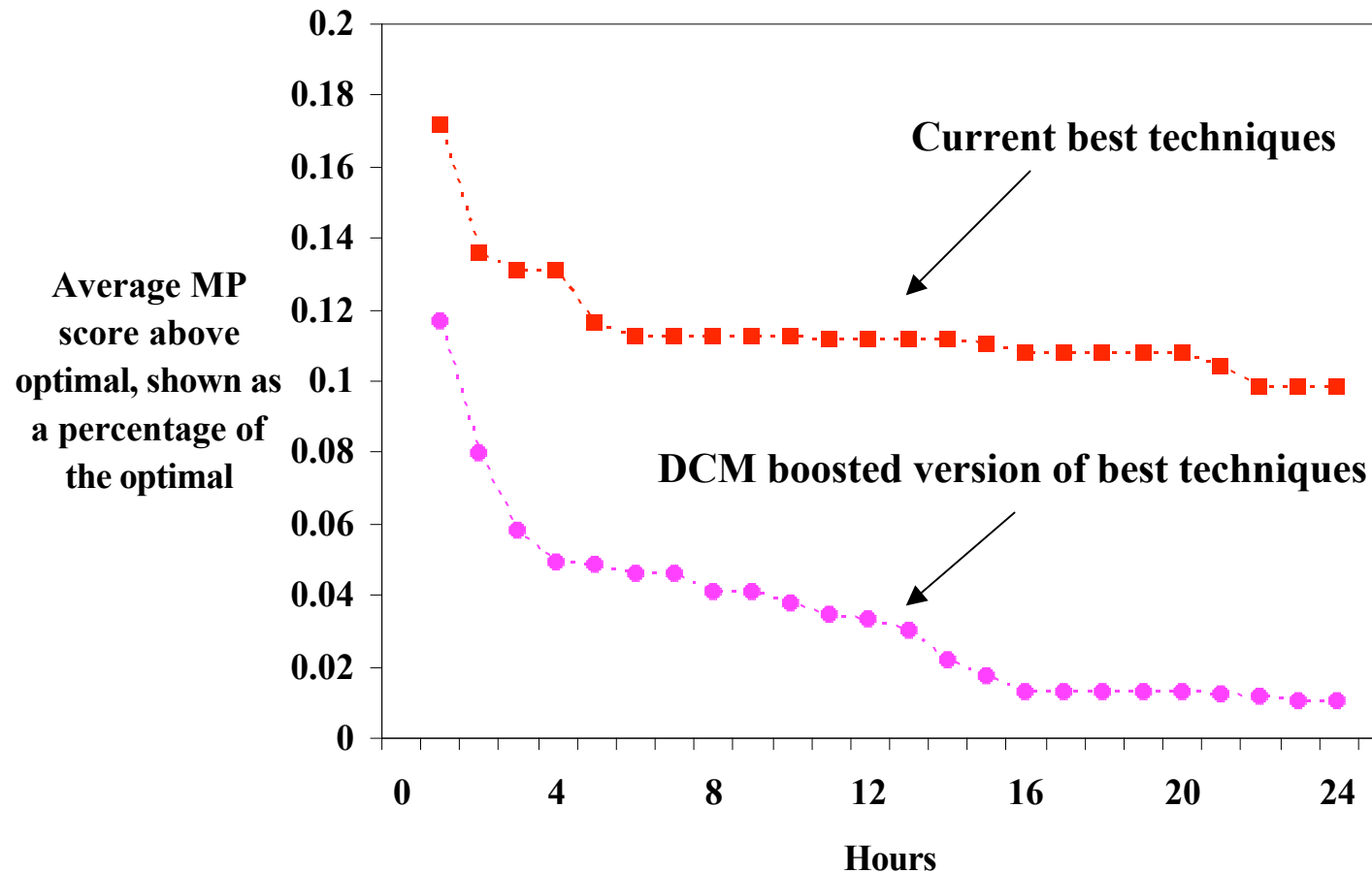
Theorem:
DCM1-NJ
converges to the
true tree from
polynomial
length sequences

Problems with current techniques for MP

Shown here is the performance of a TNT heuristic maximum parsimony analysis on a real dataset of almost 14,000 sequences. (“Optimal” here means *best score to date*, using any method for any amount of time.) Acceptable error is below 0.01%.



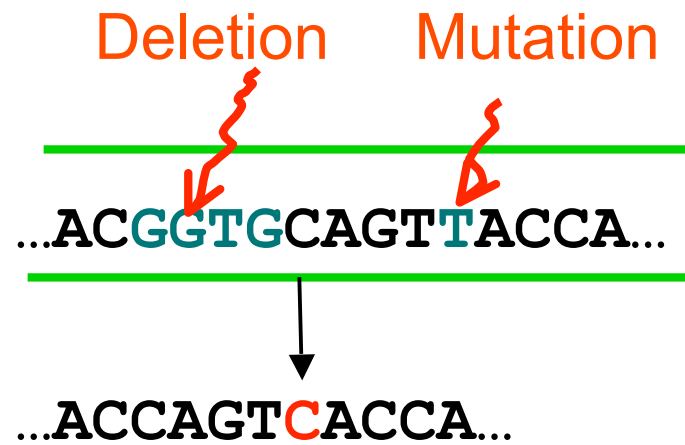
Rec-I-DCM3 significantly improves performance (Roshan et al.)



Comparison of TNT to Rec-I-DCM3(TNT) on one large dataset

Very nice, but...

- *Evolution is not as simple as these models assert!*



indels (insertions and deletions) also occur!

Basic Questions

- Does improving the alignment lead to an improved phylogeny?
- Are we getting good enough alignments from MSA methods?
- Are we getting good enough trees from the phylogeny reconstruction methods?
- Can we improve these estimations, perhaps through **simultaneous estimation** of trees and alignments?

Multiple Sequence Alignment

AGGCTATCACCTGACCTCCA	-AGGCTATCACCTGACCTCCA
TAGCTATCACGACCGC	TAG-CTATCAC--GACCGC--
TAGCTGACCGC	TAG-CT-----GACCGC--

Notes:


1. We insert gaps (dashes) to each sequence to make them “line up”.
2. Nucleotides in the same column are presumed to have a common ancestor (i.e., they are “homologous”).

Indels and substitutions at the DNA level

...ACGGTGCAGTTACCA...

Indels and substitutions at the DNA level

Deletion Mutation



...ACGGTGCAGTTACCA...

The diagram illustrates a DNA sequence with two specific changes highlighted. Above the sequence, the word "Deletion" is written in orange, with a red arrow pointing to the second 'G' in the "ACGGTGCAGTTACCA" sequence. To the right, the word "Mutation" is also written in orange, with a red arrow pointing to the 'T' in the "TACCA" portion of the sequence. The DNA sequence itself is written in black, with the "GGT" and "T" characters highlighted in teal to draw attention to the sites of the deletion and mutation respectively.

Indels and substitutions at the DNA level

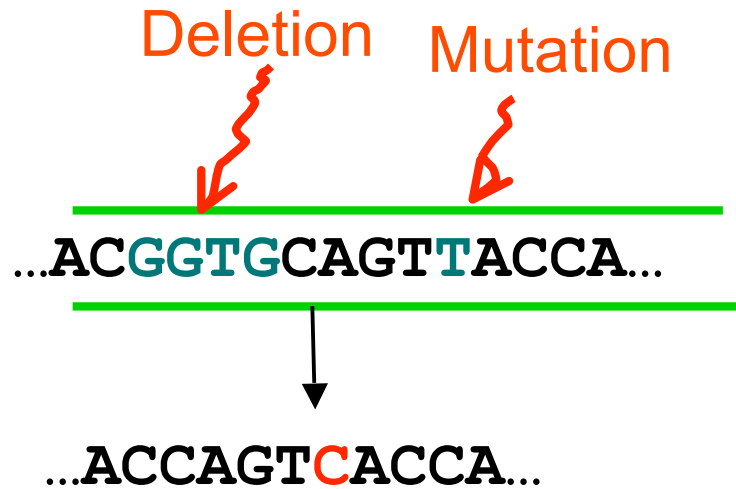
Deletion Mutation



...ACGGTGCAGTTACCA...

...ACCAGTCACCA...

The diagram illustrates a DNA sequence change. The top sequence is "...ACGGTGCAGTTACCA...". The bottom sequence is "...ACCAGTCACCA...". A red arrow points from the 'G' in the top sequence to the 'C' in the bottom sequence, indicating a deletion. A red arrow points from the 'T' in the top sequence to the 'C' in the bottom sequence, indicating a mutation.



The **true** pairwise alignment is:

...ACGGTGCAGTTACCA...

...AC-----CAGTCACCA...

The **true multiple alignment** on a set of homologous sequences is obtained by tracing their evolutionary history, and extending the pairwise alignments on the edges to a multiple alignment on the leaf sequences.

Basics about alignments

- The standard alignment method for phylogeny is Clustal (or one of its derivatives), but many new alignment methods have been developed by the protein alignment community.
- Alignments are generally evaluated in comparison to the “true alignment”, using the SP-score (percentage of truly homologous pairs that show up in the estimated alignment).
- On the basis of SP-scores (and some other criteria), methods like **ProbCons**, **Mafft**, and **Muscle** are generally considered “better” than Clustal.

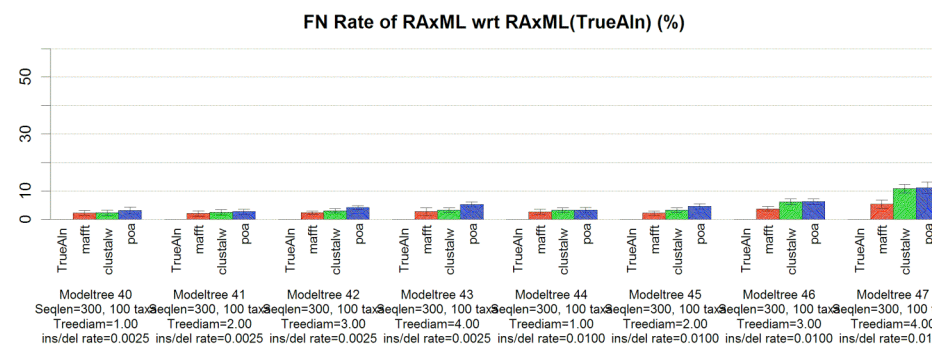
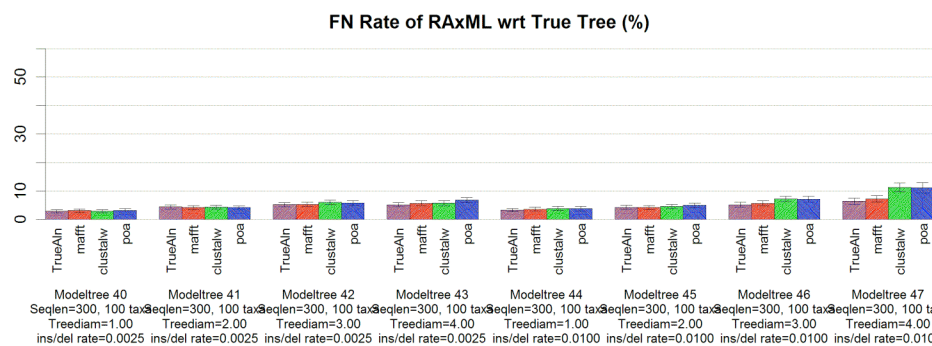
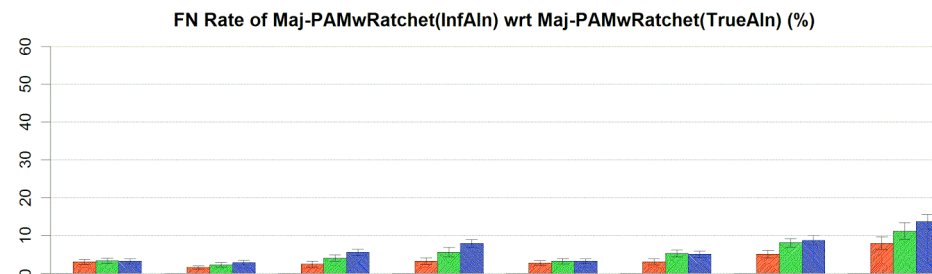
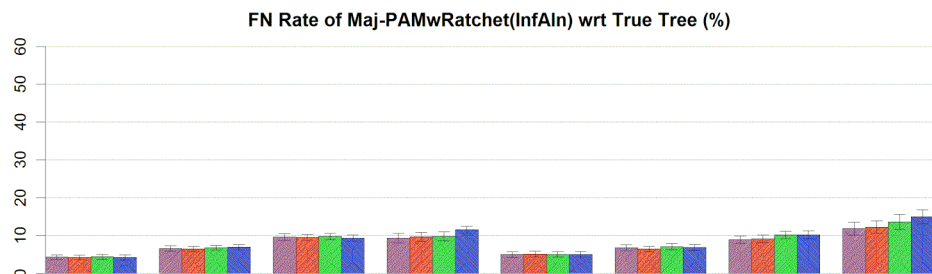
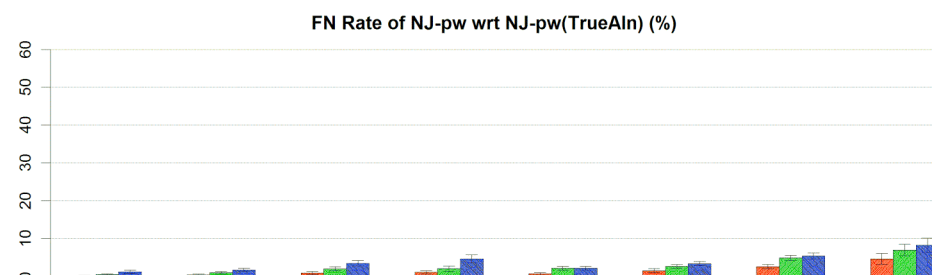
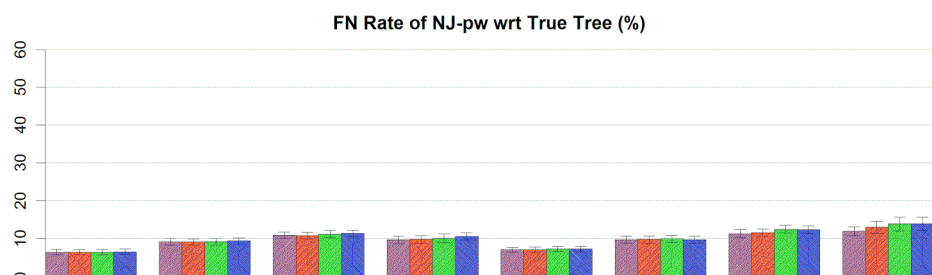
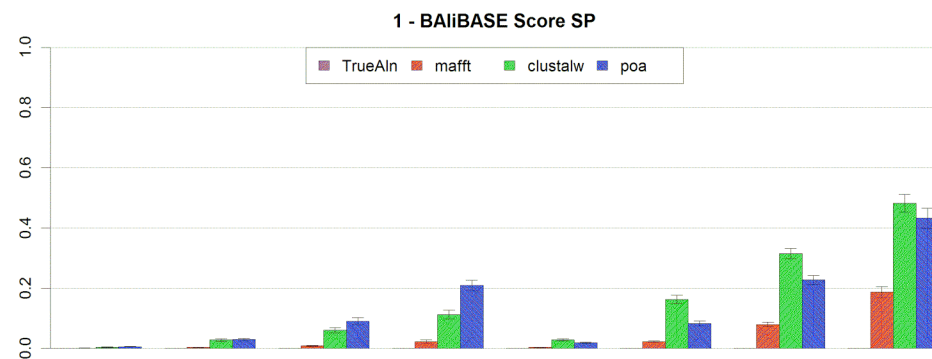
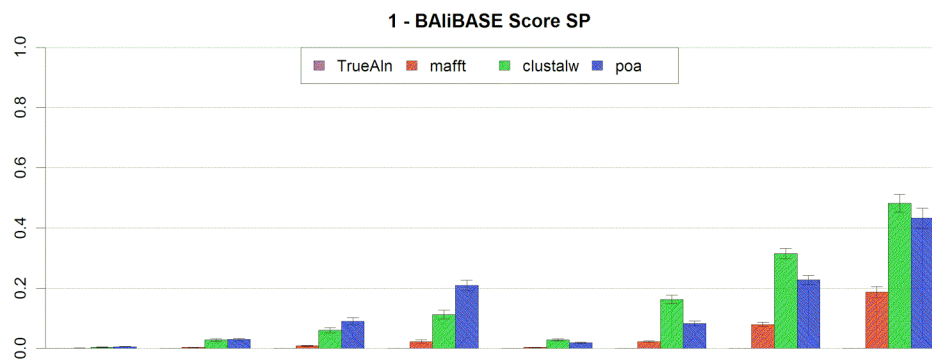
Questions

- Many new MSA methods improve on ClustalW on biological benchmarks (e.g., BaliBASE) and in simulation. Does this lead to improved phylogenetic estimations?
- The phylogeny community has tended to assume that alignment has a big impact on final phylogenetic accuracy. But does it? Does this depend upon the model conditions?
- What are the best two-phase methods?

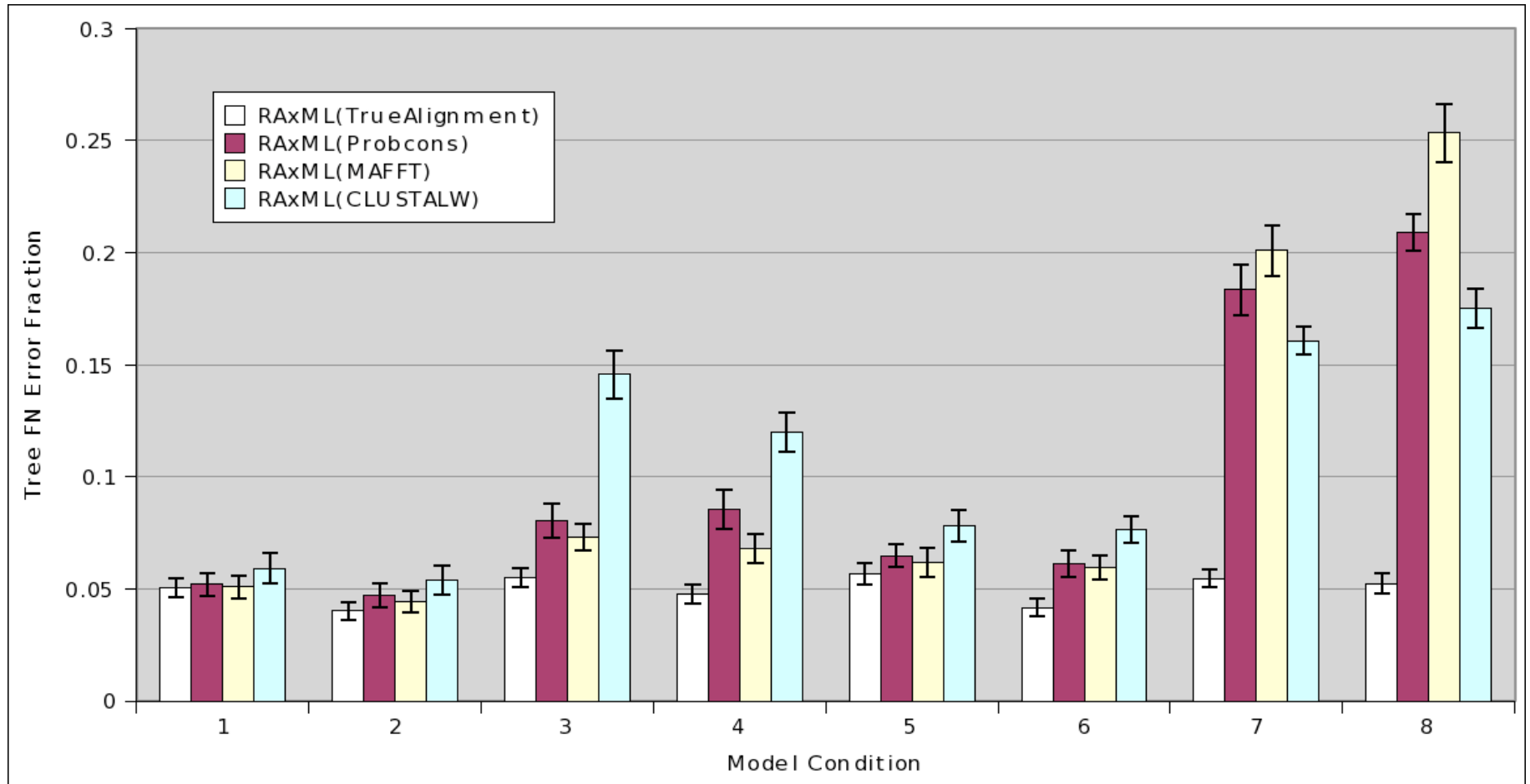
Our simulation studies (both using ROSE*)

- Amino-acid evolution (Wang et al., unpublished): Model trees based upon BaliBase and random birth-death trees, with 12 taxa to 100 taxa. Average gap length 3.4. Average identity ranges from 23% to 57%. Average gappiness ranges from 3% to 60%.
- DNA sequence evolution (Liu et al., unpublished): Random birth-death trees, 25 to 500 taxa. Two gap length distributions (short and long). Average p-distance ranges from 43% to 63%. Average gappiness ranges from 40% to 80%.

**ROSE has limitations!*



Non-coding DNA evolution



Models 1-4 have “long gaps”, and models 5-8 have “short gaps”

Observations

- Phylogenetic tree accuracy is positively correlated with alignment accuracy (measured using SP), but the degree of improvement in tree accuracy is *much smaller*. (However, note that sometimes “worse” alignments yield better trees.)
- The best two-phase methods are generally (but not always!) obtained by using either ProbCons or MAFFT, followed by ML.
- However, even the best two-phase methods don’t do well enough.

Two problems with two-phase methods

- All current methods for multiple alignment have high error rates when sequences evolve with many indels and substitutions.
- All current methods for phylogeny estimation treat indel events inadequately (either treating as missing data, or giving too much weight to each gap).

Simultaneous estimation?

- Statistical methods (e.g., AliFritz and BaliPhy) cannot be applied to datasets above ~ 20 sequences.
- POY (Wheeler et al.) attempts to find tree/alignment pairs of minimum total edit distance. POY can be applied to larger datasets, but has not performed as well as the best two-phase methods.

SATe:

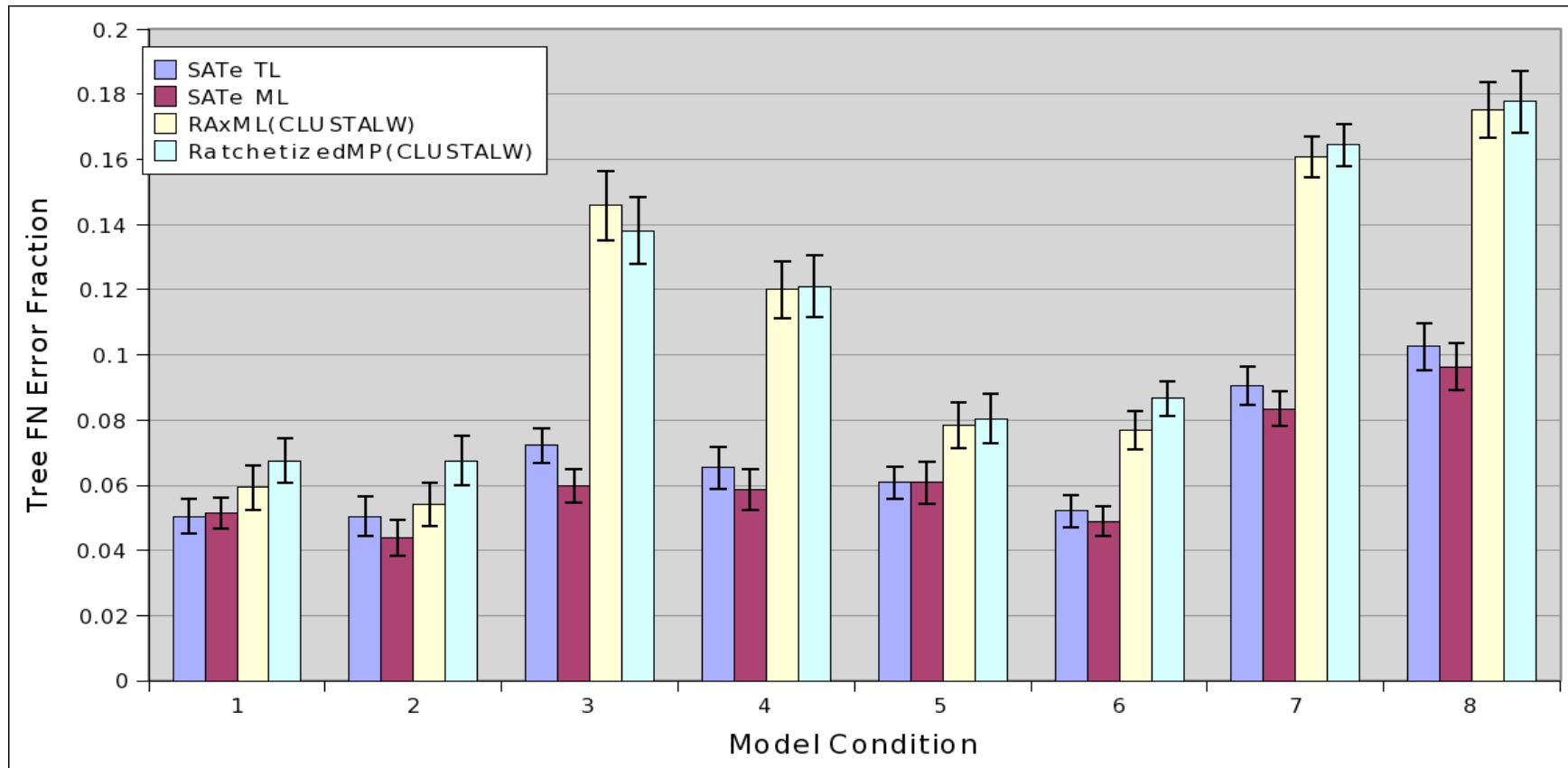
(Simultaneous Alignment and Tree Estimation)

- Developers: Warnow, Linder, Liu, Nelesen, and Zhao.
- Search strategy: search through tree space, and align sequences on each tree by *heuristically estimating ancestral sequences*.
- **SATe-TL** is the alignment/tree pair of minimum total length (default affine gap penalty has gap-open cost 2, gap-extend cost 0.5 and mismatch cost 1).
- **SATe-ML** is the alignment/tree pair that optimizes maximum likelihood under GTR+Gamma+I.

Simulation study

- 100 taxon model trees (generated by r8s and then modified, so as to deviate from the molecular clock).
- DNA sequences evolved under ROSE (indel events of blocks of nucleotides, plus HKY site evolution). The root sequence has 1000 sites.
- We vary the gap length distribution, probability of gaps, and probability of substitutions, to produce 8 model conditions: models 1-4 have “long gaps” and 5-8 have “short gaps”.

SATe-TL vs. SATe-ML vs. Clustal



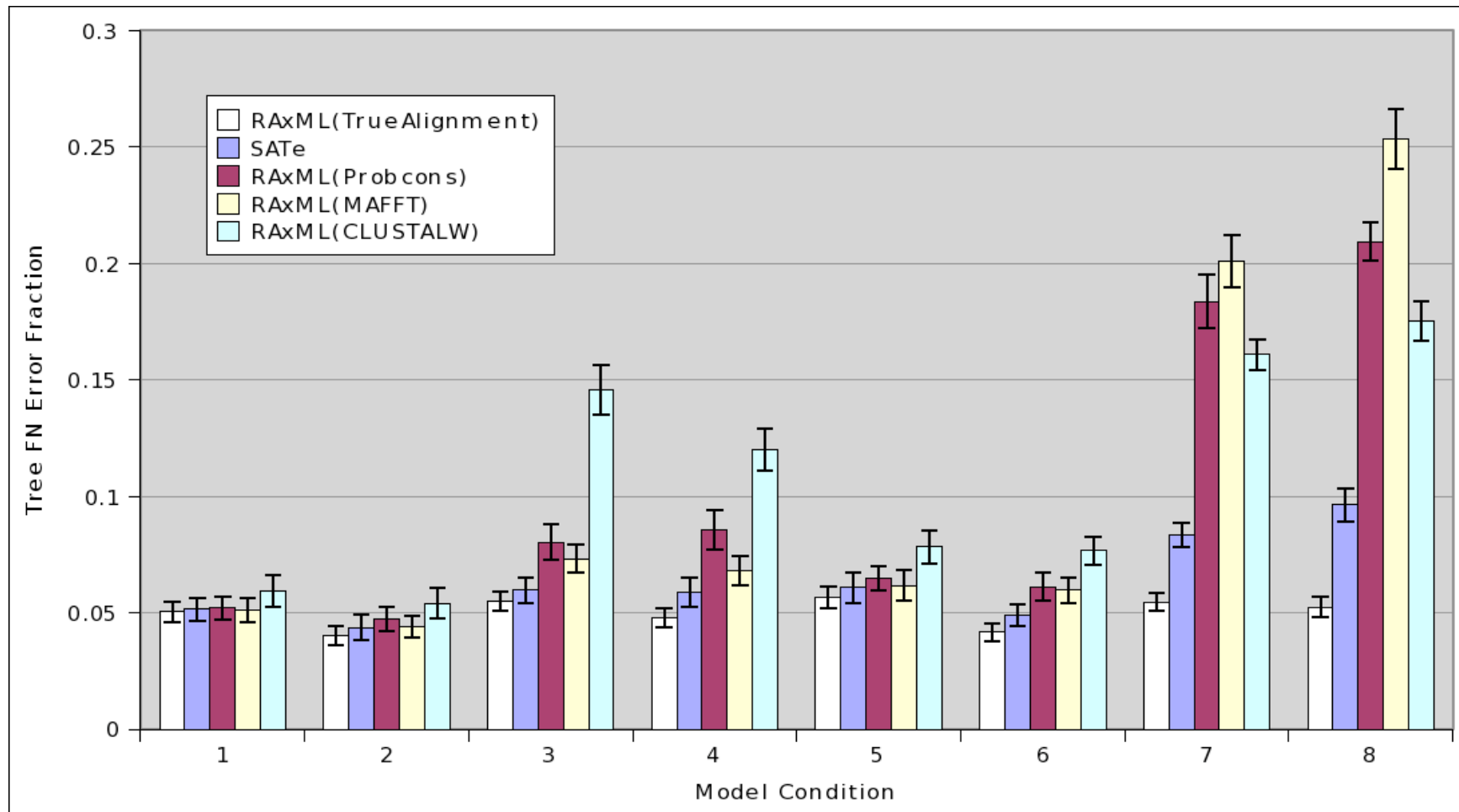
- Model conditions 1-4 have long gaps (100 taxa)
- Model conditions 5-8 have short gaps (100 taxa)

Comments

- *The affine gap penalty matters! (Performance of SATe-TL for simple gap penalties is not as good.)*
- SATe-ML produces trees that are more accurate than SATe-TL, and both improve upon trees estimated using Clustal (contrary to Ogden and Rosenberg's 2007 Systematic Biology paper).

What about comparisons to other two-phase methods?

Our method (SATe-ML) vs. other methods

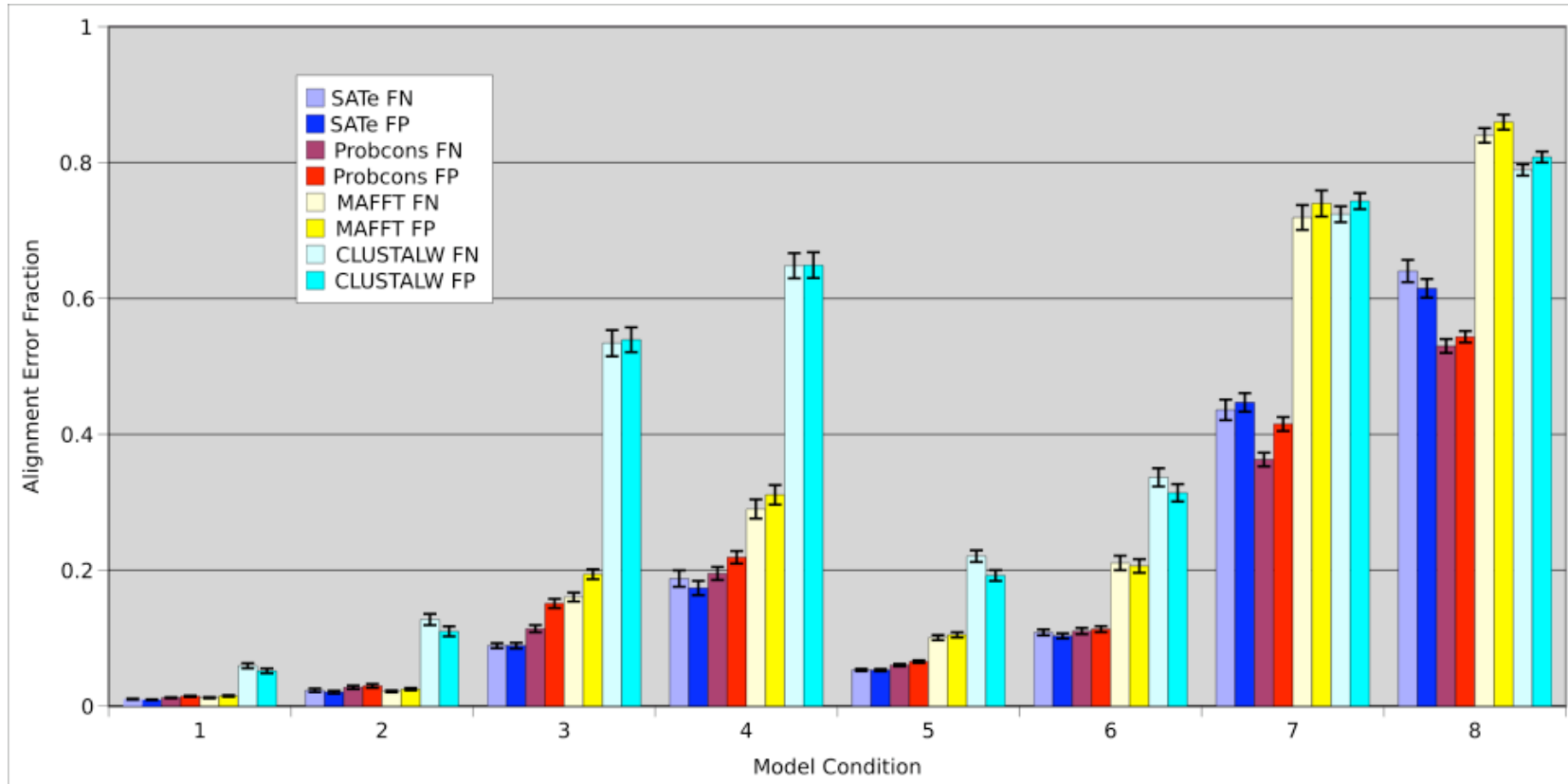


- Long gap models 1-4, Short gap models 5-8

Alignment accuracy

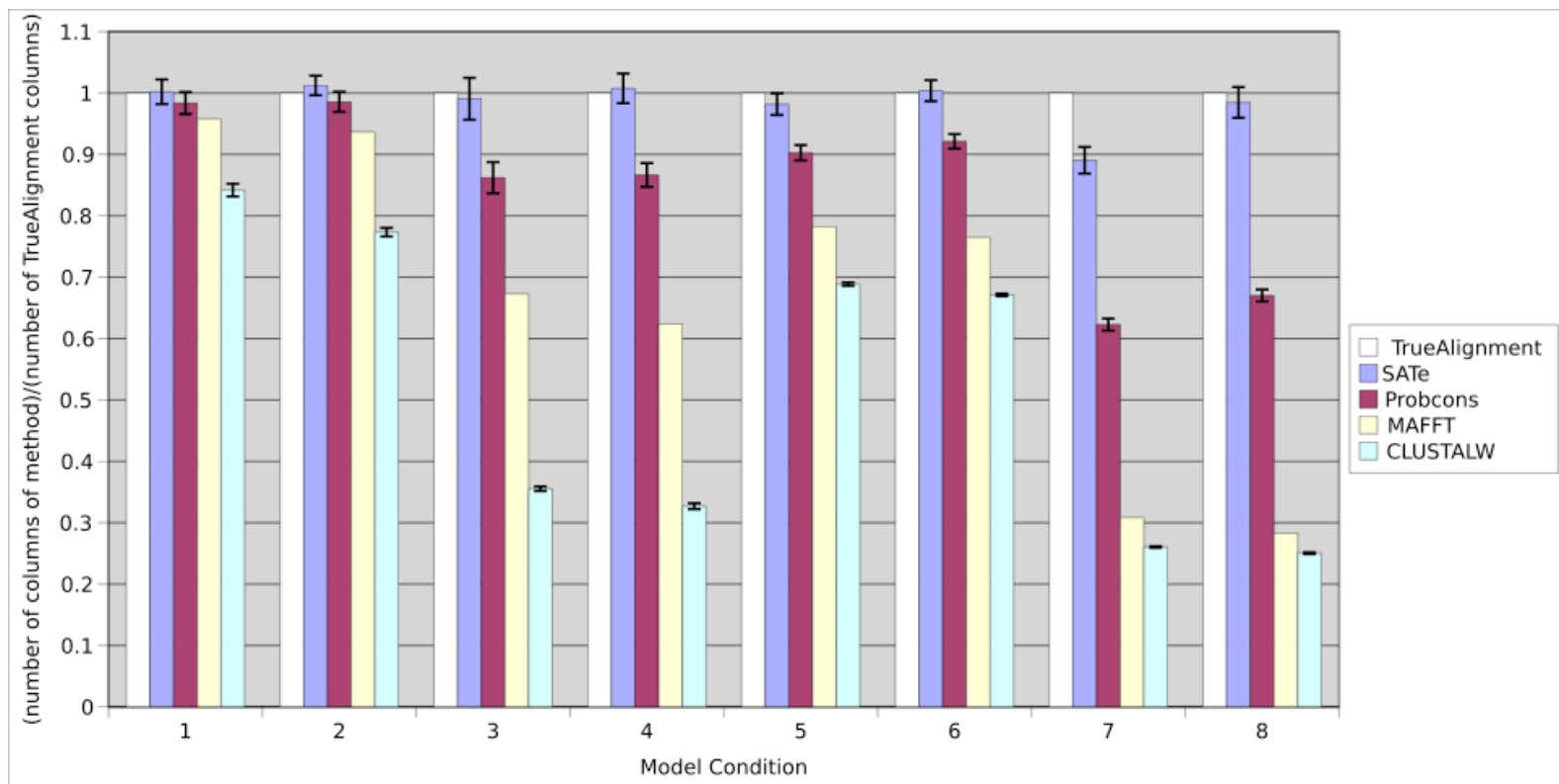
FN: proportion of correctly homologous pairs of nucleotides *missing* from the estimated alignment (i.e., 1-SP score).

FP: proportion of incorrect pairings of nucleotides in the estimated alignment.



Alignment length accuracy

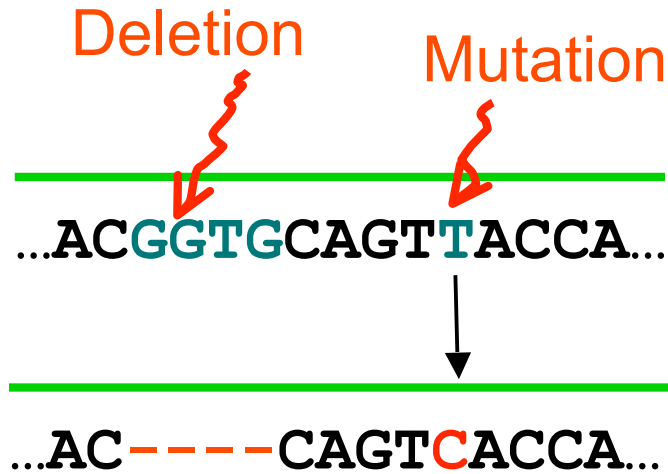
- Normalized number of columns in the estimated alignment relative to the true alignment.



Summary about SATe

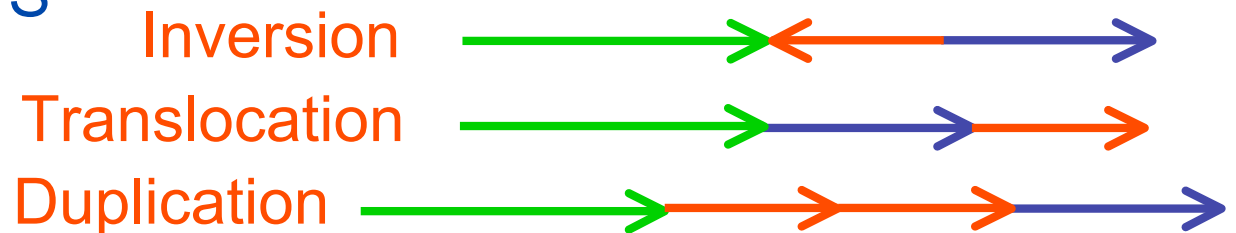
- SATe-ML improves upon the two-phase techniques we studied with respect to tree accuracy, and with respect to alignment length. (Alignment accuracy, measured using SP, varies.)
- SATe-TL is not as accurate as SATe-ML, but is fairly close in accuracy. The affine gap penalty impacts alignment and tree accuracy!
- SATe is slow, so that performance depends upon how long you run it (these experiments limited to 48 hours).

(but evolution is more complicated than that!)



SEQUENCE EDITS

REARRANGEMENTS



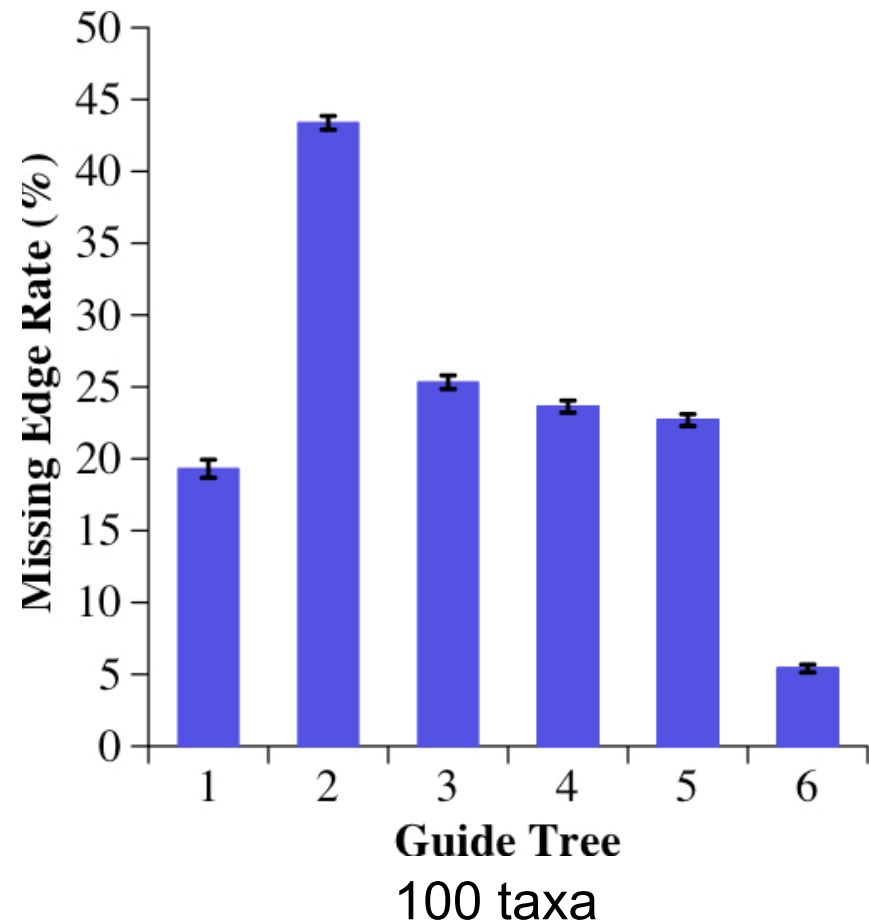
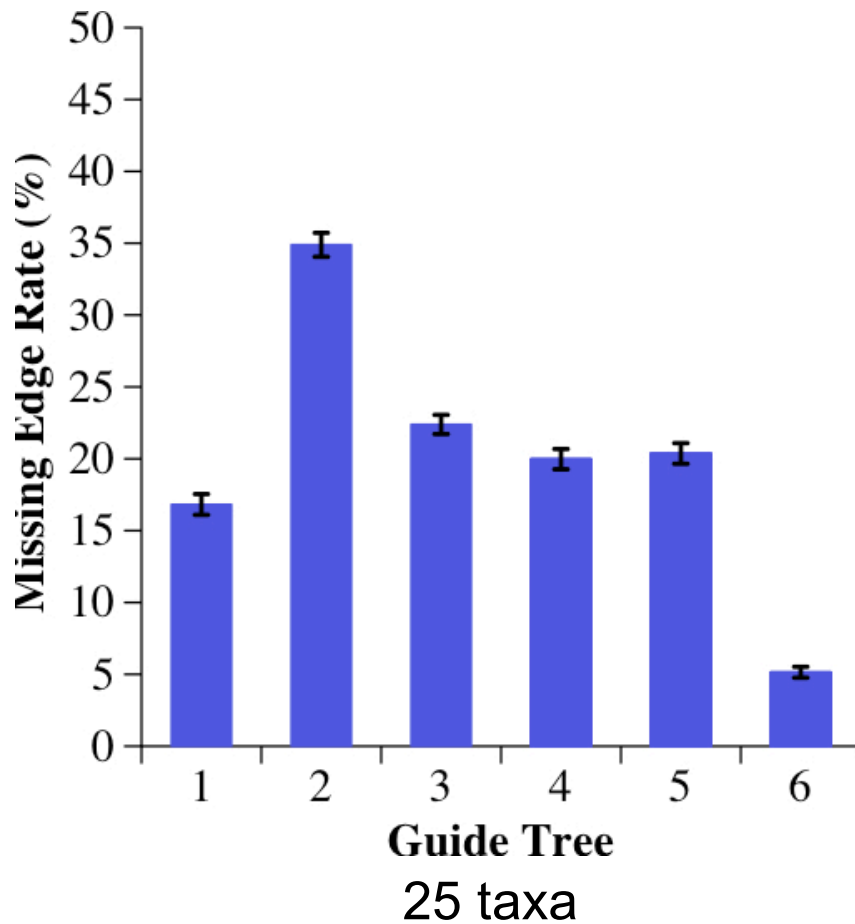
Future work

- Better models and better simulators!!! (ROSE is limited)
- Better metrics for alignment accuracy that are predictive of phylogenetic accuracy
- New data structures and visualization tools for representing homologies
- Extension of SATe-ML to models that include gap events (indels, duplications, and rearrangements)

Acknowledgements

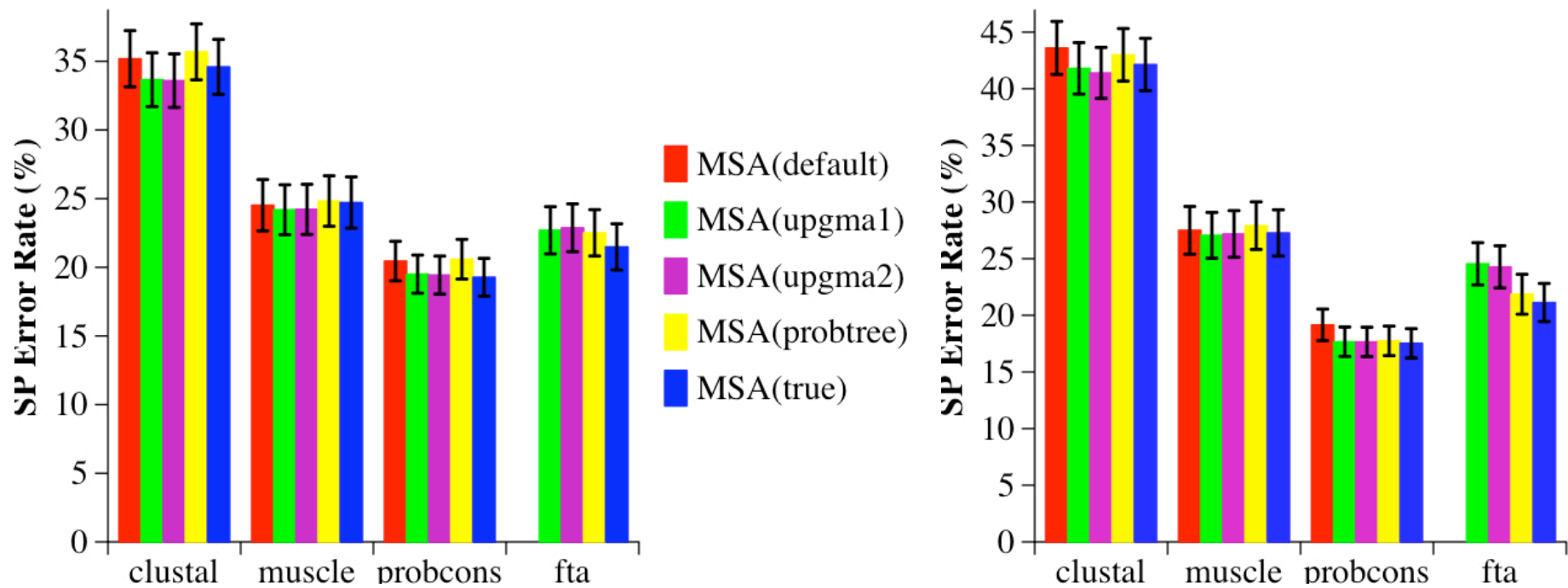
- Funding: NSF, The David and Lucile Packard Foundation, The Program in Evolutionary Dynamics at Harvard, and The Institute for Cellular and Molecular Biology at UT-Austin.
- Collaborators: Claude de Pamphilis, Peter Erdos, Daniel Huson, Jim Leebens-Mack, Randy Linder, Kevin Liu, Bernard Moret, Serita Nelesen, Usman Roshan, Mike Steel, Katherine St. John, Laszlo Szekely, Li-San Wang, Tiffani Williams, and David Zhao.
- Thanks also to Li-San Wang and Serafim Batzoglou (slides)

Guide Tree Accuracy



1-Clustal default, 2- ProbCons default, 3-Muscle default, 4-UPGMA1, 5-UPGMA2, 6-ProbTree

SP-Error Rates



Error Rates (100 Taxa)

